

WRLFMD Quarterly Report April to June 2017

Foot-and-Mouth Disease





CONTENTS

1.	Summary of samples tested and reported FMD outbreaks	3
1.1.	Asia	3
1.2.	Africa	5
1.3.	South America.....	6
1.4.	Uncharacterised FMD viruses	6
2.	Detailed Analysis.....	8
2.1.	ASIA.....	9
2.2.	AFRICA.....	22
3.	Vaccine matching	25
4.	Annex 1	26
4.1.	Summary of Submissions	26
4.2.	Clinical Samples.....	26
4.3.	Antigenic Characterisation	31
5.	Annex 2	33
6.	Annex 3	41

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



1. Summary of samples tested and reported FMD outbreaks

1.1. Asia

Cambodia

Samples were received on 23rd May 2017 from the OIE Regional Reference Laboratory (RRL) in Pakchong (Thailand). Virus isolation and typing results are pending.

Iran

Twenty four samples from cattle and sheep were received on 8th May 2017. They were collected between October 2016 and February 2017 in various locations. **FMDV type O** was isolated from nine samples and **type A** also from nine samples. VP1 sequencing showed that all nine **type O** viruses belonged to the ME-SA/PanAsia-2 lineage with eight belonging to the ANT-10 sub-lineage and one belonging to the QOM-15 sub-lineage (see below). Two of the **type A** viruses belonged to the ASIA/G-VII lineage and the other seven belonged to the Iran-05 lineage (with six belonging to the FAR-09 sublineage and one belonging to the FAR-11 sublineage) (see below); findings that indicate that multiple serotype A lineages (with different antigenic properties) are currently co-circulating in the region.

Israel

Two outbreaks of **FMD type A** were reported in cattle in the Hazafon (Northern) district on 1st May 2017 and 22nd May 2017. VP1 sequencing, performed at the Kimron Veterinary Institute, showed that the causative virus belonged to the ASIA/G-VII lineage (see below). Another outbreak, due to **FMD type O**, in cattle at Haifa was shown to be due to the EA-3 topotype and closely related to viruses present in the West Bank of the Palestinian Autonomous Territories (see below).

Mongolia

Between 2nd February and 11th April 2017, 13 outbreaks of **FMD type O** were reported in cattle, sheep and goats in four provinces (Sukhbaatar, Dornod, Khentii and Dornogovi) in the east of the country. A batch of 16 samples were received on the

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



08/05/2017 which had been collected between March 2015 and May 2017. These were typed as **FMDV type O** (two collected in 2015 and eight collected between January and April 2017) and **type A** (collected from cattle and sheep in September 2016). The **type O** viruses from 2015 belonged to SEA/Mya-98 and ME-SA/PanAsia lineages, while those from 2017 all belonged to the PanAsia lineage (see below). The **type A** viruses from 2016 belonged to the ASIA/Sea-97 lineage (see below).

Myanmar

An outbreak of **FMD type Asia 1** was reported in cattle in Rakhine State in January 2017. The last time this serotype was recorded in Myanmar was 2001. A sample received from the OIE RRL in Pakchong (Thailand) was typed as Asia 1 (although not virus could be isolated in cell cultures). VP1 sequencing of this sample, along with sequence received from the RRL, confirmed the presence of the Asia 1/G-VIII lineage showing the closest relationship to viruses from Bangladesh (see below). Additionally three samples, collected from cattle in January and April 2017 (Sagaing and Yangon regions), were typed as **FMD type O** and belonged to the ME-SA/Ind-2001d lineage (see below).

Laos

Two samples were received from the RRL on 23rd May 2017. They had been collected from cattle in January 2017 in Vientiane City and Champasak. Both were **FMDV type O** and belonged to the SEA/Mya-98 lineage (see below).

Palestinian Autonomous Territories

Four outbreaks of **FMD type O** were reported in cattle, sheep and goats in the West Bank between 1st May 2017 and 5th June 2017. VP1 sequencing, performed at the Kimron Veterinary Institute (Israel) showed the causal virus to belong to the EA-3 toptype (see below).

People's Republic of China

Two outbreaks of **FMD type O** have been reported in Guangdong province on 23rd March 2017 (pigs) and 15th May 2017 (cattle). Another outbreak of **FMD type O** was reported in cattle in the Xinjiang Autonomous region on 16th April 2017. Two outbreaks of **FMD type A** were also reported in cattle in Xinjiang on the 24th April 2017 and 28th April 2017 (these had previously been notified on 9th May within an OIE follow-up about foot-and-mouth disease serotype O). No genotyping has so far been reported.

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Thailand

Twenty eight samples, collected between August 2016 and February 2017 from various locations, were received from the RRL. **FMDV type O** was identified in 18 and **type A** in 10. VP1 sequencing revealed all the type O viruses to belong to the SEA/Mya-98 lineage and all the type A viruses to the ASIA/Sea-97 lineage (see below).

1.2. Africa

Algeria

Three further outbreaks of **FMD type A** were reported in cattle at Medea and Bordj Bou Arreridj on 2nd April 2017 and Setif on 7th April 2017. The latter outbreak was originally thought to be caused by SAT 1, but laboratory investigations confirmed that only type A was present. Genotyping at IZSLER and the WRLFMD showed the virus to belong to genotype IV of the AFRICA toptotype (see below).

Ethiopia

Two batches of samples were received, one on 11th April 2017 and the other on 15th May 2017.

The first batch were samples collected from cattle between February 2016 and March 2017 from various locations in Oromia Region and Addis Ababa. **FMD type O** viruses were isolated from 17 samples and VP1 sequencing showed that those collected in 2016 (n=13) all belonged to the EA-4 toptotype while the four collected in 2017 belonged to the EA-3 toptotype.

The second batch consisted of 54 samples (some original epithelium and other cell culture isolates) collected from cattle between January 2016 and April 2017 in the Oromia Region and Addis Ababa. **FMD type O** viruses were recovered from only 14 samples (six epithelium and eight cell culture passaged), while FMDV genome was detected in a further 19 samples and 21 were NVD. VP1 sequencing revealed all 14 viruses isolated belonged to the EA-4 toptotype (all were from 2016).

South Africa

A further outbreak of **FMD type SAT 2** was reported in cattle on 24th March 2017 at Bushbuckridge, Mpumalanga. No genotyping has yet been reported. This outbreak is located in South Africa's FMD Protection Zone, which is not part of the FMD Free Zone and therefore does not affect South Africa's OIE recognised FMD free status.

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Tunisia

Two outbreaks of **FMD type A** were reported in cattle in Bizerte on 24th April 2017. Local sequencing suggested a close relationship of the virus to the AFRICA/G-IV virus from Algeria. Analysis in one of the OIE Reference Laboratories is awaited.

Zambia

An outbreak of **FMD (untyped)** occurred in cattle in Mbala on 22nd March 2017. Further analyses are awaited.

Zimbabwe

Two outbreaks of **FMD type SAT 2** were reported in cattle on 22nd March 2017 and 17th May 2017. They occurred in Matabeleland North and Midlands provinces, respectively, and are a continuation of outbreaks which have occurred since June 2015. No genotyping has been reported.

1.3. South America

Colombia

An outbreak of FMD type O was reported in cattle at La Marota, Curipao, Tame, Arauca on 11th June 2017. This is the first FMD outbreak in Colombia since August 2009, and anywhere in the continent of South America since outbreaks in Venezuela were recognised in 2013. Genotyping results are awaited.

1.4. Uncharacterised FMD viruses

A number of outbreaks have occurred where samples have not been sent to the WRLFMD. It is probable that the countries involved have performed their own genetic characterisation; however, through the OIE/FAO laboratory network we would also like to encourage the submission of samples (or complete VP1 sequences) to the WRLFMD.

An up-to-date list and reports of FMD viruses characterised by sequencing can be found at the following website: http://www.wrlfmd.org/fmd_genotyping/2017.htm.

Results from samples or sequences received at WRLFMD (status of samples being tested) are shown in Table 1 and a complete list of clinical sample diagnostics made by the WRLFMD from April to June 2017 is shown in Annex 1 (Summary of

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Submissions). A record of all samples received by WRLFMD (April to June 2017) is shown in Annex 1 (Clinical Samples).

Table 1: Status of sequencing of samples or sequences received by the WRLFMD from April to June 2017 (* indicates samples carried over from the last quarter)

WRLFMD Batch No.	Date received	Country	Serotype	No. of samples	No. of sequences	Sequencing status
WRLFMD/2017/00006	18/04/2017	Algeria	A	3	3	Completed
WRLFMD/2017/00007	11/04/2017	Ethiopia	O	17	17	Completed
WRLFMD/2017/00008	13/04/2017	Iran	O	9	9	Completed
WRLFMD/2017/00008	13/04/2017	Iran	A	9	9	Completed
WRLFMD/2017/00009	15/05/2017	Ethiopia	O	14	14	Completed
WRLFMD/2017/00010	08/05/2017	Mongolia	O	8	8	Completed
WRLFMD/2017/00010	08/05/2017	Mongolia	A	2	2	Completed
WRLFMD/2017/00011	23/05/2017	Thailand	O	18	18	Completed
WRLFMD/2017/00011	23/05/2017	Thailand	A	10	10	Completed
WRLFMD/2017/00012	23/05/2017	Myanmar	O	3	3	Completed
WRLFMD/2017/00012	23/05/2017	Myanmar	Asia 1*	1	1	Completed
WRLFMD/2017/00013	23/05/2017	Laos	O	2	2	Completed
WRLFMD/2017/00015	23/05/2017	Cambodia	Pending	-	-	Pending
Total				96	96	

*, no virus isolated on cell culture

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



2. Detailed Analysis

Key for maps and trees:

Serotype O
Serotype A
Serotype C
Serotype Asia-1
Serotype SAT 1
Serotype SAT 2
Serotype SAT 3
FMDV Genome Detected
No Virus Detected

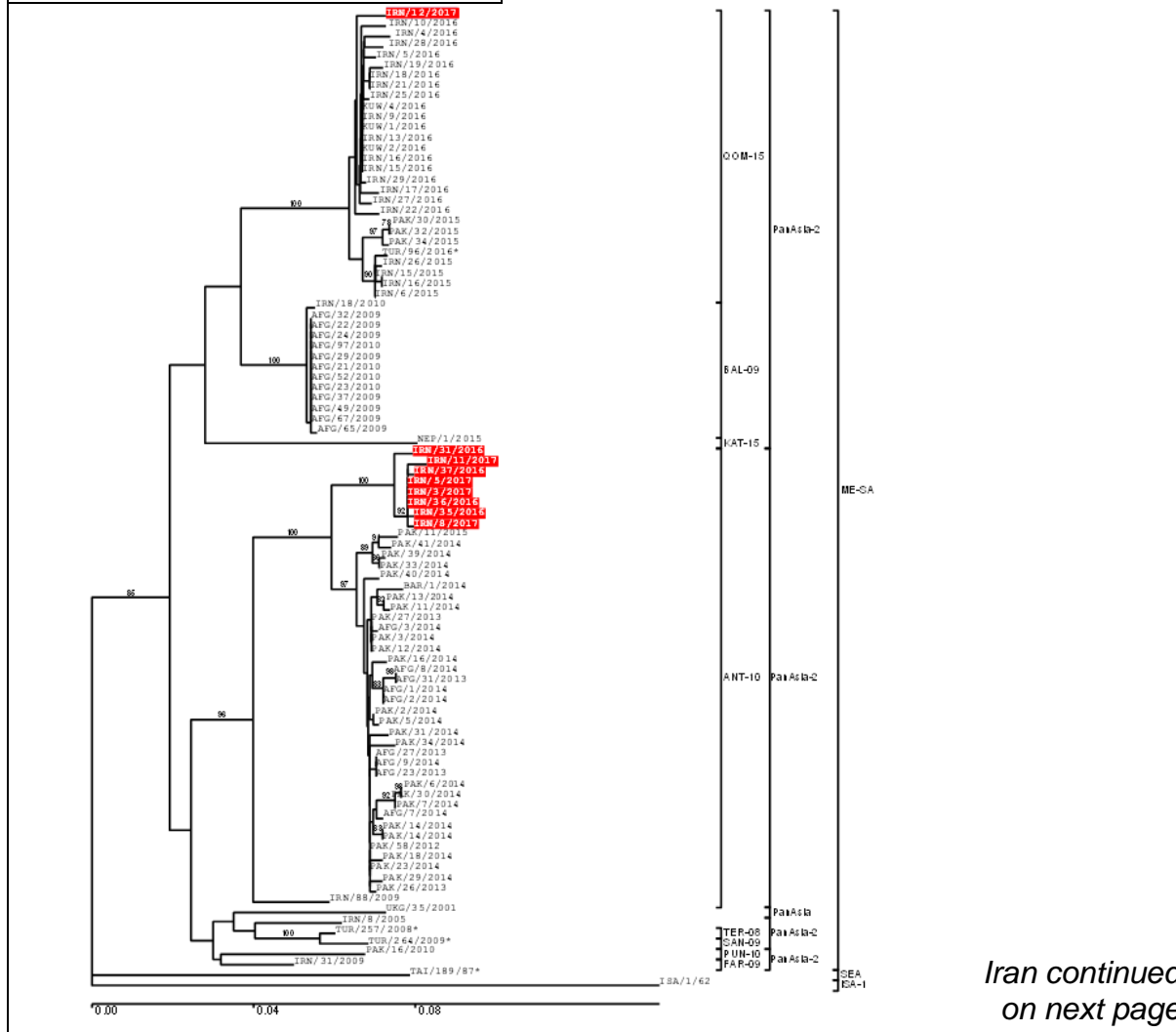
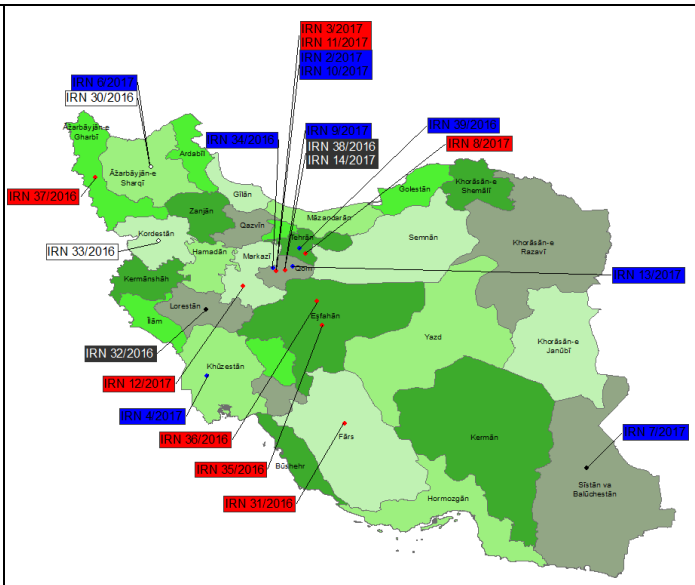
The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



2.1. ASIA

Iran
Batch: WRLFMD/2017/00008
Date received: 13/04/2017
No. of samples: 24
O (ME-SA/PanAsia-2/ANT-10): 8
O (ME-SA/PanAsia-2/QOM-15): 1
A (ASIA/G-VII): 2
A (ASIA/Iran-05/FAR-09): 6
A (ASIA/Iran-05/FAR-11): 1
FMDV-GD: 4
NVD: 2



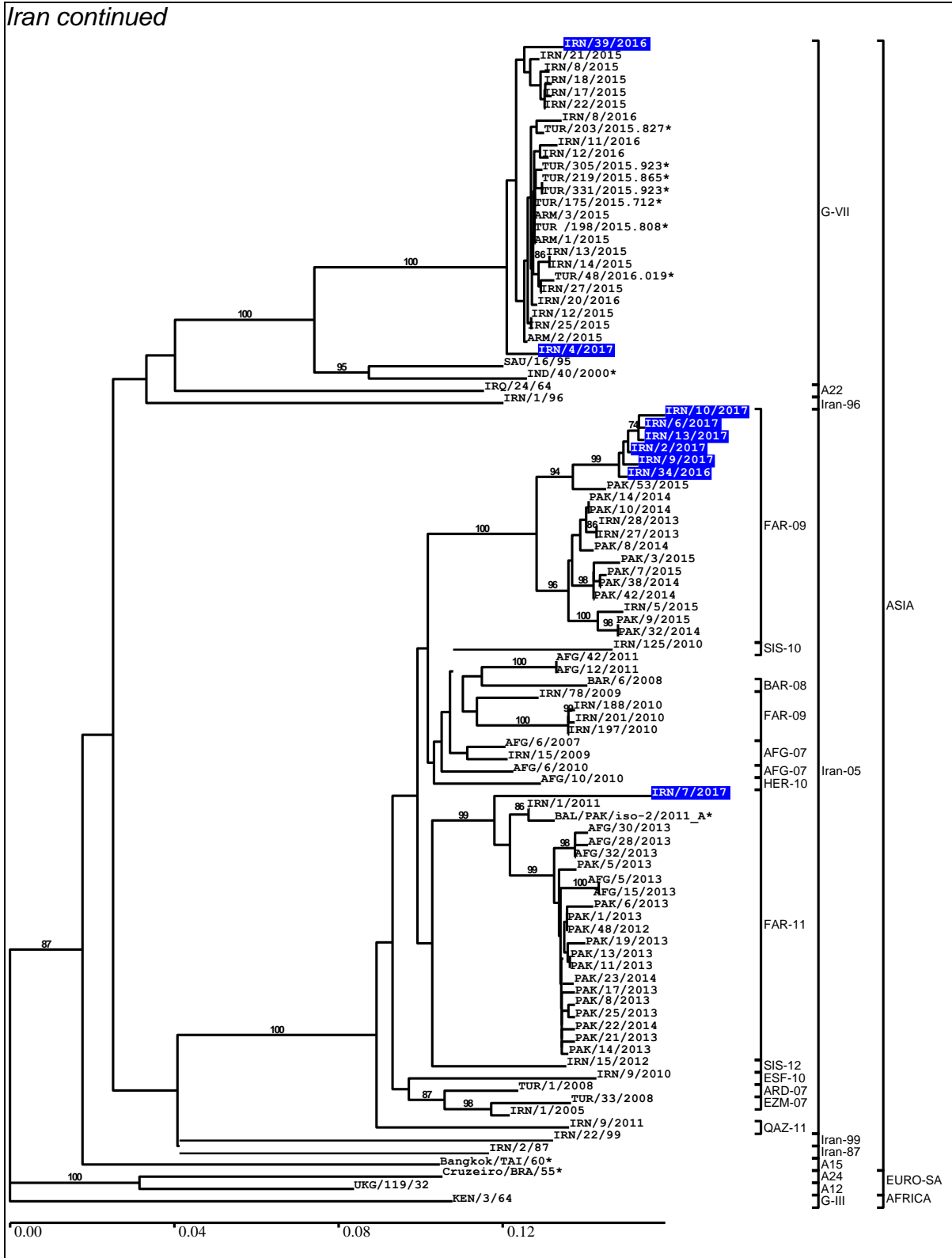
Iran continued on next page

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk

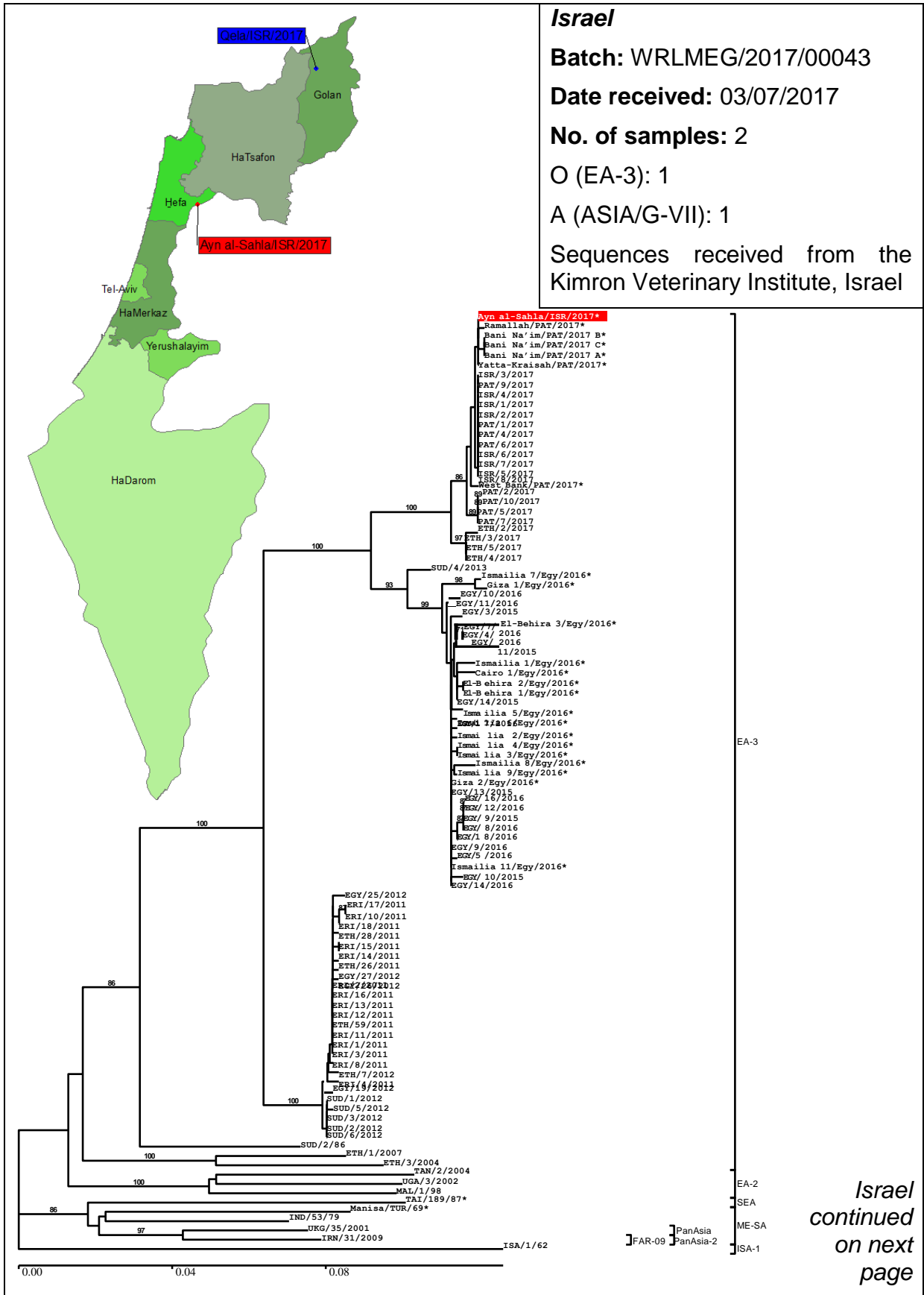


Iran continued



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk

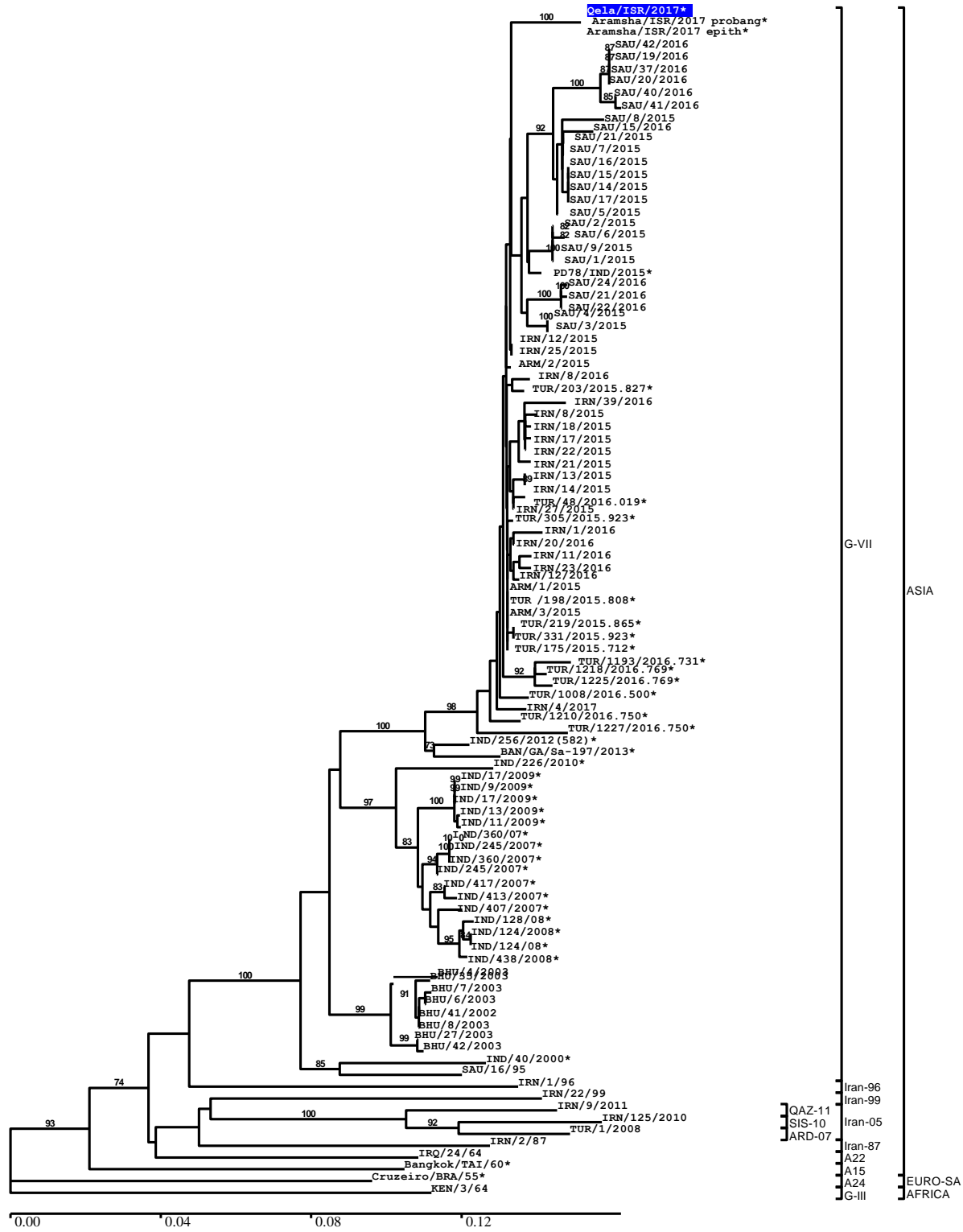


The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Israel continued



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



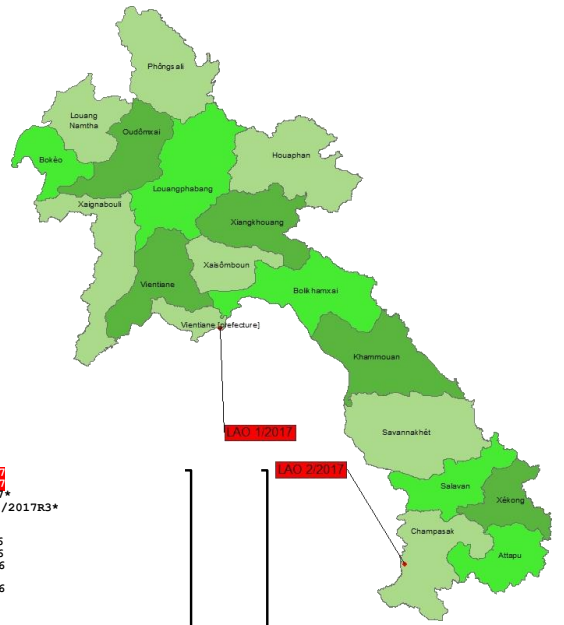
Laos

Batch: WRLFMD/2017/00013

Date received: 23/05/2017

No. of samples: 2

O (SEA/Mya-98): 2



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Mongolia

Batch: WRLFMD/2017/00010

Date received: 08/05/2017

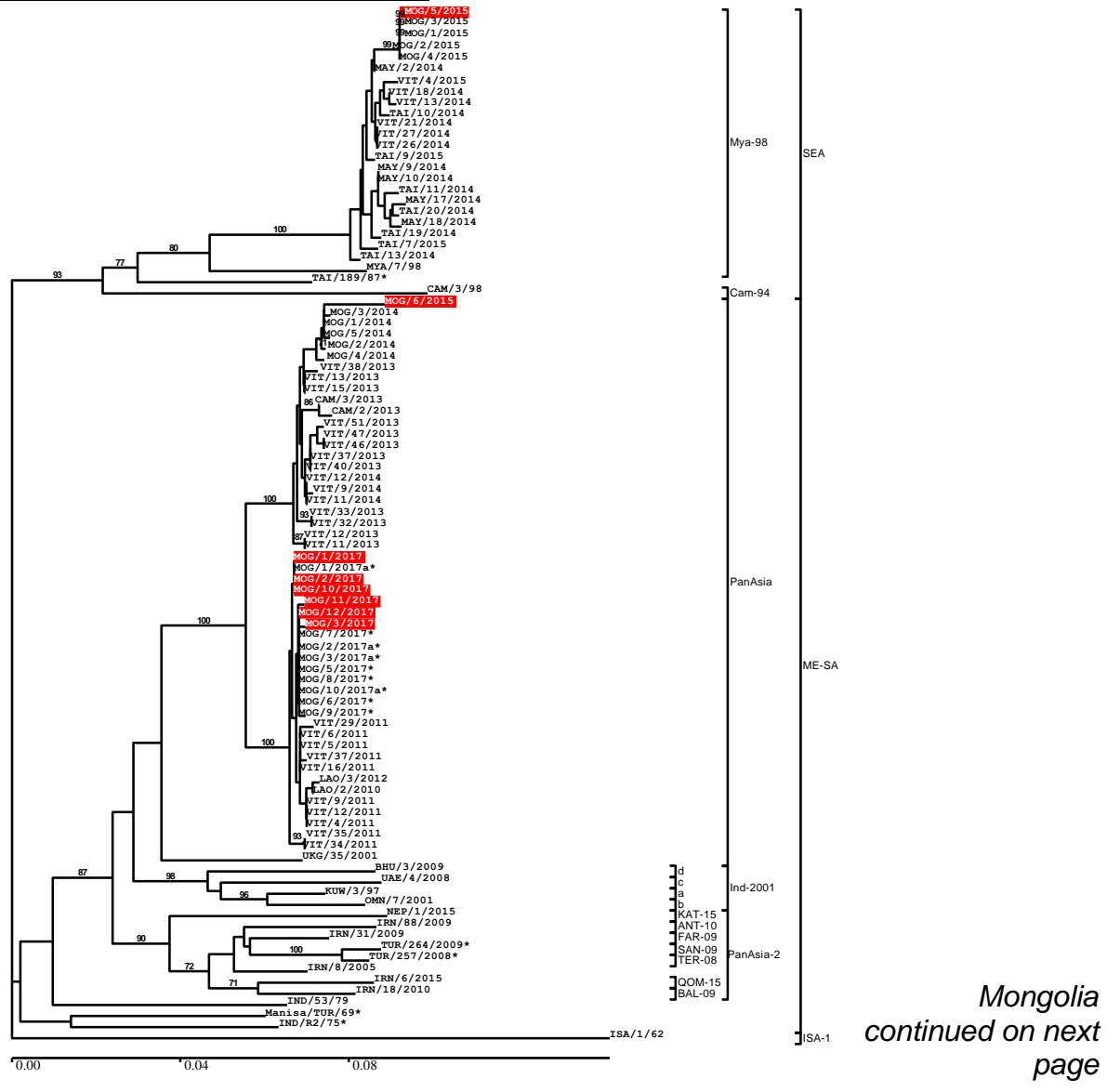
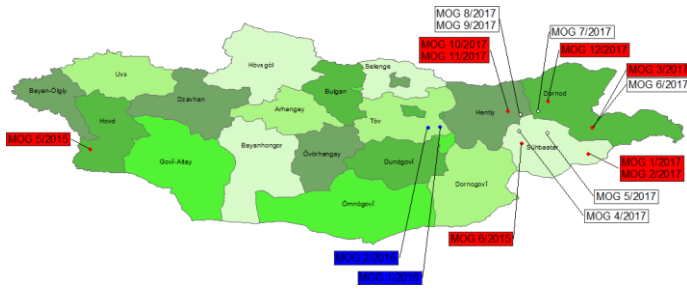
No. of samples: 16

O (ME-SA/PanAsia): 7

O (SEA/Mya-98): 1

A (ASIA/Sea-97): 2

NVD: 6



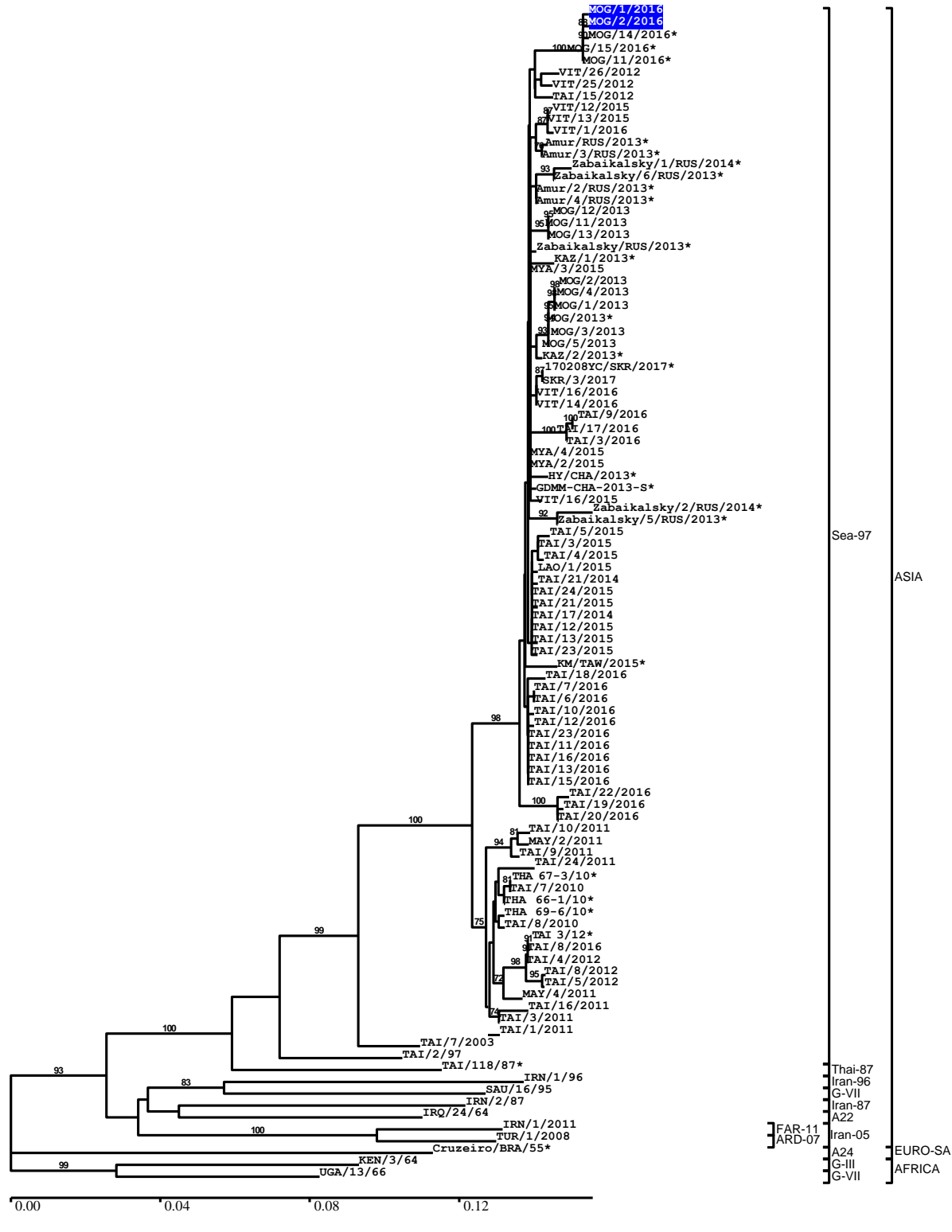
*Mongolia
continued on next
page*

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Mongolia continued



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Myanmar

Batch: WRLFMD/2017/00012

Date received: 23/05/2017

No. of samples: 5

O (ME-SA/Ind-2001d): 3

Asia 1 (ASIA/G-VIII): 1

FMDV-GD: 1

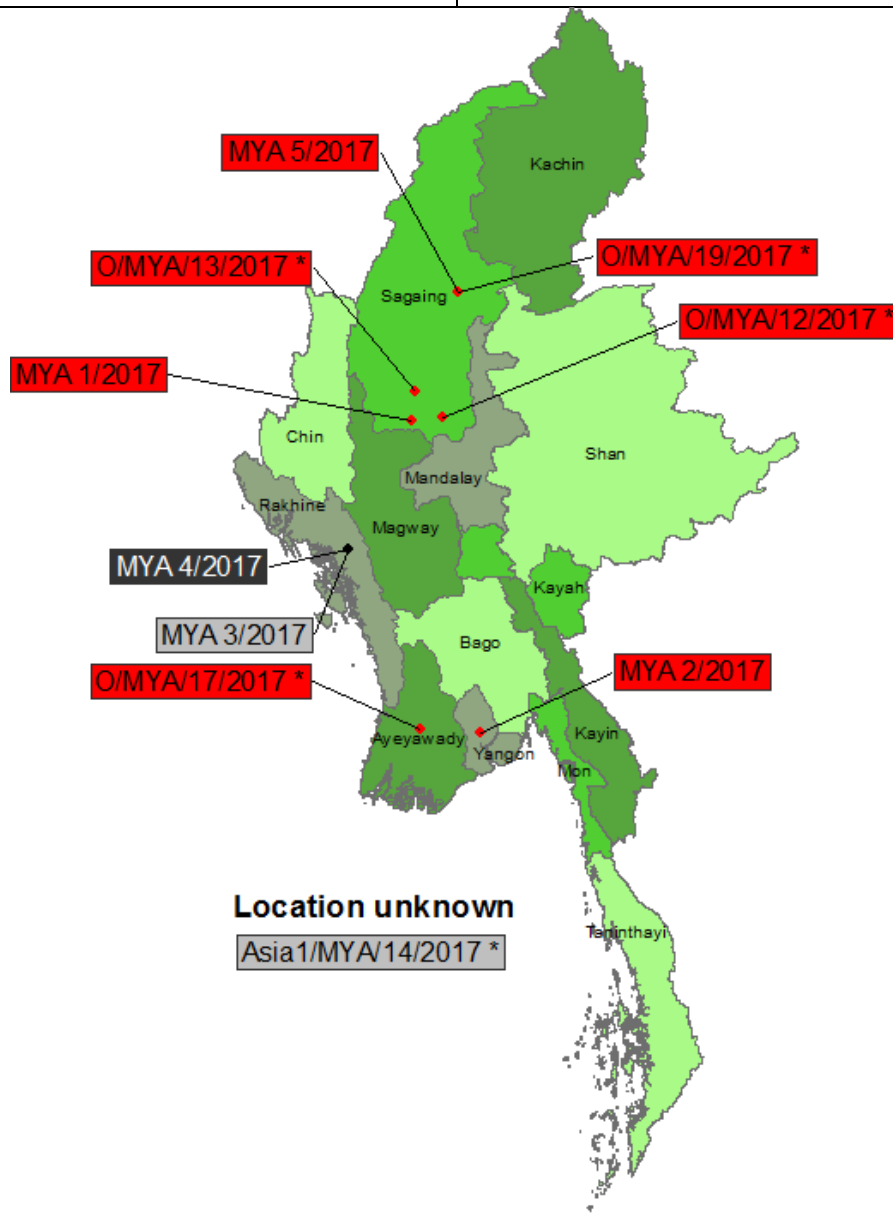
Batch: WRLMEG/2017/00034

Date received: 24/04/2017

No. of sequence: 5

O (ME-SA/Ind-2001d): 4

Asia 1 (ASIA/G-VIII): 1



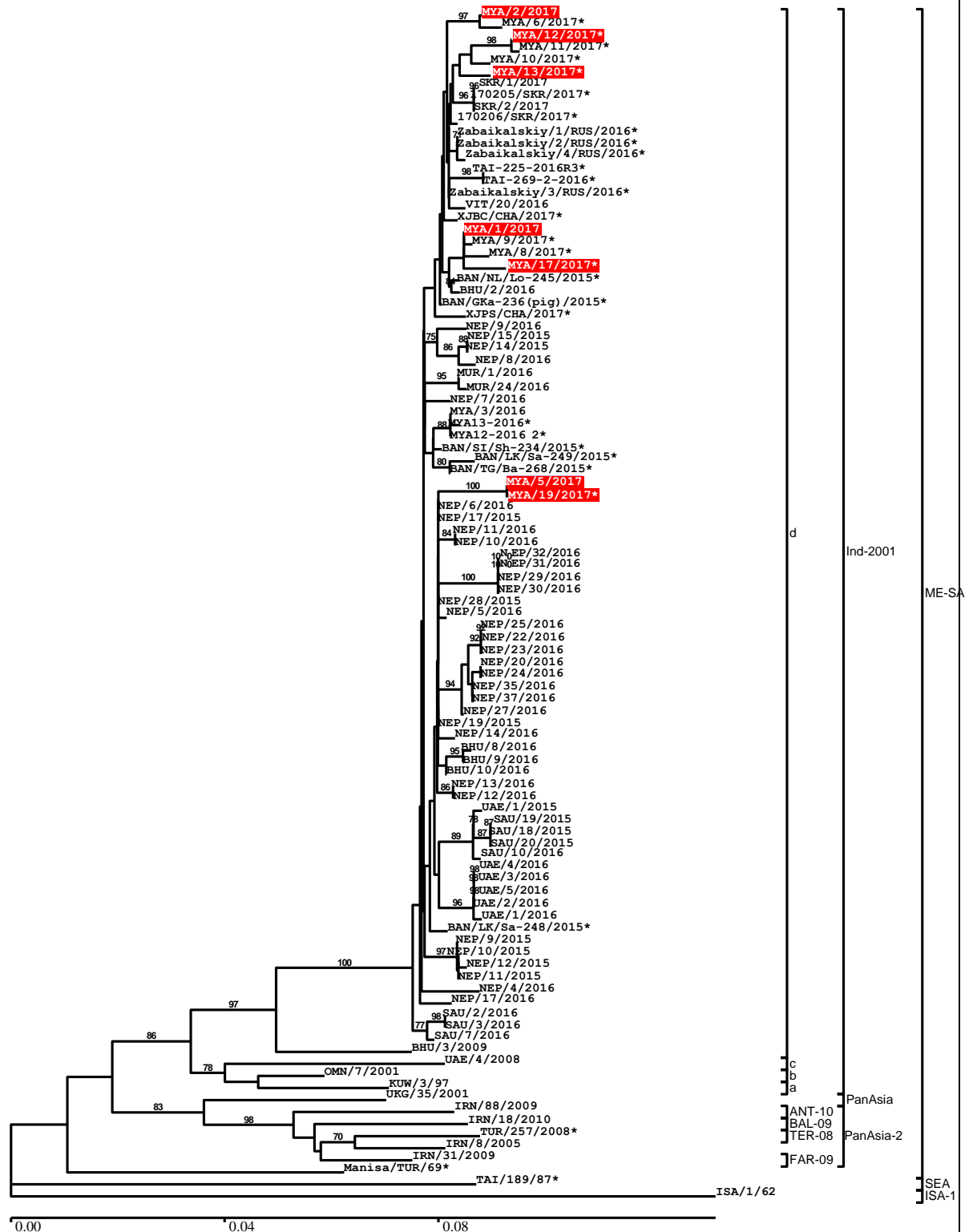
Myanmar continued on next page

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Myanmar continued



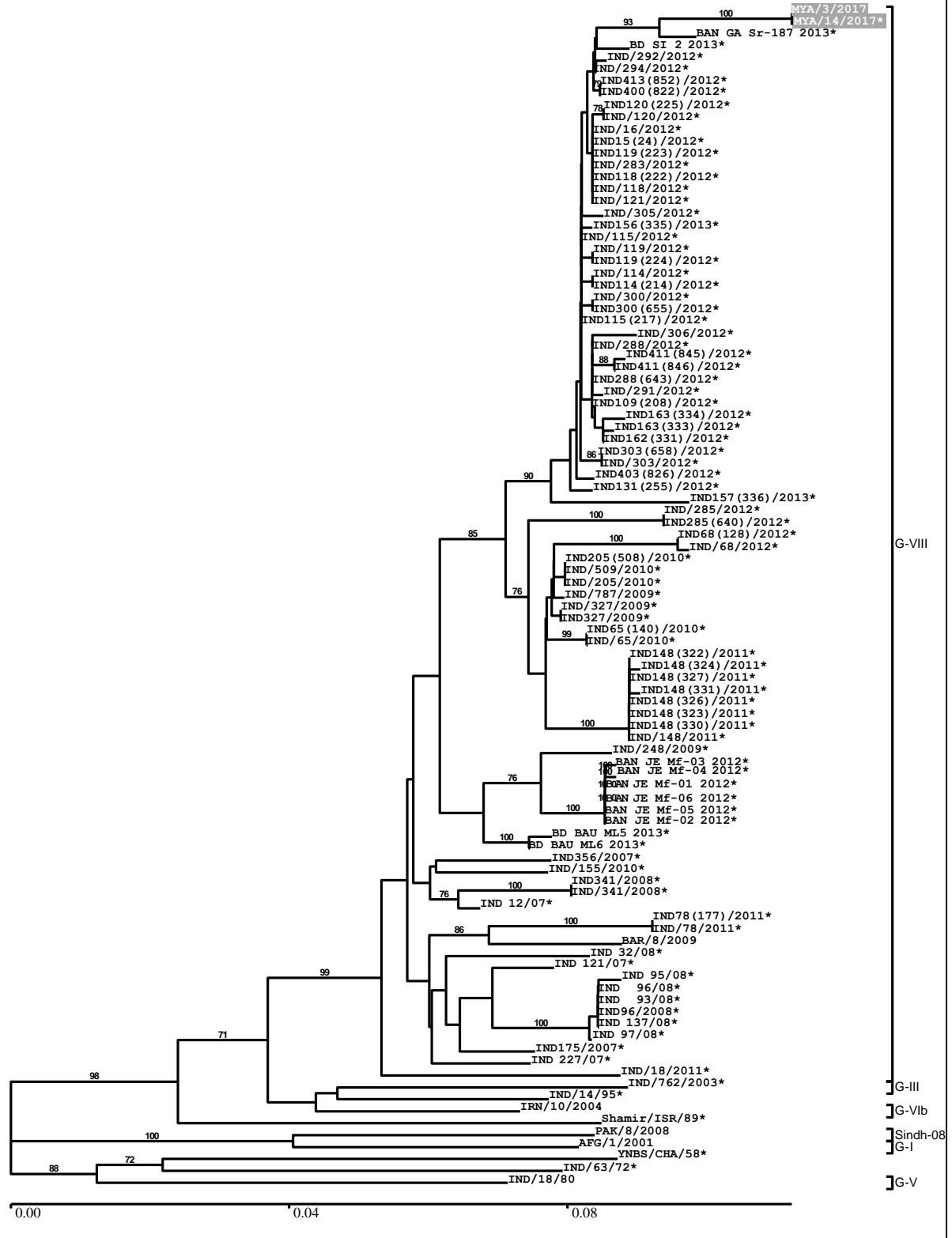
Myanmar continued on next page

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk

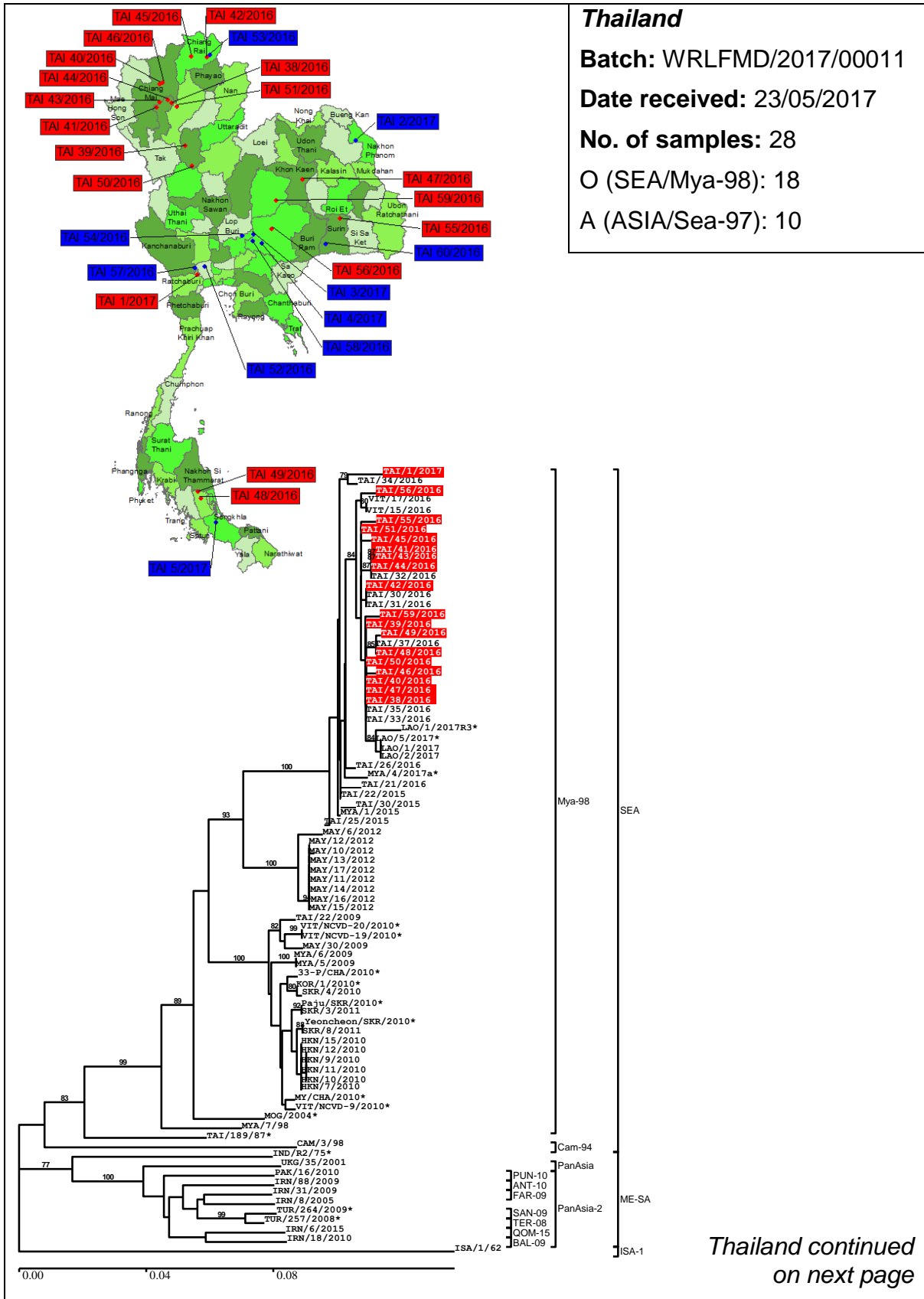


Myanmar continued



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk

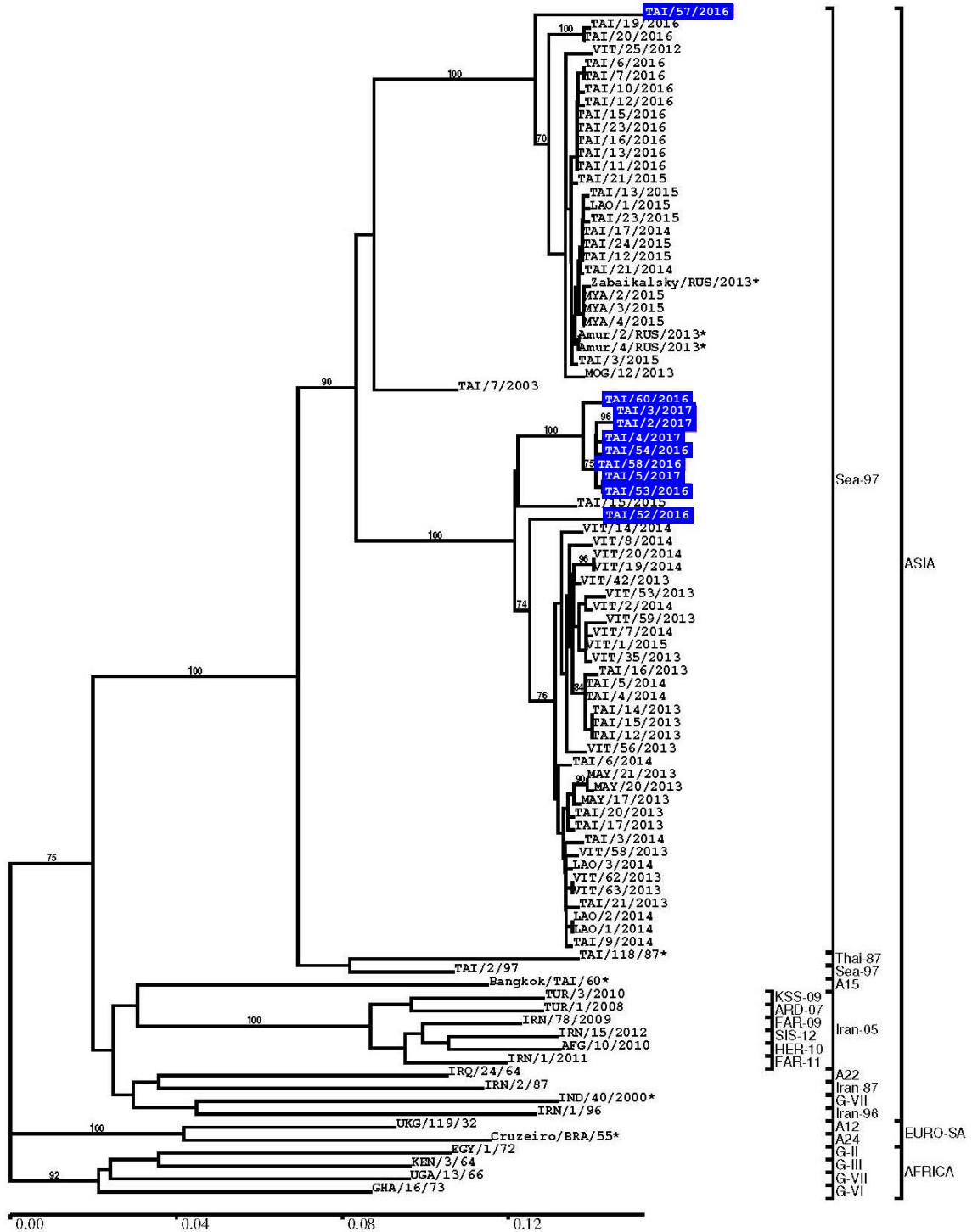


The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Thailand continued



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



2.2. AFRICA

Algeria

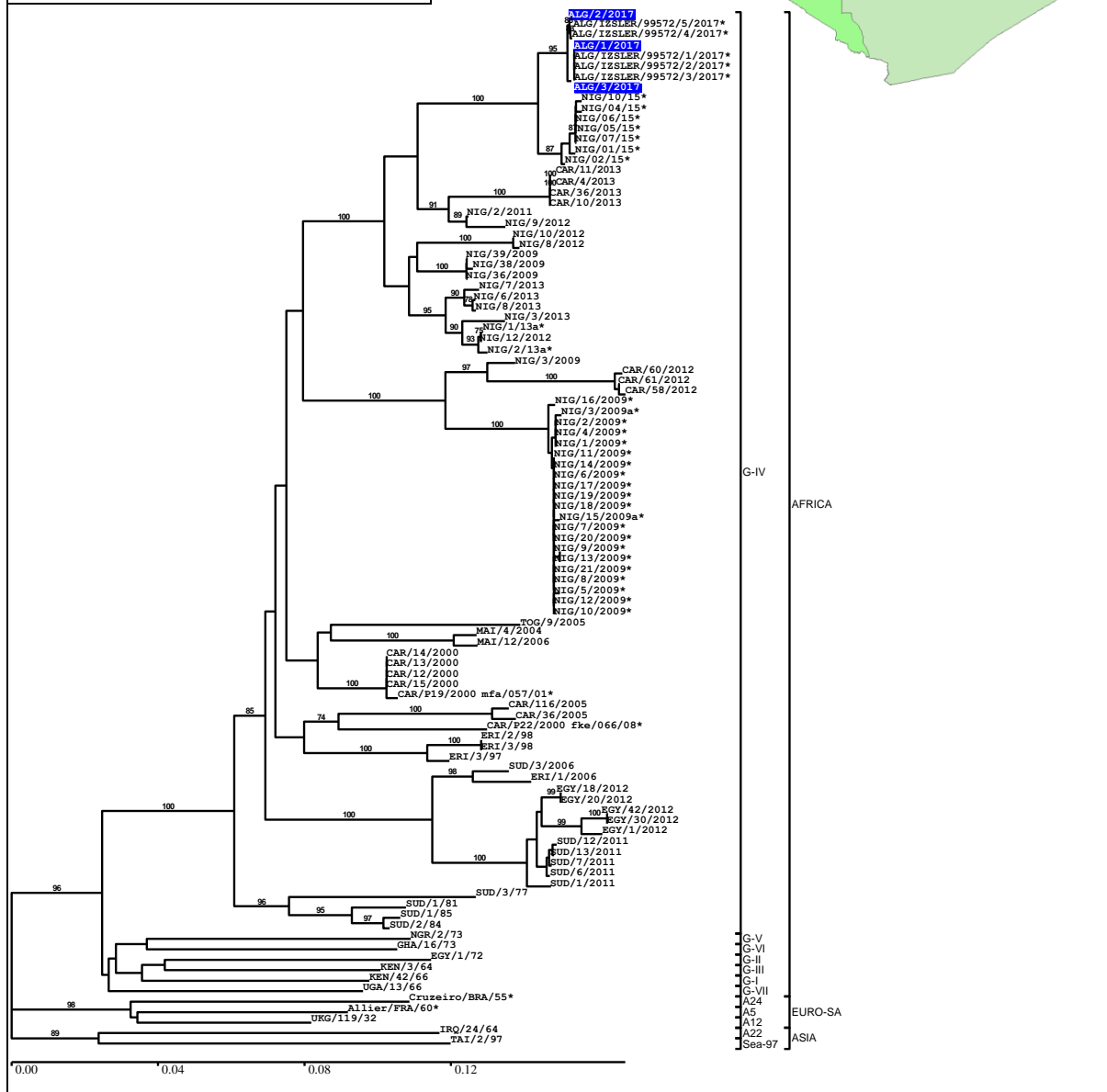
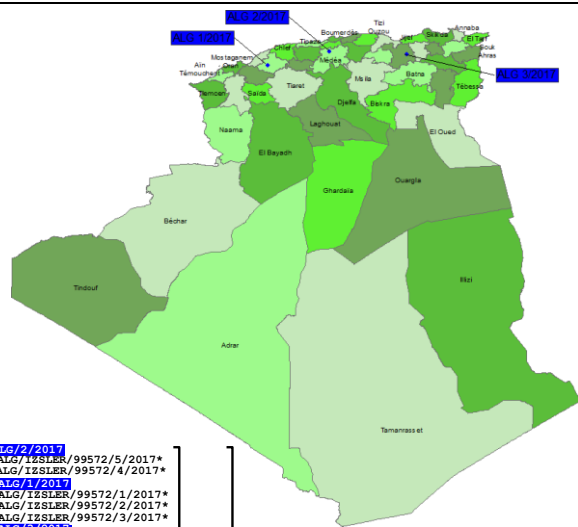
Batch: WRLFMD/2017/00006

Date received: 18/04/2017

No. of samples: 3

A (AFRICA/G-IV): 3

* 5 sequences from the same locations as ALG 1/2017 and ALG 2/2017 for analysis were also received



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Ethiopia

Batch: WRLFMD/2017/00007

Date received: 11/04/2017

No. of samples: 27

O (EA-3): 4

O (EA-4): 13

FMDV-GD: 3

NVD: 7

Batch: WRLFMD/2017/00009

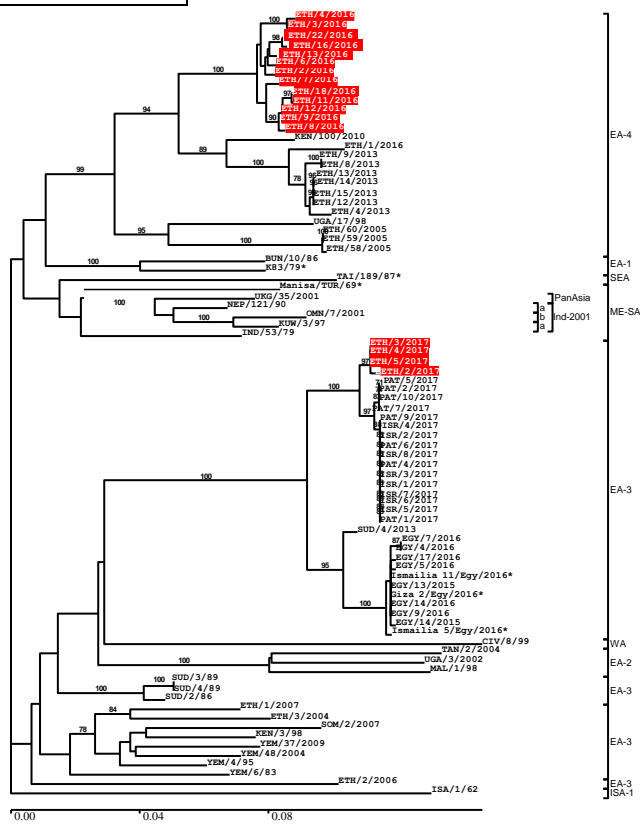
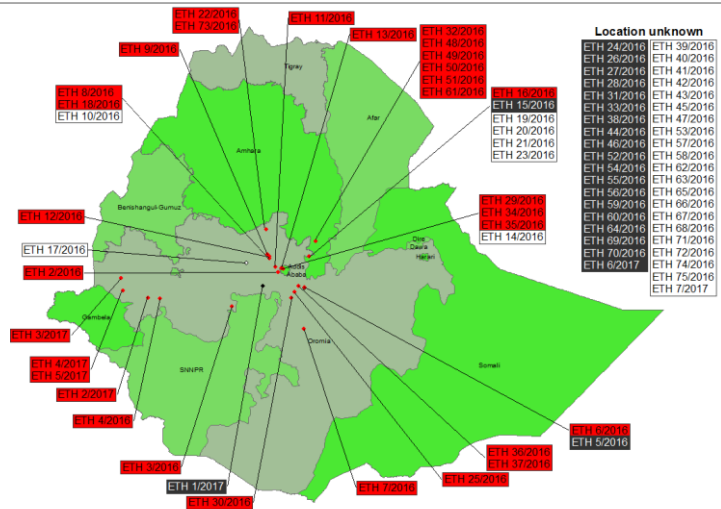
Date received: 15/05/2017

No. of samples: 54

O (EA-4): 14

FMDV-GD: 19

NVD: 21

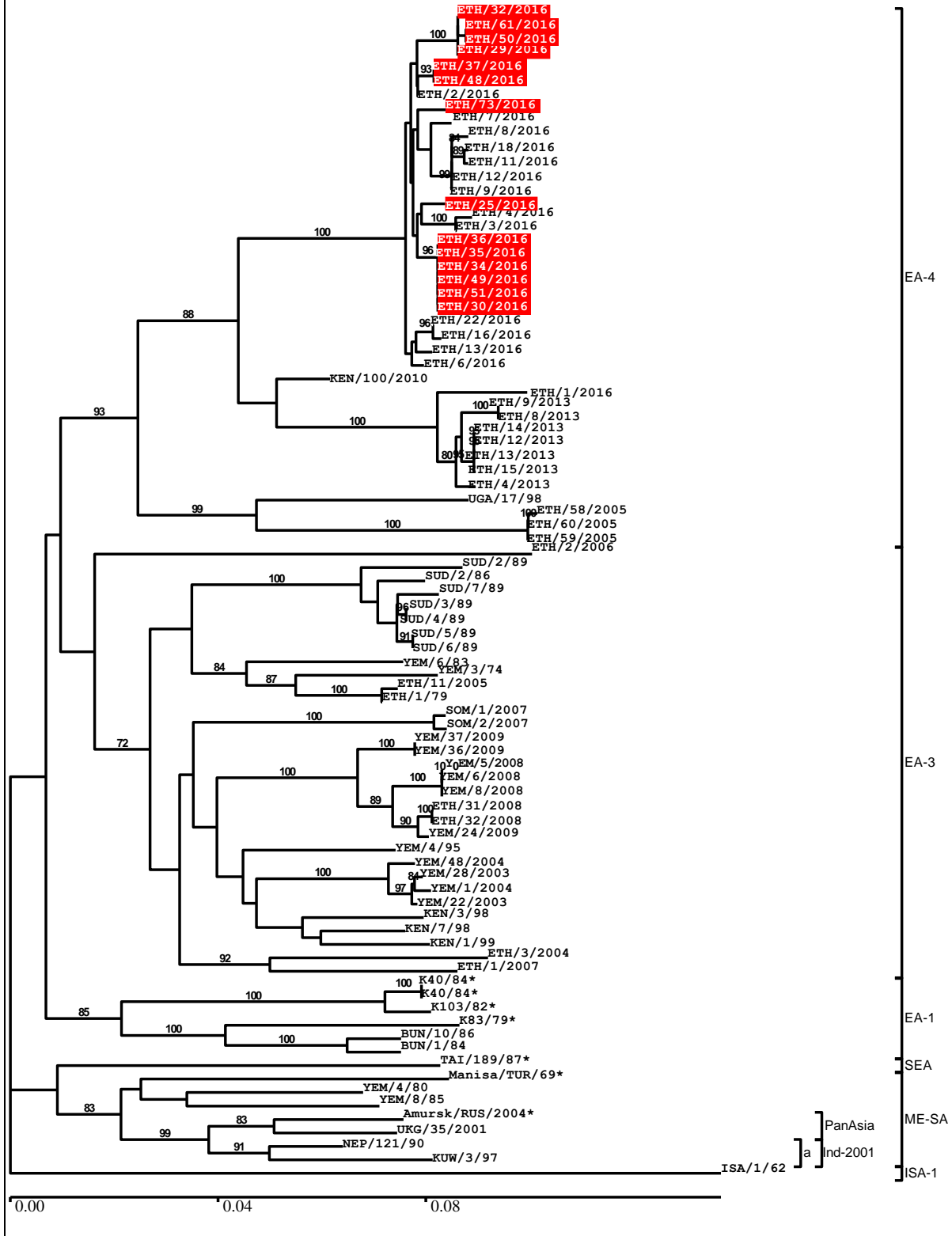


The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Ethiopia continued



The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



3. Vaccine matching

During this reporting period vaccine matching has been undertaken for 4 FMD virus field strains: serotype O (n=2) and serotype A (n=2). These are samples from Ethiopia and Iran.

For individual data see Annex 1, section 4.3 (Antigenic Characterisation).

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



4. Annex 1

4.1. Summary of Submissions

Table 2: Summary of samples collected and received to WRLFMD (April to June 2017)

Country	Nº of samples	Virus isolation in cell culture/ELISA								RT-PCR for FMD (or SVD)	
		FMD virus serotypes								virus (where appropriate)	
		O	A	C	SAT 1	SAT 2	SAT 3	ASIA -1	No Virus Detected	Positive	Negative
ALGERIA	3	-	3	-	-	-	-	-	-	3	-
CAMBODIA	4	-	-	-	-	-	-	-	-	-	-
ETHIOPIA	81	31	-	-	-	-	-	-	50	52	29
IRAN	24	9	9	-	-	-	-	-	6	22	2
LAOS	2	-	-	-	-	-	-	-	2	2	-
MONGOLIA	16	8	2	-	-	-	-	-	6	10	6
MYANMAR	5	3	-	-	-	-	-	-	2	5	-
THAILAND	28	18	10	-	-	-	-	-	-	28	-
TOTAL	163	69	24	-	-	-	-	-	66	122	37

Abbreviations used in table

VI / ELISA	FMD (or SVD) virus serotype identified following virus isolation in cell culture and antigen detection ELISA
FMD	Foot-and-mouth disease
SVD	Swine vesicular disease
NVD	No FMD, SVD or vesicular stomatitis virus detected
NT	Not tested
rRT-PCR	Real-time reverse transcription polymerase chain reaction for FMD (or SVD) viral genome

4.2. Clinical Samples

Table 3: Clinical sample diagnostics made by the WRLFMD® April to June 2017

Country	WRL for FMD Sample Identification	Animal	Date of Collection	Results		Final report
				VI/ELISA	RT-PCR	

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Country	WRL for FMD Sample Identification	Animal	Date of Collection	Results		
				VI/ELISA	RT-PCR	Final report
ALGERIA	ALG 1/2017	CATTLE	09-Apr-17	A	POS	A
	ALG 2/2017	CATTLE	09-Apr-17	A	POS	A
	ALG 3/2017	CATTLE	11-Apr-17	A	POS	A
CAMBODIA	CAM 1/2016	Awaiting information from sender	02-Dec-16	Pending	Pending	Pending
	CAM 2/2016	Awaiting information from sender	02-Dec-16	Pending	Pending	Pending
	CAM 3/2016	Awaiting information from sender	02-Dec-16	Pending	Pending	Pending
	CAM 4/2016	Awaiting information from sender	02-Dec-16	Pending	Pending	Pending
ETHIOPIA	ETH 2/2016	CATTLE	10-Feb-16	O	POS	O
	ETH 3/2016	CATTLE	23-Feb-16	O	NEG	O
	ETH 4/2016	CATTLE	04-Aug-16	O	POS	O
	ETH 5/2016	CATTLE	05-Aug-16	NEG	POS	FMDV GD
	ETH 6/2016	CATTLE	03-Sep-16	O	POS	O
	ETH 7/2016	CATTLE	05-Sep-16	O	POS	O
	ETH 8/2016	CATTLE	31-Oct-16	O	POS	O
	ETH 9/2016	CATTLE	31-Oct-16	NEG	NEG	NVD
	ETH 10/2016	CATTLE	31-Oct-16	O	POS	O
	ETH 11/2016	CATTLE	11-Nov-16	O	POS	O
	ETH 12/2016	CATTLE	11-Nov-16	O	POS	O
	ETH 13/2016	CATTLE	18-Nov-16	NEG	NEG	NVD
	ETH 14/2016	CATTLE	18-Nov-16	NEG	POS	FMDV GD
	ETH 15/2016	CATTLE	20-Nov-16	O	POS	O
	ETH 16/2016	CATTLE	27-Nov-16	NEG	NEG	NVD
	ETH 17/2016	CATTLE	29-Nov-16	O	POS	O
	ETH 18/2016	CATTLE	31-Nov-16	NEG	NEG	NVD
	ETH 19/2016	CATTLE	18-Dec-16	NEG	NEG	NVD
	ETH 20/2016	CATTLE	18-Dec-16	NEG	NEG	NVD
	ETH 21/2016	CATTLE	18-Dec-16	O	POS	O
	ETH 22/2016	CATTLE	27-Dec-16	NEG	NEG	NVD
	ETH 23/2016	CATTLE	31-Dec-16	NEG	POS	FMDV GD
	ETH 24/2016	BOVINE	03-Jan-16	NEG	POS	FMDV GD
	ETH 25/2016	BOVINE	03-Jan-16	O	POS	O
	ETH 26/2016	BOVINE	27-Jan-16	NEG	POS	FMDV GD
	ETH 27/2016	BOVINE	27-Jan-16	NEG	POS	FMDV GD
	ETH 28/2016	BOVINE	03-Mar-16	NEG	POS	FMDV GD
	ETH 29/2016	BOVINE	03-Mar-16	O	POS	O
	ETH 30/2016	BOVINE	16-Mar-16	O	POS	O
	ETH 31/2016	BOVINE	16-Mar-16	NEG	POS	FMDV GD
	ETH 32/2016	BOVINE	12-Apr-16	O	POS	O
	ETH 33/2016	BOVINE	12-Apr-16	NEG	POS	FMDV GD
	ETH 34/2016	BOVINE	27-Jun-16	O	POS	O
	ETH 35/2016	BOVINE	27-Jun-16	O	POS	O
	ETH 36/2016	BOVINE	08-Aug-16	O	POS	O
	ETH 37/2016	BOVINE	08-Aug-16	O	POS	O
	ETH 38/2016	BOVINE	11-Aug-16	NEG	POS	FMDV GD
	ETH 39/2016	BOVINE	11-Aug-16	NEG	NEG	NVD
	ETH 40/2016	BOVINE	12-Sep-16	NEG	NEG	NVD
	ETH 41/2016	BOVINE	12-Sep-16	NEG	NEG	NVD
	ETH 42/2016	BOVINE	02-Oct-16	NEG	NEG	NVD
	ETH 43/2016	BOVINE	02-Oct-16	NEG	NEG	NVD
	ETH 44/2016	BOVINE	12-Oct-16	NEG	POS	FMDV GD

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Country	WRL for FMD Sample Identification	Animal	Date of Collection	Results		Final report
				VI/ELISA	RT-PCR	
	ETH 45/2016	BOVINE	12-Oct-16	NEG	NEG	NVD
	ETH 46/2016	BOVINE	12-Oct-16	NEG	POS	FMDV GD
	ETH 47/2016	BOVINE	12-Oct-16	NEG	NEG	NVD
	ETH 48/2016	BOVINE	13-Oct-16	O	POS	O
	ETH 49/2016	BOVINE	13-Oct-16	O	POS	O
	ETH 50/2016	BOVINE	21-Oct-16	O	POS	O
	ETH 51/2016	BOVINE	21-Oct-16	O	POS	O
	ETH 52/2016	BOVINE	28-Oct-16	NEG	POS	FMDV GD
	ETH 53/2016	BOVINE	28-Oct-16	NEG	NEG	NVD
	ETH 54/2016	BOVINE	28-Oct-16	NEG	POS	FMDV GD
	ETH 55/2016	BOVINE	28-Oct-16	NEG	POS	FMDV GD
	ETH 56/2016	BOVINE	11-Nov-16	NEG	POS	FMDV GD
	ETH 57/2016	BOVINE	11-Nov-16	NEG	NEG	NVD
	ETH 58/2016	BOVINE	11-Nov-16	NEG	NEG	NVD
	ETH 59/2016	BOVINE	11-Nov-16	NEG	POS	FMDV GD
	ETH 60/2016	BOVINE	23-Nov-16	NEG	POS	FMDV GD
	ETH 61/2016	BOVINE	23-Nov-16	O	POS	O
	ETH 62/2016	BOVINE	13-Dec-16	NEG	NEG	NVD
	ETH 63/2016	BOVINE	13-Dec-16	NEG	NEG	NVD
	ETH 64/2016	BOVINE	18-Dec-16	NEG	POS	FMDV GD
	ETH 65/2016	BOVINE	18-Dec-16	NEG	NEG	NVD
	ETH 66/2016	BOVINE	19-Dec-16	NEG	NEG	NVD
	ETH 67/2016	BOVINE	19-Dec-16	NEG	NEG	NVD
	ETH 68/2016	BOVINE	19-Dec-16	NEG	NEG	NVD
	ETH 69/2016	BOVINE	19-Dec-16	NEG	POS	FMDV GD
	ETH 70/2016	BOVINE	20-Dec-16	NEG	POS	FMDV GD
	ETH 71/2016	BOVINE	20-Dec-16	NEG	NEG	NVD
	ETH 72/2016	BOVINE	20-Dec-16	NEG	NEG	NVD
	ETH 73/2016	BOVINE	20-Dec-16	O	POS	O
	ETH 74/2016	BOVINE	27-Dec-16	NEG	NEG	NVD
	ETH 75/2016	BOVINE	27-Dec-16	NEG	NEG	NVD
	ETH 1/2017	CATTLE	18-Mar-17	O	POS	O
	ETH 2/2017	CATTLE	18-Mar-17	O	POS	O
	ETH 3/2017	CATTLE	18-Mar-17	O	POS	O
	ETH 4/2017	CATTLE	18-Mar-17	O	POS	O
	ETH 5/2017	CATTLE	18-Mar-17	O	POS	O
	ETH 6/2017	BOVINE	01-Apr-17	NEG	POS	FMDV GD
	ETH 7/2017	BOVINE	01-Apr-17	NEG	NEG	NVD
	IRN 30/2016	SHEEP	21-Oct-16	NEG	NEG	NVD
	IRN 31/2016	CATTLE	14-Nov-16	O	POS	O
	IRN 32/2016	CATTLE	28-Nov-16	NEG	POS	FMDV GD
	IRN 33/2016	SHEEP	30-Nov-16	NEG	NEG	NVD
	IRN 34/2016	CATTLE	06-Dec-16	A	POS	A
	IRN 35/2016	CATTLE	14-Dec-16	O	POS	O
	IRN 36/2016	SHEEP	22-Dec-16	O	POS	O
IRAN	IRN 37/2016	CATTLE	24-Dec-16	O	POS	O
	IRN 38/2016	SHEEP	26-Dec-16	NEG	POS	FMDV GD
	IRN 39/2016	CATTLE	30-Dec-16	A	POS	A
	IRN 1/2017	CATTLE	01-Jan-17	NEG	POS	FMDV GD
	IRN 2/2017	CATTLE	07-Jan-17	A	POS	A
	IRN 3/2017	SHEEP	07-Jan-17	O	POS	O
	IRN 4/2017	CATTLE	11-Jan-17	A	POS	A
	IRN 5/2017	CATTLE	12-Jan-17	O	POS	O

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Country	WRL for FMD Sample Identification	Animal	Date of Collection	Results		Final report	
				VI/ELISA	RT-PCR		
	IRN 6/2017	SHEEP	25-Jan-17	A	POS	A	
	IRN 7/2017	CATTLE	31-Jan-17	A	POS	A	
	IRN 8/2017	CATTLE	02-Feb-17	O	POS	O	
	IRN 9/2017	CATTLE	15-Feb-17	A	POS	A	
	IRN 10/2017	CATTLE	20-Feb-17	A	POS	A	
	IRN 11/2017	CATTLE	20-Feb-17	O	POS	O	
	IRN 12/2017	CATTLE	23-Feb-17	O	POS	O	
	IRN 13/2017	SHEEP	25-Feb-17	A	POS	A	
	IRN 14/2017	CATTLE	25-Feb-17	NEG	POS	FMDV GD	
LAOS	LAO 1/2017	CATTLE	30-Jan-17	NEG	POS	FMDV GD	
	LAO 2/2017	CATTLE	30-Jan-17	NEG	POS	FMDV GD	
	MOG 5/2015	CATTLE	03-Mar-15	O	POS	O	
	MOG 6/2015	CATTLE	12-May-15	O	POS	O	
	MOG 1/2016	CATTLE	29-Aug-16	A	POS	A	
	MOG 2/2016	SHEEP	20-Sep-16	A	POS	A	
	MOG 1/2017	CATTLE	28-Jan-17	O	POS	O	
	MOG 2/2017	CATTLE	28-Jan-17	O	POS	O	
	MOG 3/2017	CATTLE	31-Jan-17	O	POS	O	
MONGOLIA	MOG 4/2017	CATTLE	09-Feb-17	NEG	NEG	NVD	
	MOG 5/2017	CATTLE	20-Feb-17	NEG	NEG	NVD	
	MOG 6/2017	CATTLE	20-Feb-17	NEG	NEG	NVD	
	MOG 7/2017	CATTLE	20-Feb-17	NEG	NEG	NVD	
	MOG 8/2017	CATTLE	23-Feb-17	NEG	NEG	NVD	
	MOG 9/2017	CATTLE	23-Feb-17	NEG	NEG	NVD	
	MOG 10/2017	CATTLE	23-Mar-17	O	POS	O	
	MOG 11/2017	CATTLE	23-Mar-17	O	POS	O	
	MOG 12/2017	CATTLE	03-Apr-17	O	POS	O	
	MYANMAR	MYA 1/2017	CATTLE	27-Jan-17	O	POS	O
		MYA 2/2017	CATTLE	27-Jan-17	O	POS	O
		MYA 3/2017	CATTLE	08-Apr-17	NEG	POS	FMDV GD
MYA 4/2017		CATTLE	08-Apr-17	NEG	POS	FMDV GD	
MYA 5/2017		CATTLE	08-Apr-17	O	POS	O	
	TAI 38/2016	CATTLE	03-Aug-16	O	POS	O	
	TAI 39/2016	CATTLE	03-Aug-16	O	POS	O	
	TAI 40/2016	CATTLE	09-Aug-16	O	POS	O	
	TAI 41/2016	CATTLE	09-Aug-16	O	POS	O	
	TAI 42/2016	CATTLE	09-Aug-16	O	POS	O	
	TAI 43/2016	CATTLE	09-Aug-16	O	POS	O	
	TAI 44/2016	CATTLE	09-Aug-16	O	POS	O	
	TAI 45/2016	CATTLE	15-Aug-16	O	POS	O	
THAILAND	TAI 46/2016	CATTLE	22-Aug-16	O	POS	O	
	TAI 47/2016	CATTLE	26-Aug-16	O	POS	O	
	TAI 48/2016	CATTLE	29-Aug-16	O	POS	O	
	TAI 49/2016	CATTLE	01-Sep-16	O	POS	O	
	TAI 50/2016	CATTLE	02-Sep-16	O	POS	O	
	TAI 51/2016	CATTLE	02-Sep-16	O	POS	O	
	TAI 52/2016	CATTLE	04-Nov-16	A	POS	A	
	TAI 53/2016	CATTLE	08-Nov-16	A	POS	A	
	TAI 54/2016	CATTLE	11-Nov-16	A	POS	A	
	TAI 55/2016	CATTLE	28-Nov-16	O	POS	O	
	TAI 56/2016	CATTLE	29-Nov-16	O	POS	O	

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Country	WRL for FMD Sample Identification	Animal	Date of Collection	Results		Final report
				VI/ELISA	RT-PCR	
	TAI 57/2016	CATTLE	29-Nov-16	A	POS	A
	TAI 58/2016	CATTLE	30-Nov-16	A	POS	A
	TAI 59/2016	CATTLE	30-Nov-16	O	POS	O
	TAI 60/2016	CATTLE	30-Nov-16	A	POS	A
	TAI 1/2017	SWINE	10-Jan-17	O	POS	O
	TAI 2/2017	CATTLE	11-Jan-17	A	POS	A
	TAI 3/2017	CATTLE	13-Jan-17	A	POS	A
	TAI 4/2017	CATTLE	19-Jan-17	A	POS	A
	TAI 5/2017	CATTLE	03-Feb-17	A	POS	A

Abbreviations used in table

FMD(V)	Foot-and-mouth disease (virus)
FMDV GD	Genome detected
FMDV NGD	Genome not detected (samples submitted in Trizol, only rRT-PCR carried out)
VI/ELISA	FMDV serotype identified following virus isolation in cell culture and antigen ELISA
rRT-PCR	Real-time reverse transcription polymerase chain reaction on epithelial suspension for FMD (or SVD) viral genome
NVD	No foot-and-mouth disease, swine vesicular disease or vesicular stomatitis virus detected
NT	Not tested

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



4.3. Antigenic Characterisation

Antigenic characterisation of FMD field isolates by matching with vaccine strains by 2dmVNT from April to June 2017.

Table 4: Vaccine matching studies for O FMDV by VNT

Strain	Serotype	Topotype	Strain	O 3039	O1 Manisa	O/TUR/5/2009
ETH/2/2017	O	EA-3	n/d	N	N	Y
ETH/11/2016	O	EA-4	n/d	N	N	Y

Table 5: Vaccine matching studies for A FMDV by VNT

Strain	Serotype	Topotype	Strain	A/IRN/05	A/TUR/20/06	A22 IRAQ	A TUR 11	A TUR 14	A CRUZ BVS
ALG/2/2017	A	AFRICA	G-IV	N	N	B	n/d	n/d	N
ALG/3/2017	A	AFRICA	G-IV	N	N	B	n/d	n/d	N

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



Abbreviations used in tables

M	Vaccine Match <i>$r_1 = \geq 0.3$. Suggests that there is a close relationship between field isolate and vaccine strain. A potent vaccine containing the vaccine strain is likely to confer protection.</i>
N	No Vaccine Match <i>$r_1 = < 0.3$. Suggests that the field isolate is so different from the vaccine strain that the vaccine is unlikely to protect</i>
B	Borderline <i>Any r_1 values between 0.28 to 0.32</i>
NT	Not tested against this vaccine

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



5. Annex 2

Recent FMD Publications (April to June 2016) cited by Web of Science (Pirbright Institute papers and authors are highlighted in **BOLD AND GREY**)

1. Abd El-Rahim, I.H.A., A.H. Asghar, A.M. Mohamed, and S.M. Fat'hi (2016). The impact of importation of live ruminants on the epizootiology of Foot and mouth disease in Saudi Arabia. *Revue scientifique et technique (International Office of Epizootics)*, **35**(3): 769-778.
2. Ahmed, A., A. Latif, R. Zahra, A.B. Zahur, U. Farooq, A. Ullah, M. Abubakar, A. Jamil, and M. Afzal (2017). Current appraisal of the suitability of Foot-and-mouth disease strains in two commonly used commercial vaccines for control of FMD in Pakistan. *Journal of Animal and Plant Sciences*, **27**(2): 446-450.
3. Amjad, K., M.H. Mushtaq, A. Mansur-ud-Din, F. Zahida, and K. Asghar (2017). Seasonal trends in seroprevalence of FMD in bovines under different environmental conditions in rural KPK, Pakistan. *Pakistan Veterinary Journal*, **37**(1): 55-58.
4. Arzt, J., B. Brito, S.J. Pauszek, E.J. Hartwig, G.R. Smoliga, L.T. Vu, P.P. Vu, C. Stenfeldt, L.L. Rodriguez, N.T. Long, and D.H. Dung (2017). Genome Sequence of Foot-and-Mouth Disease Virus Serotype O Lineage Ind-2001d Collected in Vietnam in 2015. *Genome Announcements*, **5**(18).
5. Arzt, J., J.M. Pacheco, C. Stenfeldt, and L.L. Rodriguez (2017). Pathogenesis of virulent and attenuated *Foot-and-mouth disease virus* in cattle. *Virology Journal*, **14**(1): 89-89.
6. Bahar e, M., A. Muhammad, A. Sibtain, A. Muhammad, M.A. Ashraf, U.H. Khan, F. Ahad, and W. Abdul (2016). Experimental trials of Foot-and-mouth disease vaccines with different vegetable oil adjuvants in goats under field conditions. *Pakistan Journal of Life and Social Sciences*, **14**(3): 178-182.
7. Barrette, R.W., J.M. Rowland, F.R. Grau, and M.T. McLntosh (2017). Development of a Feature and Template-Assisted Assembler and Application to the Analysis of a Foot-and-Mouth Disease Virus Genotyping Microarray. *Plos One*, **12**(1).
8. Bohorquez, J.A., S. Defaus, S. Munoz-Gonzalez, M. Perez-Simo, R. Rosell, L. Fraile, F. Sobrino, D. Andreu, and L. Ganges (2017). A bivalent dendrimeric peptide bearing a T-cell epitope from *Foot-and-mouth disease virus* protein 3A improves humoral response against *Classical swine fever virus*. *Virus Research*, **238**: 8-12.
9. Boklund, A., S. Mortensen, M.H. Johansen, and T. Halasa (2017). Resource Estimations in Contingency Planning for Foot-and-Mouth Disease. *Frontiers in Veterinary Science*, **4**: 64-64.
10. Bouguedour, R. and A. Ripani (2016). Review of the foot and mouth disease situation in North Africa and the risk of introducing the disease into Europe. *Revue scientifique et technique (International Office of Epizootics)*, **35**(3): 757-768.

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



11. Bown, C.P. and J.A. Hillman (2017). Foot-and-Mouth Disease and Argentina's Beef Exports: The WTO's US-Animals Dispute. *World Trade Review*, **16**(2): 253-277.
12. Brito, B., S.J. Pauszek, M. Eschbaumer, C. Stenfeldt, H.C. de Carvalho Ferreira, L.T. Vu, N.T. Phuong, B.H. Hoang, N.D. Tho, P.V. Dong, P.Q. Minh, N.T. Long, **D.P. King**, **N.J. Knowles**, D.H. Dung, L.L. Rodriguez, and J. Arzt (2017). Phylodynamics of *Foot-and-mouth disease virus* O/PanAsia in Vietnam 2010-2014. *Veterinary Research*, **48**(1): 24-24.
13. Byoungjuchoi, 김희진, 오세은, and 안세혁 (2017). Real-time Monitoring Method of Cattle's Temperature for FMD Prevention and Its Case Studies. *Journal of Korean Institute of Information Technology*, **15**(5): 141-150.
14. Cairns, A., T. Tolhurst, K. Poon, A.P. Ker, S. Duff, D. Jacques, and L. Yang (2017). The economic impact of a Foot-and-mouth disease outbreak for Ontario's beef sector. *Canadian Journal of Agricultural Economics*, **65**(1): 159-183.
15. Cha, C.-N., E.-K. Park, C.-Y. Yoo, S. Kim, Y.W. Yun, and H.-J. Lee (2017). Blood parameter changes in Korean traditional calves and pigs after foot-and-mouth disease vaccination. *Korean Journal of Veterinary Research*, **57**(1): 43-45.
16. Chen, Z., S. Zhang, Z. Li, G. Ma, and Z. Su (2017). Construction of a stable w/o nano-emulsion as a potential adjuvant for *Foot-and-mouth disease virus* vaccine. *Artificial Cells Nanomedicine and Biotechnology*, **45**(5): 897-906.
17. Countryman, A.M. and A.D. Hagerman (2017). Retrospective Economic Analysis of Foot and Mouth Disease Eradication in the Latin American Beef Sector. *Agribusiness*, **33**(2): 257-273.
18. de la Higuera, I., C. Ferrer-Orta, A.I. de Avila, C. Perales, M. Sierra, K. Singh, S.G. Sarafianos, Y. Dehouck, U. Bastolla, N. Verdaguer, and E. Domingo (2017). Molecular and Functional Bases of Selection against a Mutation Bias in an RNA Virus. *Genome Biology and Evolution*, **9**(5): 1212-1228.
19. Dill, V., M. Beer, and B. Hoffmann (2017). Simple, quick and cost-efficient: A universal RT-PCR and sequencing strategy for genomic characterisation of Foot-and-mouth disease viruses. *Journal of Virological Methods*, **246**: 58-64.
20. dos Santos, D.V., G. Sousa e Silva, E.J. Weber, H. Hasenack, F.H. Sautter Groff, B. Todeschini, M.R. Borba, A.A. Rosa Medeiros, V.B. Leotti, C.W. Canal, and L.G. Corbellini (2017). Identification of Foot-and-mouth disease risk areas using a multi-criteria analysis approach. *Plos One*, **12**(5).
21. Durrani, A.Z., F.A. Khan, A. Khan, I. Haq, and A. Khan. *Comparative study of Immune potency of two different trivalent FMD Vaccines in Cattle*, in *Proceedings of 2017 14th International Bhurban Conference on Applied Sciences and Technology*, M. Zafaruzzaman, et al., Editors. 2017. p. 170-173.
22. El-Khabaz, K.A.S. and A.A.T. Al-Hosary (2017). Detection and identification of *Foot-and-mouth disease virus* serotypes in Assiut governorate, Egypt. *Journal of Advanced Veterinary and Animal Research*, **4**(1): 32-38.
23. El-Rahim, I.H.A.A., A.H. Asghar, A.M. Mohamed, and S.M. Fat'hi (2016). The impact of importation of live ruminants on the epizootiology of Foot-and-mouth disease in Saudi Arabia. *Revue Scientifique Et Technique-Office International Des Epizooties*, **35**(3): 769-778.

The content of this report is the property of WRLFMD®, The Pirbright Institute.



24. Enright, J.A. and A. O'Hare (2017). Reconstructing disease transmission dynamics from animal movements and test data. *Stochastic Environmental Research and Risk Assessment*, **31**(2): 369-377.
25. Ferreira, H.C.d.C., S.J. Pauszek, **A. Ludi**, C.L. Huston, J.M. Pacheco, V.T. Le, P.T. Nguyen, H.H. Bui, T.D. Nguyen, T. Nguyen, T.T. Nguyen, L.T. Ngo, D.H. Do, L. Rodriguez, and J. Arzt (2017). An Integrative Analysis of *Foot-and-Mouth Disease Virus* Carriers in Vietnam Achieved Through Targeted Surveillance and Molecular Epidemiology. *Transboundary and Emerging Diseases*, **64**(2): 547-563.
26. **Fishbourne, E., A.B. Ludi, G. Wilsden, P. Hamblin, B. Statham, A. Bin-Tarif, E. Brocchi, S. Grazioli, A. Dekker, P. Eble, and D.P. King** (2017). Efficacy of a high potency O1 Manisa foot-and-mouth disease vaccine in cattle against heterologous challenge with a field virus from the O/ME-SA/Ind-2001 lineage collected in North Africa. *Vaccine*.
27. Galdo Novo, S., V. Malirat, E.D. Maradei, A.M. Espinoza, E. Smitsaart, A.R. Pedemonte, N. Mattion, and I.E. Bergmann (2017). Antigenic and immunogenic spectrum of Foot-and-mouth disease vaccine strain O1 Campos against representative viruses of topotypes that circulated in Asia over the past decade. *Vaccine*.
28. Govindaraj, G., B. Ganeshkumar, K.R. Nethrayini, R. Shalini, V. Balamurugan, B. Pattnaik, and H. Rahman (2017). Farm Community Impacts of Foot-and-Mouth Disease Outbreaks in Cattle and Buffaloes in Karnataka State, India. *Transboundary and Emerging Diseases*, **64**(3): 849-860.
29. Guan, S.H. and G.J. Belsham (2017). Separation of *Foot-and-mouth disease virus* leader protein activities; identification of mutants that retain efficient self-processing activity but poorly induce eIF4G cleavage. *Journal of General Virology*, **98**(4): 671-680.
30. **Gubbins, S., J. Forster, S. Clive, D. Schley, S. Zuber, J. Schaaf, and D. Corley** (2016). Thermal inactivation of *Foot-and-mouth disease virus* in extruded pet food. *Revue Scientifique Et Technique-Office International Des Epizooties*, **35**(3): 965-972.
31. Hatem, A.A. and M.A. Talal (2016). The effect of FMD disease on 3rd trimester pregnant cows and their fetuses. *Basrah Journal of Veterinary Research*, **15**(3): 475-480.
32. Hegde, R., S. Kowalli, K. Nagaraja, N.K. Dharanesha, C.M. Seema, T.A. Khan, G.V. Nagaraj, K. Srikala, K.J. Sudharshana, D. Nagaraju, S. Rao, P. Giridhara, and S.M. Byregowda (2016). Serosurveillance of Foot-and-mouth disease in Karnataka state, India: a 3 years study. *Virusdisease*, **27**(3): 294-302.
33. Hole, K., F. Ahmadpour, J. Krishnan, C. Stansfield, J. Copps, and C. Nfon (2017). Efficacy of accelerated hydrogen peroxide(®) disinfectant on *Foot-and-mouth disease virus*, *Swine vesicular disease virus* and *Senecavirus A*. *Journal of Applied Microbiology*, **122**(3): 634-639.
34. Houndje, E.M.B., C.A. Ogni, N. Noudeke, S. Farougou, A.K.I. Youssao, and T.M. Kpodekon (2016). Ethno-veterinary recipes of medicinal plants using for the treatment of Foot-and-mouth disease in Benin. *International Journal of Biological and Chemical Sciences*, **10**(5): 2090-2107.

The content of this report is the property of WRLFMD®, The Pirbright Institute.



35. **Howson, E.L.A., B. Armson, M. Madi**, C.J. Kasanga, S. Kandusi, R. Sallu, E. Chepkwony, A. Siddle, P. Martin, J. Wood, **V. Mioulet, D.P. King**, T. Lembo, S. Cleaveland, and **V.L. Fowler** (2017). Evaluation of Two Lyophilized Molecular Assays to Rapidly Detect *Foot-and-mouth disease virus* Directly from Clinical Samples in Field Settings. *Transboundary and Emerging Diseases*, **64**(3): 861-871.
36. Imran, I., I. Altaf, M. Ashraf, A. Javeed, N. Munir, and R. Bashir (2016). *In vitro* evaluation of antiviral activity of leaf extracts of *Azadirachta indica*, *Moringa oleifera*, and *Morus alba* against the *Foot-and-mouth disease virus* on BHK-21 cell line. *Scienceasia*, **42**(6): 392-396.
37. Isabel de Avila, A., E. Moreno, C. Perales, and E. Domingo (2017). Favipiravir can evoke lethal mutagenesis and extinction of *Foot-and-mouth disease virus*. *Virus Research*, **233**: 105-112.
38. Jonguk, L., Yongwha Chung, 노병준, and 박대희 (2017). Crisis Management Analysis of Foot-and-Mouth Disease Using Multi-dimensional Data Cube. *The Journal of the Korea Contents Association*, **17**(5): 565-573.
39. Kaszta, Z., J. Marino, and E. Wolff (2017). Fine-scale spatial and seasonal rangeland use by cattle in a foot-and-mouth disease control zones. *Agriculture Ecosystems & Environment*, **239**: 161-172.
40. Kim, S., H. Kwon, S. Park, H. Jeon, J.-k. Park, and J. Park (2017). Pilot-Scale Bio-Augmented Aerobic Composting of Excavated Foot-And-Mouth Disease Carcasses. *Sustainability*, **9**(3).
41. Kim, T., S. Ryoo, J. Nah, M. Sagong, S. Lee, K. Lee, Y. Ko, J. Park, M. Lee, S. Wee, D. Tark, and B. Ku (2017). Complete genome sequence of a *Foot-and-mouth disease virus* of serotype O, isolated from Gochang, Republic of Korea, in 2016. *Genome Announcements*, **5**(10): e01671-16.
42. **Knowles, N.J., J. Wadsworth, K. Bachanek-Bankowska**, and **D.P. King** (2016). VP1 sequencing protocol for *Foot and mouth disease virus* molecular epidemiology. *Revue scientifique et technique (International Office of Epizootics)*, **35**(3): 741-755.
43. Ko, C.-R., S.-S. Seol, and G. Kim (2017). Political Response to Foot-and-Mouth Disease: A Review of Korean News. *Sustainability*, **9**(3).
44. Lawrence, P. and E. Rieder (2017). Insights into Jumonji C-domain containing protein 6 (JMJD6): a multifactorial role in *Foot-and-mouth disease virus* replication in cells. *Virus Genes*.
45. Lee, S.-Y., Y.-J. Lee, R.-H. Kim, J.-N. Park, M.-E. Park, M.-K. Ko, J.-H. Choi, J.-Q. Chu, K.-N. Lee, S.-M. Kim, D. Tark, H.-S. Lee, Y.-J. Ko, M.-G. Seo, J.-W. Park, B. Kim, M.-H. Lee, J.-S. Lee, and J.-H. Park (2017). Rapid Engineering for Vaccine and Challenge Viruses against Foot-and-Mouth Disease. *Journal of Virology*.
46. Leng, Y., M. Ren, Z. Meng, P. Zhang, Z. Yang, X. Yao, and Y. Wang (2017). Development of multiplex PCR for detection of 5 pathogenic viruses to pigs. *Zhongguo Yufang Shouyi Xuebao*, **39**(2): 123-126.
47. Liu, X., Y. Fang, P. Zhou, Y. Lu, Q. Zhang, S. Xiao, Z. Dong, L. Pan, J. Lv, Z. Zhang, Y. Zhang, and Y. Wang (2017). Chimeric virus-like particles elicit protective immunity against serotype O *Foot-and-mouth disease virus* in guinea pigs. *Applied Microbiology and Biotechnology*.

The content of this report is the property of WRLFMD®, The Pirbright Institute.



48. Liu, Z., J. Shao, F. Zhao, G. Zhou, S. Gao, W. Liu, J. Lv, X. Li, Y. Li, H. Chang, and Y. Zhang (2017). Chemiluminescence immunoassay for the detection of antibodies against the 2C and 3ABC nonstructural proteins induced by infecting pigs with the *Foot-and-mouth disease virus*. *Clinical and vaccine immunology : CVI*.
49. Lotufo, C.M., I.E. Bergmann, N.M. Mattion, M. Wilda, and P.R. Grigera (2017). Recombinant *Foot-and-mouth disease virus* (FMDV) non-structural protein 3A fused to enhanced green fluorescent protein (EGFP) as a candidate probe to identify FMDV-infected cattle in serosurveys. *Archives of Virology*.
50. Lozovoi, D. (2016). Implementation of a complex of joint measures by member states of the Commonwealth of Independent States for prophylaxis and control of foot and mouth disease. *Mezhdunarodnyi Sel'skokhozyaistvennyi Zhurnal*, (4): 53-58.
51. Mahmoud, M.A. and S.A. Galbat (2017). Outbreak of foot and mouth disease and Peste des petits ruminants in sheep flock imported for immediate slaughter in Riyadh. *Veterinary World*, **10**(2): 238-243.
52. Meyer, A., L. Zamir, A. Ben Yair Gilboa, B. Gelman, D.U. Pfeiffer, and T. Vergne (2017). Quantitative Assessment of the Risk of Release of *Foot-and-Mouth Disease Virus* via Export of Bull Semen from Israel. *Risk analysis : an official publication of the Society for Risk Analysis*.
53. Mischenko, A.V., V.A. Mischenko, P. Bolortuya, T. Purevkhoo, and K. Batamgalan (2017). Distribution of Foot-and-mouth disease in Mongolia. *Veterinariya*, (2): 23-25.
54. Mohanty, N.N., B. Das, L.N. Sarangi, S. Subramaniam, J.K. Mohapatra, and H.K. Panda (2016). Isolation and characterization of foot-and-mouth disease virus from Odisha, India. *Tropical Biomedicine*, **33**(4): 753-760.
55. Nelson, N., **D.J. Paton, S. Gubbins, C. Colenutt, E. Brown, S. Hodgson**, and J.L. Gonzales (2017). Preclinical diagnosis: predicting its ability to improve control of farm-to-farm Foot-and-mouth disease transmission in cattle. *Journal of Clinical Microbiology*.
56. Pamaranon, N., S. Kasemsuwan, and T. Rukkwamsuk (2016). Quantitative risk assessment of the introduction of *Foot-and-mouth disease virus* by human into fattening pig farms in the compartmentalization. *Journal of Kasetsart Veterinarians*, **26**(2): 83-99.
57. Paprocka, G. and A. Kesy (2017). Epidemiological situation of foot-and-mouth disease in 2014-2015 and the beginning of 2016. *Medycyna Weterynaryjna*, **73**(3): 131-135.
58. Pomeroy, L.W., S. Bansal, M. Tildesley, K.I. Moreno-Torres, M. Moritz, N. Xiao, T.E. Carpenter, and R.B. Garabed (2017). Data-Driven Models of Foot-and-Mouth Disease Dynamics: A Review. *Transboundary and Emerging Diseases*, **64**(3): 716-728.
59. Prajapati, B.M., J.P. Gupta, D.P. Pandey, G.A. Parmar, and J.D. Chaudhari (2017). Molecular markers for resistance against infectious diseases of economic importance. *Veterinary World*, **10**(1): 112-120.
60. Puckette, M., T. Burrage, J.G. Neilan, and M. Rasmussen (2017). Evaluation of *Gussia luciferase* and *Foot-and-mouth disease virus* 2A translational

The content of this report is the property of WRLFMD®, The Pirbright Institute.



- interrupter chimeras as polycistronic reporters for transgene expression. *Bmc Biotechnology*, **17**.
61. Rai, D.K., F. Diaz-San Segundo, G. Campagnola, A. Keith, E.A. Schafer, A. Kloc, T. de Los Santos, O. Peersen, and E. Rieder (2017). Attenuation of *Foot-and-Mouth Disease Virus* by Engineering Viral Polymerase Fidelity. *Journal of virology*.
 62. Rodriguez Pulido, M. and M. Saiz (2017). Molecular Mechanisms of *Foot-and-Mouth Disease Virus* Targeting the Host Antiviral Response. *Frontiers in Cellular and Infection Microbiology*, **7**.
 63. Rout, M., S. Subramaniam, B. Das, J.K. Mohapatra, B.B. Dash, A. Sanyal, and B. Pattnaik (2017). Foot-and-mouth disease in wildlife population of India. *Indian Journal of Animal Research*, **51**(2): 344-346.
 64. Ryoo, S., T. Kim, J. Nah, M. Sagong, S. Lee, K. Lee, Y. Ko, J. Park, M. Lee, S. Wee, D. Tark, and B. Ku (2017). Complete genome sequence of a *Foot-and-mouth disease virus* of serotype O isolated from Gimje, Republic of Korea, in 2016. *Genome Announcements*, **5**(10): e01694-16.
 65. Santos, D.V.D., G.S.E. Silva, E.J. Weber, H. Hasenack, F.H.S. Groff, B. Todeschini, M.R. Borba, A.A.R. Medeiros, V.B. Leotti, C.W. Canal, and L.G. Corbellini (2017). Identification of Foot-and-mouth disease risk areas using a multi-criteria analysis approach. *PloS one*, **12**(5): e0178464-e0178464.
 66. Scott, K.A., N.M. Rathogwa, A.V. Capozzo, and F.F. Maree (2017). Evaluation of immune responses of stabilised SAT2 antigens of foot-and-mouth disease in cattle. *Vaccine*.
 67. Senthilkumaran, C., M. Yang, H. Bittner, A. Ambagala, O. Lung, J. Zimmerman, L.G. Gimenez-Lirola, and C. Nfon (2017). Detection of genome, antigen, and antibodies in oral fluids from pigs infected with *Foot-and-mouth disease virus*. *Canadian Journal of Veterinary Research*, **81**(2): 82-90.
 68. Shi, X., G. Ma, Y. Bai, J. Hao, C. Qiao, Q. Liu, Y. Li, and B. Li (2017). Simultaneous detection of five bovine viruses by multiplex ligation-dependent probe amplification. *Chinese Journal of Veterinary Science*, **37**(2): 224-230.
 69. Singh, R., R. Alex, U. Singh, S. Kumar, G.S. Sengar, T.V. Raja, R.R. Alyethodi, A. Kumar, and R. Deb (2017). A Synonymous Mutation at Bovine α Vitronectin Domain of Integrin Host Receptor (ITGAV) Gene Effect the Susceptibility of Foot-and-Mouth Disease in Crossbred Cattle. *Advances in Experimental Medicine and Biology*.
 70. Song, S., B. Zhao, P. Qu, D. Hu, Y. Sun, H. Dong, L. Yang, X. Song, and S. Pan (2017). Establishment of multiplex TaqMan real-time PCR for the detection of bovine viral diarrhoea virus, *Infectious bovine rhinotracheitis virus* and *Foot-mouth disease virus*. *Zhongguo Yufang Shouyi Xuebao*, **39**(2): 133-136.
 71. Souley Kouato, B., F.M. Elliot, **D.P. King**, J. Hyera, **N.J. Knowles**, **A.B. Ludi**, **V. Mioulet**, G. Matlho, K. De Clercq, E. Thys, H. Marichatou, S. Issa, and C. Saegerman (2017). Outbreak investigations and molecular characterization of Foot-and-mouth disease viruses circulating in south-west Niger. *Transboundary and Emerging Diseases*.
 72. Stenfeldt, C., J. Arzt, G. Smoliga, M. LaRocco, J. Gutkoska, and P. Lawrence (2017). Proof-of-concept study: profile of circulating microRNAs in Bovine

The content of this report is the property of WRLFMD®, The Pirbright Institute.



- serum harvested during acute and persistent FMDV infection. *Virology Journal*, **14**.
73. Tekleghiorghis, T., K. Weerdmeester, F. van Hemert-Kluitenberg, R.J.M. Moormann, and A. Dekker (2017). Foot-and-Mouth Disease Seroprevalence in Cattle in Eritrea. *Transboundary and Emerging Diseases*, **64**(3): 754-763.
 74. Van der Wiele, A. (2017). Foot-and-mouth disease training in Nepal. *Tijdschrift Voor Diergeneeskunde*, **142**(2): 46-47.
 75. Vandebussche, F., D.J. Lefebvre, U. De Leeuw, S. Van Borm, and K. De Clercq (2017). Laboratory validation of two real-time RT-PCR methods with 5'-tailed primers for an enhanced detection of *Foot-and-mouth disease virus*. *Journal of Virological Methods*, **246**: 90-94.
 76. Vosloo, W., A.D. Bastos, E. Kirkbride, D.J. Esterhuysen, D.J. vanRensburg, R.G. Bengis, D.W. Keet, and G.R. Thomson (1996). Persistent infection of African buffalo (*Syncerus caffer*) with SAT-type Foot-and-mouth disease viruses: Rate of fixation of mutations, antigenic change and interspecies transmission. *Journal of General Virology*, **77**: 1457-1467.
 77. Vu, L.T., N.T. Long, B. Brito, C. Stenfeldt, N.T. Phuong, B.H. Hoang, S.J. Pauszek, E.J. Hartwig, G.R. Smoliga, P.P. Vu, L.T.V. Quang, V.V. Hung, N.D. Tho, P.V. Dong, P.Q. Minh, M. Bertram, I.H. Fish, L.L. Rodriguez, D.H. Dung, and J. Arzt (2017). First detection of *Foot-and-mouth disease virus* O/Ind-2001d in Vietnam. *Plos One*, **12**(6).
 78. Wada, M., M. Stevenson, N. Cogger, and T. Carpenter (2017). Evaluation of the Control Strategy for the 2010 Foot-and-Mouth Disease Outbreak in Japan Using Disease Simulation. *Transboundary and Emerging Diseases*, **64**(3): 978-989.
 79. Willeberg, P.W., M. Alkhamis, A. Boklund, A.M. Perez, C. Enoe, and T. Halasa (2017). Semiquantitative Decision Tools for FMD Emergency Vaccination Informed by Field Observations and Simulated Outbreak Data. *Frontiers in Veterinary Science*, **4**: 43-43.
 80. Wnek, J. (2016). Eradication of Foot-and-mouth disease in Poland in 1918-1939. *Zycie Weterynaryjne*, **91**(12): 944-945.
 81. Wungak, Y.S., O.O. Ishola, B.O. Olugasa, D.D. Lazarus, D.O. Ehizibolo, and H.G. Ularamu (2017). Spatial pattern of foot-and-mouth disease virus serotypes in North Central Nigeria. *Veterinary World*, **10**(4): 450-456.
 82. Yan, D., Z. Teng, S. Sun, S. Jiang, H. Dong, Y. Gao, Y. Wei, W. Qin, X. Liu, H. Yin, and H. Guo (2017). *Foot-and-mouth disease virus*-like particles as integrin-based drug delivery system achieve targeting anti-tumor efficacy. *Nanomedicine: Nanotechnology, Biology, and Medicine*, **13**(3): 1061-1070.
 83. Yan, H., J. Wang, M. Zhu, H. Miao, Z. Peng, H. Li, and A. Xin (2017). Complete Genome Sequence of *Foot-and-Mouth Disease Virus* Serotype O Strain YNTBa from Yunnan Province of China. *Genome Announcements*, **5**(22).
 84. Yang, Y., Q. Zhao, Z. Li, L. Sun, G. Ma, S. Zhang, and Z. Su (2017). Stabilization study of inactivated *Foot and mouth disease virus* vaccine by size-exclusion HPLC and differential scanning calorimetry. *Vaccine*.
 85. You, S.-H., T. Kim, J.-H. Choi, G. Park, K.-N. Lee, B. Kim, M.-H. Lee, H.-S. Kim, S.-M. Kim, and J.-H. Park (2017). Coinjection of a vaccine and anti-viral

The content of this report is the property of WRLFMD®, The Pirbright Institute.



- agents can provide fast-acting protection from Foot-and-mouth disease. *Antiviral Research*, **143**: 195-204.
86. Zeng, B., R. Li, H. Xiao, and S. Chen (2017). Advances in molecular genetic basis of disease resistance differences among domestic cattle breeds. *Acta Veterinaria et Zootechnica Sinica*, **48**(2): 193-200.
 87. **Zhang, F., E. Perez-Martin, N. Juleff, B. Charleston, and J. Seago** (2017). A replication-competent *Foot-and-mouth disease virus* expressing a luciferase reporter. *Journal of Virological Methods*, **247**: 38-44.
 88. Zhao, B., Y. Liu, Y. Chen, P. Ji, J. Wang, C. Liu, S. Yang, and G. Zhang (2017). Soluble expression and immunoreactivity analysis of FMDV VP0 and VP1 protein. *Journal of Henan Agricultural Sciences*, **46**(2): 105-110.
 89. Zhao, F.-R., Y.-L. Xie, Z.-Z. Liu, J.-J. Shao, S.-F. Li, P. Zhou, Y.-G. Zhang, and H.-Y. Chang (2017). Transcriptomic Analysis of porcine PBMCs in Response to FMDV Infection. *Acta Tropica*.
 90. Zhao, H., G. Teni, J. Lu, T. Huang, Q. Xiao, and L. Geri (2017). Effect of various types FMDV VP1 gene epitope embedded protein SLP-MultiVP1 on IgG subtypes of different immune mice. *Chinese Journal of Veterinary Science*, **37**(2): 262-265.
 91. Zhu, Z., C. Li, X. Du, G. Wang, W. Cao, F. Yang, H. Feng, X. Zhang, Z. Shi, H. Liu, H. Tian, D. Li, K. Zhang, X. Liu, and H. Zheng (2017). *Foot-and-mouth disease virus* infection inhibits LGP2 protein expression to exaggerate inflammatory response and promote viral replication. *Cell Death and Disease*, **8**(4): e2747-e2747.

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk



6. Annex 3

RECOMMENDATIONS FROM WRLFMD® ON FMD VIRUS STRAINS TO BE INCLUDED IN FMDV ANTIGEN BANKS (FOR FMD-FREE COUNTRIES)

June 2017:

Note: Virus strains are NOT listed in order of importance

High Priority	A/ASIA/G-VII(G-18)* O Manisa O PanAsia-2 (or equivalent) Asia 1 Shamir A Iran-05 (or A TUR 06) A22 Iraq A24 Cruzeiro O BFS or Campos SAT 2 Saudi Arabia (or equivalent i.e. SAT 2 Eritrea)
Medium Priority	A Eritrea-98 SAT 2 Zimbabwe SAT 1 South Africa A Malaysia 97 (or Thai equivalent such as A/Sakolnakorn/97) A Argentina 2001 O Taiwan 97 (pig-adapted strain or Philippine equivalent)
Low Priority	A Iran '96 A Iran '99 A Iran 87 or A Saudi Arabia 23/86 (or equivalent) A15 Bangkok related strain A87 Argentina related strain C Noville SAT 2 Kenya SAT 1 Kenya SAT 3 Zimbabwe

Note: Discussions are currently underway to adopt a risk-based approach for different FMD viral lineages to identify priority vaccines for use in Europe and other FMD-free settings.

*Recent *in vitro* data from WRLFMD for serotype A viruses from Saudi Arabia and Iran highlights an apparent gap in vaccines supplied by international manufacturers for this viral lineage.

The content of this report is the property of WRLFMD®, The Pirbright Institute.

Copies of all the individual reports cited herein can be obtained from The Pirbright Institute and prior to presentation, publication or any other public use of these data, please contact Dr Donald King, The Pirbright Institute, donald.king@pirbright.ac.uk