



ISSN 1014-2339

ANIMAL GENETIC RESOURCES INFORMATION

BULLETIN D'INFORMATION SUR LES RESSOURCES GÉNÉTIQUES ANIMALES

BOLETÍN DE INFORMACIÓN SOBRE RECURSOS GENÉTICOS ANIMALES



Food and
Agriculture
Organization
of the
United
Nations

Organisation
des
Nations
Unies
pour
l'alimentation
et
l'agriculture

Organización
de las
Naciones
Unidas
para la
Agricultura
y la
Alimentación



*Initiative for
Domestic
Animal
Diversity*

*Initiative pour
la Diversité
des Animaux
Domestiques*

*Iniciativa para
la Diversidad
de los Animales
Domésticos*

The designations employed and the presentation of material in this publication do not imply the expression of any opinion whatsoever on the part of the Food and Agriculture Organization of the United Nations concerning the legal status of any country, territory, city or area or of its authorities, or concerning the delimitation of its frontiers or boundaries.

Les appellations employées dans cette publication et la présentation des données qui y figurent n'impliquent de la part de l'Organisation des Nations Unies pour l'alimentation et l'agriculture aucune prise de position quant au statut juridique des pays, territoires, villes ou zones, ou de leurs autorités, ni quant au tracé de leurs frontières ou limites.

Las denominaciones empleadas en esta publicación y la forma en que aparecen presentados los datos que contiene no implican de parte de la Organización de las Naciones Unidas para la Agricultura y la Alimentación juicio alguno sobre la condición jurídica de países, territorios, ciudades o zonas, o de sus autoridades, ni respecto de la delimitación de sus fronteras o límites.

All rights reserved. No part of this publication may be reproduced, stored in a retrieval system, or transmitted in any form or by any means, electronic, mechanical, photocopying or otherwise, without the prior permission of the copyright owner. Applications for such permission, with a statement of the purpose and the extent of the reproduction, should be addressed to the Director, Information Division, Food and Agriculture Organization of the United Nations, Viale delle Terme di Caracalla, 00100 Rome, Italy.

Tous droits réservés. Aucune partie de cette publication ne peut être reproduite, mise en mémoire dans un système de recherche documentaire ni transmise sous quelque forme ou par quelque procédé que ce soit: électronique, mécanique, par photocopie ou autre, sans autorisation préalable du détenteur des droits d'auteur. Adresser une demande motivée au Directeur de la Division de l'information, Organisation des Nations Unies pour l'alimentation et l'agriculture, Viale delle Terme di Caracalla, 00100 Rome, Italie, en indiquant les passages ou illustrations en cause.

Reservados todos los derechos. No se podrá reproducir ninguna parte de esta publicación, ni almacenarla en ningún sistema de recuperación de datos o transmitirla en cualquier forma o por cualquier procedimiento (electrónico, mecánico, fotocopia, etc.), sin autorización previa del titular de los derechos de autor. Las peticiones para obtener tal autorización, especificando la extensión de lo que se desea reproducir y el propósito que con ello se persigue, deberán enviarse a la Dirección de Información, Organización de las Naciones Unidas para la Agricultura y la Alimentación, Viale delle Terme di Caracalla, 00100 Roma, Italia.

Editors - Editeurs - Editores:
S. Galal & J. Boyazoglu

Viale delle Terme di Caracalla 1, 00100
Rome, Italy

Animal Genetic Resources Information is published under the joint auspices of the Food and Agriculture Organization of the United Nations (FAO) and the United Nations Environment Programme (UNEP). It is edited in the Animal Genetic Resources Group of the Animal Production and Health Division of FAO. It is available direct from FAO or through the usual FAO sales agents.

ANIMAL GENETIC RESOURCES INFORMATION will be sent free of charge to those concerned with the sustainable development conservation of domestic livestock. Anyone wishing to receive it regularly should send their name and address to the Editor, at the address shown above.

AGRI can also be found in the "Library" of DAD-IS at URL <http://www.fao.org/dad-is>.

Le Bulletin d'information sur les ressources génétiques animales est publié sous les auspices conjoints de l'Organisation des Nations Unies pour l'Alimentation et l'Agriculture (FAO) et du Programme des Nations Unies pour l'Environnement (UNEP). Cette publication est éditée par le Groupe des Ressources Génétiques de la Division de la Production et de la Santé Animales de la FAO. On peut se le procurer directement au siège de la FAO ou auprès des dépositaires et agents habituels de vente de publication de l'Organisation.

LE BULLETIN D'INFORMATION SUR LES RESSOURCES GÉNÉTIQUES ANIMALES sera envoyé gratuitement aux personnes intéressées par le développement durable et la conservation du bétail domestique. Les personnes souhaitant recevoir cette publication régulièrement voudront bien faire parvenir leurs nom et adresse à l'éditeur, à l'adresse sus-indiquée.

AGRI peut être consulté également sur la "Librairie" de DAD-IS de URL <http://www.fao.org/dad-is>.

El Boletín de Información sobre Recursos Genéticos Animales se publica bajo los auspicios de la Organización de las Naciones Unidas para la Agricultura y la Alimentación (FAO) y del Programa de las Naciones Unidas para el Medio Ambiente (UNEP). Se edita en el Grupo de Recursos Genéticos de la Dirección de Producción y Sanidad Animal de la FAO. Se puede obtener directamente de la FAO o a través de sus agentes de venta habituales.

El BOLETIN DE INFORMACION SOBRE RECURSOS GENETICOS ANIMALES será enviado gratuitamente a quienes estén interesados en el desarrollo sostenible y la conservación del ganado doméstico. Si se desea recibirlo regularmente, se ruega comunicar nombre, apellido y dirección al editor a la dirección arriba indicada. AGRI puede consultarse también en la "Librería" de DAD-IS de URL <http://www.fao.org/dad-is>.

ANIMAL GENETIC
RESOURCES INFORMATION
BULLETIN
D'INFORMATION SUR LES
RESSOURCES GÉNÉTIQUES
ANIMALES
BOLETÍN DE
INFORMACIÓN SOBRE RECURSOS
GENÉTICOS ANIMALES



CONTENTS

| | Page |
|---|------|
| Editorial | i |
| Contribution to 25 th Issue of AGRI from the Founder Editor <i>J. Hodges</i> | vii |
| Past, present and future <i>J. Boyazoglu</i> | ix |
| The state of African cattle genetic resources. I. Classification framework and identification of threatened and extinct breeds <i>J.E.O. Rege</i> | 1 |
| Characteristics of Munjal sheep <i>B.P. Kushwaha, Riyazuddin, R. N. Singh & S. Parthasarathy</i> | 27 |
| Conservation of livestock breed diversity <i>J.S.F. Barker</i> | 33 |
| The development of a system of linear measurements to provide an assessment of type and function of beef cattle <i>G.L.H. Alderson</i> | 45 |
| A programme for the inventory, characterisation, evaluation, conservation and utilisation of European rabbit (<i>Oryctolagus cuniculus</i>) genetic resources <i>G. Bolet, M. Monnerot, C. Arnal, J. Arnold, D. Bell, G. Bergoglio, U. Besenfelder, S. Bosze, S. Boucher, J.M. Brun, N. Chanteloup, M.C. Ducourouble, M. Durand-Tardif, P.J. Esteves, N. Ferrand, G. Hewitt, T. Joly, P.F. Koehl, M. Laube, S. Lechevestrier, M. Lopez, G. Masoero, R. Piccinin, G. Queney, G. Saleil, A. Surridge, W. van der Loo, J. Vanhommerig, J.S. Vicente, G. Virag & J.M. Zimmermann</i> | 57 |
| The Peel-Neck chicken of Belize and Guatemala, Central America <i>J.G. Mallia</i> | 73 |
| European approaches to conservation of farm animal genetic resources <i>D.L. Simon</i> | 79 |
| Inbreeding and loss of founder alleles in four variations of a conservation programme using circular mating, for Danish Shorthorn Cattle <i>M. Trinderup, J.N. Jørgensen & M. Hansen</i> | 101 |
| Advances in the Brazilian animal genetic resources conservation programme <i>A. da S. Mariante, M. do S. M. Albuquerque, A. A. do Egito & C. McManus</i> | 109 |
| Indexes | 125 |
| Editorial policies and procedures | 153 |

FOOD AND AGRICULTURE ORGANIZATION OF THE UNITED NATIONS
ORGANISATION DES NATIONS UNIES POUR L'ALIMENTATION ET L'AGRICULTURE
ORGANIZACION DE LAS NACIONES UNIDAS PARA LA AGRICULTURA Y LA ALIMENTACION

UNITED NATIONS ENVIRONMENT PROGRAMME
PROGRAMME DES NATIONS UNIES POUR L'ENVIRONNEMENT
PROGRAMA DE LAS NACIONES UNIDAS PARA EL MEDIO AMBIENTE

Editorial

The state of the world's animal genetic resources ?

Animal genetic resources (AnGR) contribute substantial inputs to and outputs from most food and agriculture production systems, and must be wisely used, developed and conserved, as part of efforts to achieve and sustain world food security and rural development. In view of this importance, and of the increasing pace of change, local, national and regional communities must be empowered to act based upon answers to such key questions as:

- Do we have essential baseline data and other information on the status of animal genetic resources to establish local community, country, regional and global priorities for their effective and efficient management over time?
- Do all communities and countries have the capacity to manage these resources, in an increasingly interdependent world?
- Do we have in place the appropriate methodologies and technologies to best understand, use and develop, conserve and access these resources, in a sustainable manner?
- Are AnGRs capable of increasing production and productivity, or at least productivity alone, whilst also maintaining the levels of product quality demanded by the consuming community, identified and deployed to the farmers of each of the full range of important production environments ?
- Do we have sustainable intensification programmes in place for those AnGR currently being used by farmers?
- Do we have an adequate understanding of the status of breeds that are currently not of value to farmers, and of wild relatives of the domesticated animal species, to provide the foundation for an Early Warning System for Animal Genetic Resources at

risk of being lost, and to identify opportunities for the future use of these resources?

- Are we in a position to be able to identify situations where an emergency response could be considered to prevent the loss of animal genetic resources at risk?
- Do we have in place at the local, country, regional and global levels sustainable policies for wise use of AnGR?

The answer to these questions collectively seems to be no! This presents a real challenge for all those concerned with agricultural sustainable development and food security; whether they be farmers, researchers, educators, policy makers or administrators; and particularly if they are simply members of the consuming community; even more so if they are tomorrow's decision-makers, our children and theirs!

Through the help of its Member Governments and other national and international organizations, FAO has been assisting countries to establish breed databases, in DAD-IS, and publish two editions of the World Watch List for Domestic Animal Diversity that with analysis provide substantial amount of data on AnGR globally. But this data is far short from providing the information implied in the above questions which is needed for planning and implementing sound and sustainable management of our countries' AnGR.

The issue of the State of the World's AnGR (SoW-AnGR) now and regularly updating this to monitor AnGR development, is now a topic of discussion by country delegations in FAO governing body sessions. It was an important item on the agenda of the FAO Commission on Genetic Resources for Food and Agriculture that convened during April 1999 with delegations from over 140 countries and organizations. Delegations

resolved that FAO should lead and coordinate the development of a country-driven Report on the State of the World's Animal Genetic Resource as funds become available.

The SoW-AnGR Report, will specifically establish essential baseline information in major areas: the state of animal diversity; the state of country capacity to manage animal genetic resources; the state of the art, the available methodologies and technologies to assist farmers, breeders and scientists to better describe, use, develop, conserve and access animal genetic resources, and thereby contribute to global food security and sustainable rural development.

The SoW-AnGR Report will establish the critical baseline data and information required to enable cost-effective AnGR Management activities to be planned and implemented at the local, country, regional and global levels. It will also provide the first reliable global outlook to examine potential future roles for animal genetic resources, and help project likely developments.

Finally, the first SoW-AnGR Report will help stimulate sustainable intensification of food and agriculture production of the broad range of production systems in countries, regions and globally.

The Editors

Editorial

Quel est l'état des ressources génétiques animales dans le monde?

Les ressources génétiques animales (AnGR), contribuent de façon importante autant qu'elles recueillent de la plupart des systèmes de production agricole et alimentaire. Elles doivent donc être utilisées avec précaution et développées et conservées, ceci dans le but d'atteindre et de maintenir la sécurité alimentaire et le développement rural dans le monde. Etant donné cette importance et le déroulement des changements en cours, les communautés au niveau local, national et régional doivent être en mesure d'agir en faisant face aux questions suivantes:

- Avons-nous les données de base essentielles et autres informations nécessaires sur la situation des ressources génétiques animales pour pouvoir établir les priorités, au niveau des communautés, des pays, des régions et au niveau mondial; quelle sera la gestion effective et efficace dans le futur?
- Est-ce que toutes les communautés et les pays possèdent les modalités nécessaires pour gérer ces ressources étant donné une toujours grandissant interdépendance mondiale?
- Avons-nous sur place les méthodologies et technologies appropriées pour mieux comprendre, utiliser et développer, conserver et accéder à ces ressources de façon durable?
- Est-ce que les AnGR sont vraiment un moyen pour augmenter la production et la productivité, ou tout au moins seulement la productivité, tout en conservant la qualité des produits demandés par les consommateurs; identifier et déployer ses principes vers les agriculteurs de chacun des secteurs de production les plus importants?

- Avons-nous sur place des programmes d'intensification durables pour ces AnGR utilisées normalement par les agriculteurs?
- Possédons-nous une connaissance adéquate de la situation des races qui actuellement ne sont pas d'intérêt pour les agriculteurs, ainsi que des parents sauvages de ces espèces domestiques, pour jeter les bases pour un Système d'Alerte Rapide pour les AnGR qui sont en risque de disparition, et pour identifier les opportunités d'une utilisation future de ces ressources?
- Sommes-nous en position de pouvoir identifier les situations dans lesquelles une réponse urgente doit être prise en considération pour prévenir la perte de ressources génétiques animales à risque?
- Disposons-nous sur place, au niveau local, de pays, de région et au niveau mondial, de politiques durables pour une utilisation prudente des AnGR?

La réponse à cet ensemble de questions semble être non! Ceci représente un enjeu réel pour tous ceux concernés par le développement agricole durable et la sécurité alimentaire; aussi bien s'il s'agit d'agriculteurs, de chercheurs, d'enseignants, de politiciens ou d'administrateurs, et en particulier s'il s'agit simplement de membres de la société de consommateurs, encore plus si demain ils deviennent les "décideurs"!

A travers l'aide fournie par ses Gouvernements Membres et d'autres organisations nationales et internationales, la FAO a porté son aide aux pays pour établir des bases de données sur les races, DAD-IS, et a publié deux éditions de la Liste Mondiale de Surveillance pour la Diversité des Animaux Domestiques, qui, à travers son analyse, fourni un nombre important de données sur les AnGR au niveau mondial.

Cependant, toutes ces données sont bien loin de répondre aux questions posées précédemment, et qui sont nécessaires pour mettre en oeuvre et planifier une gestion appropriée et durable des AnGR de nos pays.

La question de l'Etat Mondial des AnGR (SoW-AnGR) aujourd'hui et sa mise à jour régulière pour contrôler le développement des AnGR, est actuellement un thème de discussion durant les séances au sein des délégations des pays membres de la FAO. Il s'agissait d'un des points importants de l'agenda de la Commission pour les Ressources Génétiques pour l'Agriculture et l'Alimentation qui s'est réunie au mois d'avril 1999 avec les délégations provenant de plus de 140 pays et organisations. Les délégations ont conclu que la FAO devrait diriger et coordonner le développement d'un rapport-guide pour les pays sur l'Etat des Ressources Génétiques Animales (SoW) selon la disponibilité de fonds.

Le rapport SoW devra établir spécifiquement les bases d'information essentielles pour les thèmes principaux: la

situation de la diversité animale; la situation de chaque pays à gérer les ressources génétiques animales; la situation actuelle, les méthodologies et technologies disponibles pour assister les éleveurs, les améliorateurs et les chercheurs à mieux décrire, utiliser, développer, conserver et accéder aux ressources génétiques animales, et ainsi contribuer à la sécurité alimentaire mondiale et au développement rural durable.

Le rapport SoW-AnGR établira les données guides indispensables et l'information requise pour permettre que les activités de gestion AnGR à planifier et à mettre en oeuvre au niveau local, de pays, régional et au niveau mondial aient une bonne relation coût/efficacité. Ce rapport représentera aussi la première perspective fiable au niveau mondial pour examiner les futurs rôles potentiels des ressources génétiques animales et pour aider les projets de développement.

Finalement, le premier rapport SoW-AnGR servira à stimuler une intensification durable de la production alimentaire et agricole à large échelle dans les systèmes de production des pays, des régions et au niveau mondial.

Les Editeurs

Editorial

Cual es el estado de los recursos genéticos animales en el mundo?

Los recursos genéticos animales (AnGR) contribuyen en parte importante a los insumos y las producciones de la mayoría de los sistemas agroalimentarios. Deberían ser aprovechados, desarrollados y conservados prudentemente como parte del esfuerzo aunado para conseguir y sostener la seguridad alimentaria mundial y el desarrollo rural. Dada esta importancia, y debido al ritmo creciente de los cambios, las comunidades locales, nacionales y regionales deben tener la facultad para actuar basándose en las respuestas a preguntas clave como:

- Disponemos de datos fundamentales y de información adicional sobre el estado de los recursos genéticos animales para poder establecer prioridades en las comunidades local, nacional, regional y a nivel mundial para su gestión efectiva y eficiente a lo largo del tiempo?
- Todas las comunidades y países, tienen la capacidad de gestionar estos recursos en un mundo cada vez más interdependiente?
- Están puestas en práctica las metodologías y tecnologías apropiadas para entender, aprovechar y desarrollar al máximo los AnGR y para conservar y acceder a los mismos de una manera sostenible?
- Los recursos genéticos animales, son capaces de aumentar la producción y la productividad (o al menos esta última), manteniendo a la vez los niveles de calidad de los productos, exigidos por la comunidad de consumidores, identificados y puestos a disposición de los productores en cada parte del espectro de los medios importantes de producción?
- Existen en la práctica programas sostenibles de intensificación para los recursos genéticos animales utilizados actualmente por los ganaderos?

- Tenemos suficientes conocimientos sobre el estado de las razas que hoy día carecen de valor para los ganaderos, y de las especies salvajes relacionadas con las razas domesticadas, para establecer la base de un Sistema de Alerta para los AnGR en peligro de extinción, y para identificar las oportunidades para el aprovechamiento de dichos recursos en el futuro?
- Estamos preparados para identificar situaciones en las que una respuesta de emergencia sería necesaria para impedir la pérdida de recursos genéticos animales en peligro?
- Se están aplicando, a niveles local, nacional, regional y mundial, políticas sostenibles para un aprovechamiento juicioso de los recursos genéticos animales?

La respuesta global a estas preguntas parece ser negativa, lo cual presenta un verdadero desafío para todos los implicados en el desarrollo agrario sostenible y para la seguridad alimentaria, sean agricultores, ganaderos, investigadores, educadores, políticos o administradores; especialmente si son miembros de la comunidad de consumidores; más aún si son responsables políticos, y sobre todo ¡si de nuestros hijos y los suyos se trata!.

Mediante la ayuda de los Gobiernos de sus estados miembros y otras organizaciones nacionales e internacionales, la FAO ha estado ayudando a países a establecer bases de datos de razas, en DAD-IS, y ha publicado dos ediciones de la Lista Mundial de Vigilancia para la diversidad de animales domésticos, la cual ofrece bastantes datos sobre los recursos genéticos animales en el mundo. Pero estos datos están lejos de proporcionar la información implicada en las preguntas anteriores, necesaria para planificar y poner

.....

en práctica una gestión firme y sostenible de los recursos genéticos animales de nuestros países.

La cuestión del Estado Mundial de los Recursos Genéticos Animales (SoW-AnGR) ahora, y su actualización periódica para hacer un seguimiento del desarrollo de AnGR, es actualmente tema de debate durante las sesiones por parte de las delegaciones nacionales de los países miembros de la FAO. Ha sido una cuestión importante en el orden del día de la Comisión de la FAO sobre los Recursos Genéticos para la Alimentación y la Agricultura que se convocó durante el mes de abril de 1999, con delegaciones proveniente de más de 140 países y organizaciones. Las delegaciones decidieron que la FAO debía liderar y coordinar el desarrollo de un informe impulsado por los países sobre el Estado Mundial de los Recursos Genéticos Animales a medida que vaya habiendo disponibilidad de fondos.

El informe SoW-AnGR establecerá específicamente cuál es la información fundamental en zonas principales: el estado

de la diversidad animal; la capacidad de un país de gestionar recursos genéticos animales; la situación actual, las metodologías y tecnologías disponibles para ayudar a los ganaderos, mejoradores y científicos para mejor describir, utilizar, desarrollar, conservar y acceder a los recursos genéticos animales y así contribuir a la seguridad alimentaria global y al desarrollo rural sostenible.

El informe SoW-AnGR establecerá los datos fundamentales críticos y la información necesaria para permitir la planificación y puesta en práctica de actividades de gestión de AnGR a niveles local, nacional, regional y global. También proporcionará la primera visión global y fiable para examinar los papeles potenciales para los recursos genéticos animales en el futuro y para ayudar a proyectar posibles desarrollos.

Finalmente, el primer informe SoW-AnGR ayudará a estimular la intensificación sostenible de la producción agroalimentaria en el amplio espectro de sistemas de producción en los países, en las regiones y en el mundo.

Los Editores

.....

Contribution to 25th Issue of AGRI from the Founder Editor

Animal Genetic Resources Information Bulletin (AGRI) was established jointly by FAO and UNEP in 1983 as a new global journal. Funds for the first four years were provided by UNEP while development, editorial and publication tasks were undertaken by FAO.

To understand the reason for the start of AGRI it may be recalled that a mere 15-20 years ago the conservation of domestic animals was a novel idea that, for many people, ran counter to the aims of livestock improvement. The concept of conservation as an important component of agricultural biodiversity surfaced at the first UN Conference on the Environment in Stockholm in 1972 as a result of which UNEP was created.

FAO worked closely with UNEP and in 1980 they convened a Joint FAO/UNEP Member Country Expert Consultation in Rome on "The conservation and management of animal genetic resources" under the independent chair of Dr. Helen Newton-Turner of Australia. This Consultation was not for experts in their own right. Participants were nominated by their governments and came as representatives of the member countries of FAO and UNEP. Thus, the recommendations were directed to FAO and UNEP from the member countries. I represented Canada at this Consultation.

Until that time, the thrust of FAO's support to governments for livestock improvement had been to increase production and productivity. A major component of the FAO and of some government bi-lateral programmes had been the provision of semen from Black and White cattle to developing countries. This practice was later judged to be misplaced from a production point of view

and was discontinued. Further, the practice was diluting the gene pools of indigenous cattle.

Little was known about the numbers, condition and genetic parameters of indigenous breeds at that time in either developed or developing countries. Documentation was sparse, often unpublished, poorly classified and inaccessible. The 1980 Consultation recommended that FAO and UNEP should co-finance and that FAO should establish:

1. Classification criteria for endangered breeds;
2. Technical methods and standards for live and cryogenic conservation;
3. Methods of conservation;
4. Regional data and gene banks;
5. Comprehensive documentation of livestock breeds in China and the USSR;
6. Training in conservation methods;
7. A series of technical publications on animal conservation;
8. A Newsletter.

Based upon these recommendations, UNEP provided funds and FAO undertook the tasks by adding them to the existing livestock improvement responsibilities of the Senior Officer for Animal Breeding in the Animal Production and Health Division (AGA) without at that time, it might be added, any increase in human resources. I was appointed to the enlarged position in 1982, thereby becoming the Founder Editor of AGRI and serving until 1990. All the recommendations of the 1980 Consultation were either completed or launched during the 1980s.

The Newsletter was intended to support the increasing number of individuals, NGOs and a few governments throughout the world that were beginning to work on the conservation of animal genetic resources. These people were isolated, frequently lacked technical direction, experience and funds. The Consultation urged that the Newsletter should be sent free of cost to all who were interested. There were two special aims:

1. To provide all those involved in Animal Genetic Resources Conservation and Management with information;
2. To offer a specialized journal for publication of papers on Animal Genetic Resources Conservation and Management, since this topic was often viewed as irrelevant at the time by some of the major animal science journals.

25 issues and 16 years later it seems strange that this major field of animal genetics should have been so late in developing. The Newsletter was enthusiastically supported from the start by a growing number of contributors and readers and, under the present Editorship, has continued to expand and to contribute meaningfully to the conservation of animal genetic resources. In my view, it has been a significant factor in changing the perspectives of scientists, governments and development practitioners to include both livestock utilization and conservation as essential partners in livestock improvement now and in the future.

Dr. John Hodges
Founder Editor of AGRI

Past, present and future

Is issue 25 of a journal of any particular importance? At least in this case it means 16 years of interactions and some 2 000 pages of inputs. It all started at a time when people interested in the conservation of Animal Genetic Resources (AGR) were *a minute minority*, euphemistically speaking *a species on its way to extinction*. Today, seven years after the Rio de Janeiro Summit, the terms *AGR* and *Biodiversity* are truly *a la mode*.

The extraordinary development of science and technology these past decades was particularly marked by the progress of biotechnologies, the extent of their actual application and the potential and dangers they could introduce in the future. Thus, the many crises we had to address these past couple of years oblige us to renewed attention in the relationship Biosciences-Society. Ethics are now again *news* and questioning the advances of science, the benefits and implications, the successes and the challenges is today integral part of any daily exercise, actions and reactions. In this complex, new but fundamental thinking, biodiversity becomes a master-card.

It is known that indigenous AGRs are well adapted to the marginal regions and extensive difficult grazing territories of the world and are mostly linked to the manufacture and availability of traditional products of animal origin. Their conservation must thus be seen as a mission of local and regional collective interest. They must be fully incorporated as one of the decisive factors of the prevailing livestock production systems.

Having said that and having been directly and actively involved with AGR conservation and use since 1960, I can dare wonder if we have not been in recent years overstating sometimes the *specific importance* of this field

of animal agriculture. AGR should be dealt within the overall context of the Animal Production Sector, including Livestock Systems and Global Rural Biodiversity, not as a mystified study field apart.

The Animal Genetic Resources Information Bulletin (AGRI) launched in the early 80s by John Hodges, stopped appearing in the late 80s because of cutbacks in the FAO Regular Programme Budget. As soon as this problem could be solved, around 1991 with UNEP's further meaningful support, it seemed indispensable to resume this publication medium through which colleagues from around the world and in particular those from developing countries could share their information and know-how on farm animal breeds without the constraints of a too sharp formalism. This is also true for the great number of interested people who are not directly concerned with research, such as breed associations, NGOs, INGOs and hobby breeders.

The new AGRI was designed and developed as a support for the circulation of results, reviews, point of views and opinions that a strictly scientific publication would not publish.

It is necessary to underline, at this stage, that this bulletin is one of the most cost-effective venues of its sort and is partially run on a voluntary basis.

There are three distinct periods in AGRI's life (figure 1). The first one can be called the *birth and first difficult steps* (1983-1987) followed by two years of no activity due to financial constraints and the departure of the founder and first editor, John Hodges. The second period, (1991 to 1995; Co-Editors: Daniel Chupin and Jean Boyazoglu) corresponds to a time of *growth and worldwide*

Figure 1. Number of pages published per year.

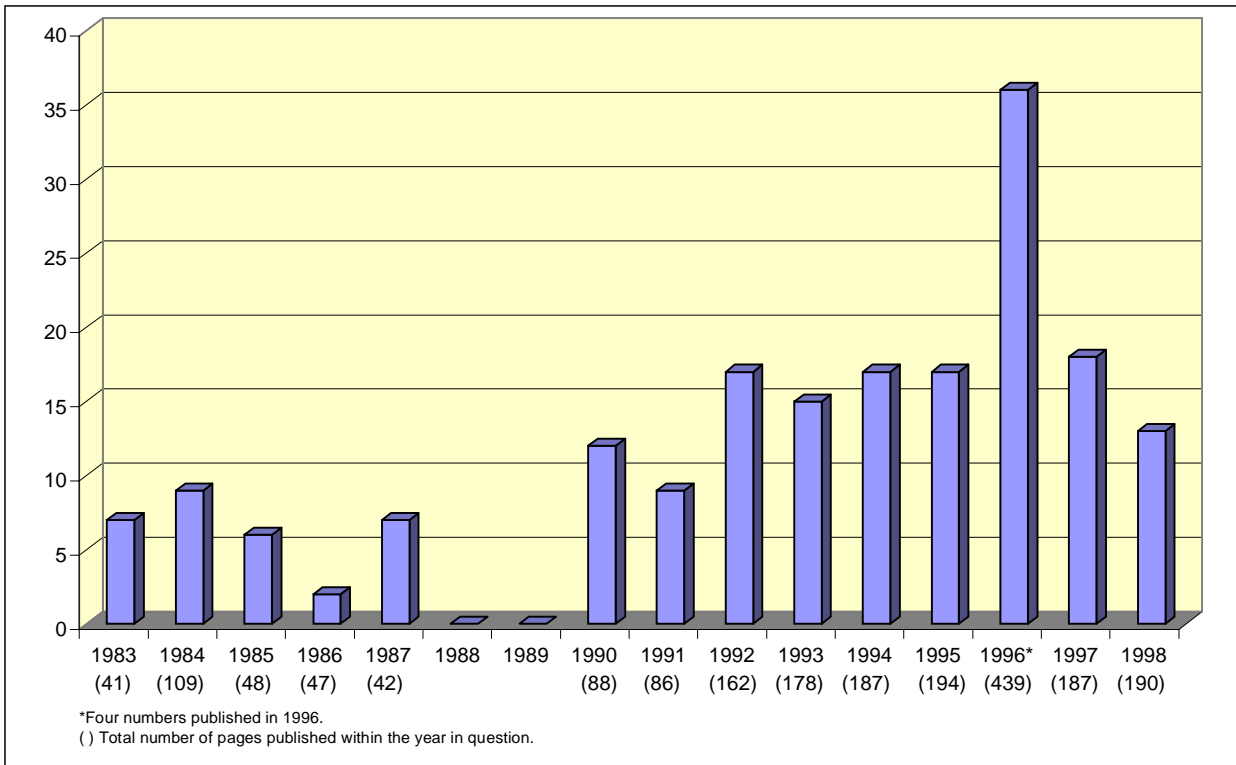


Figure 2. Number of articles published from different regions of the world.

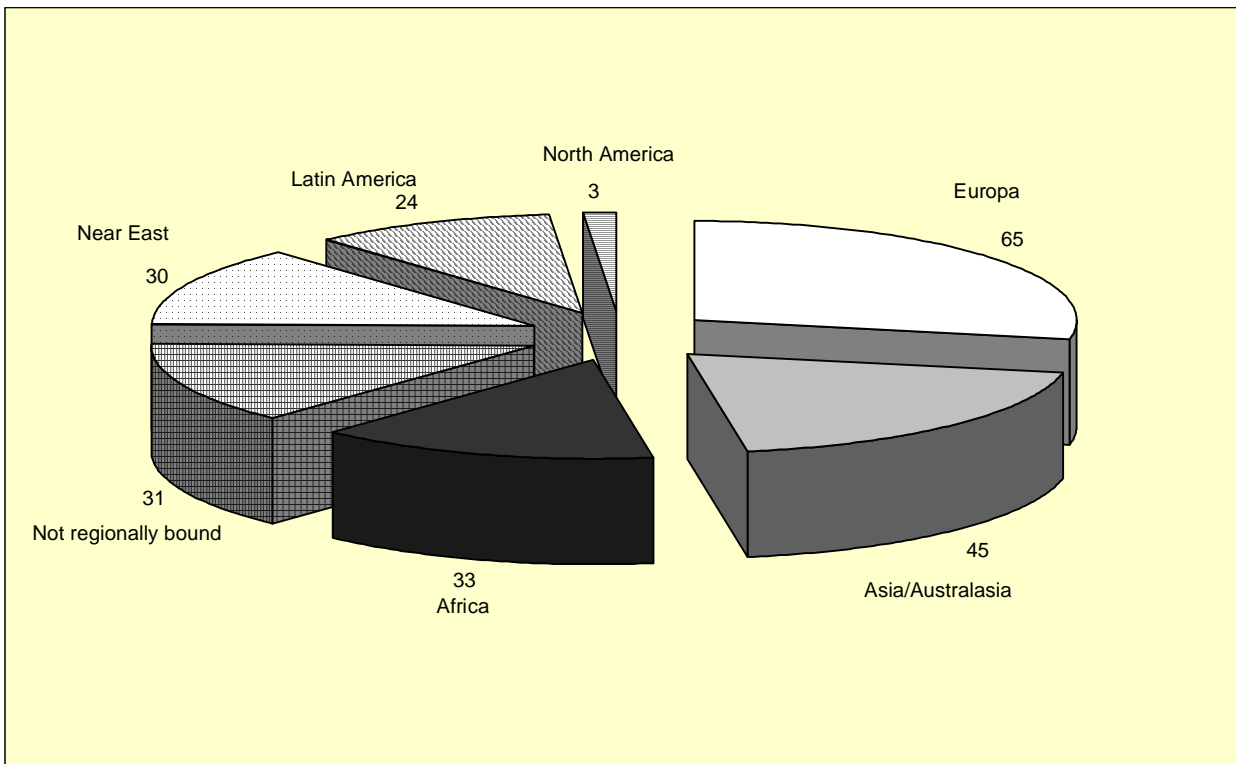
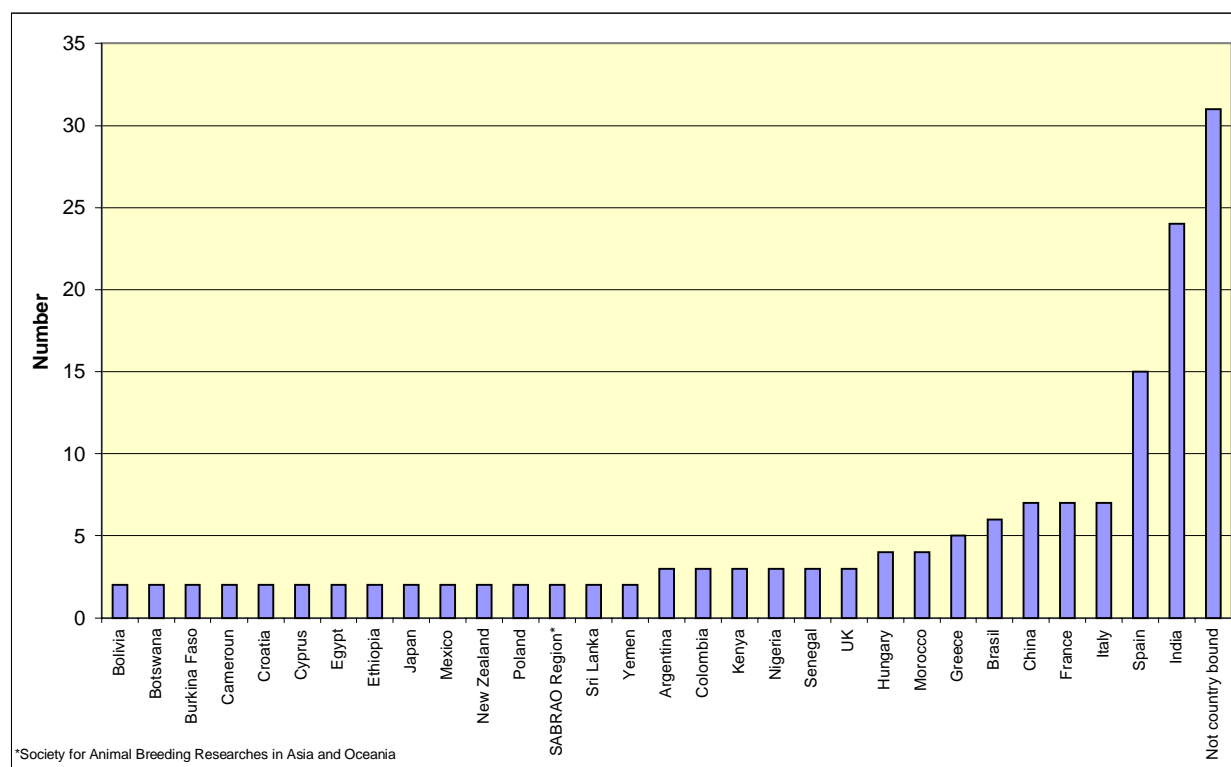


Figure 3. Countries with more than one paper published in AGRI.



recognition of the bulletin's value. The third period (1996 to date; Co-Editors: Salah Galal and Jean Boyazoglu) is one of *enhancing the journal's status, important changes in format and presentation, much more selective choice of the material published and the electronic publication on the Internet and in a CD ROM*. This period also correspond to the development of FAO's Global Strategy. While in the early years we had to struggle to obtain manuscripts, and in many cases had to rewrite them completely to meet linguistic and technical standards and needs, today we can be more discriminative: from 61 papers received since AGRI no. 19, 45 manuscripts were published or accepted for publication, while 5 are pending; thus a rejection rate of around 20%.

We mostly try to publish two issues per year, though there are some exceptions such as the year 1996 with four issues and 439 pages published; we had to find extra funding to meet the need of clearing the backlog of manuscripts. The average number of pages published per issue was around 50

in the 80s, moving up to 100 in the 90s. The issues dated 1990 correspond to the backlog of articles that were received during the 1987-1989 period; they were edited for publication in early 1991.

The 201 articles published in the 25 issues (figure 2) originated from Europe(33%), from Asia/ Australasia (24%), from Africa (16%), from Latin America (12%), from Near East (4%) and from North America (2%); the remaining articles (19%) did not concern a specific region of the world. In particular, the geographic origin (country/subregion) of the material analysed in the published papers is given in figure 3 and table 1. The distribution of the articles shows that papers concerning the main ruminants (cattle, sheep and goats) and horses are in the great majority (figure 4) although the interest goes from pigeons to rabbits and from donkeys to yak and buffaloes.

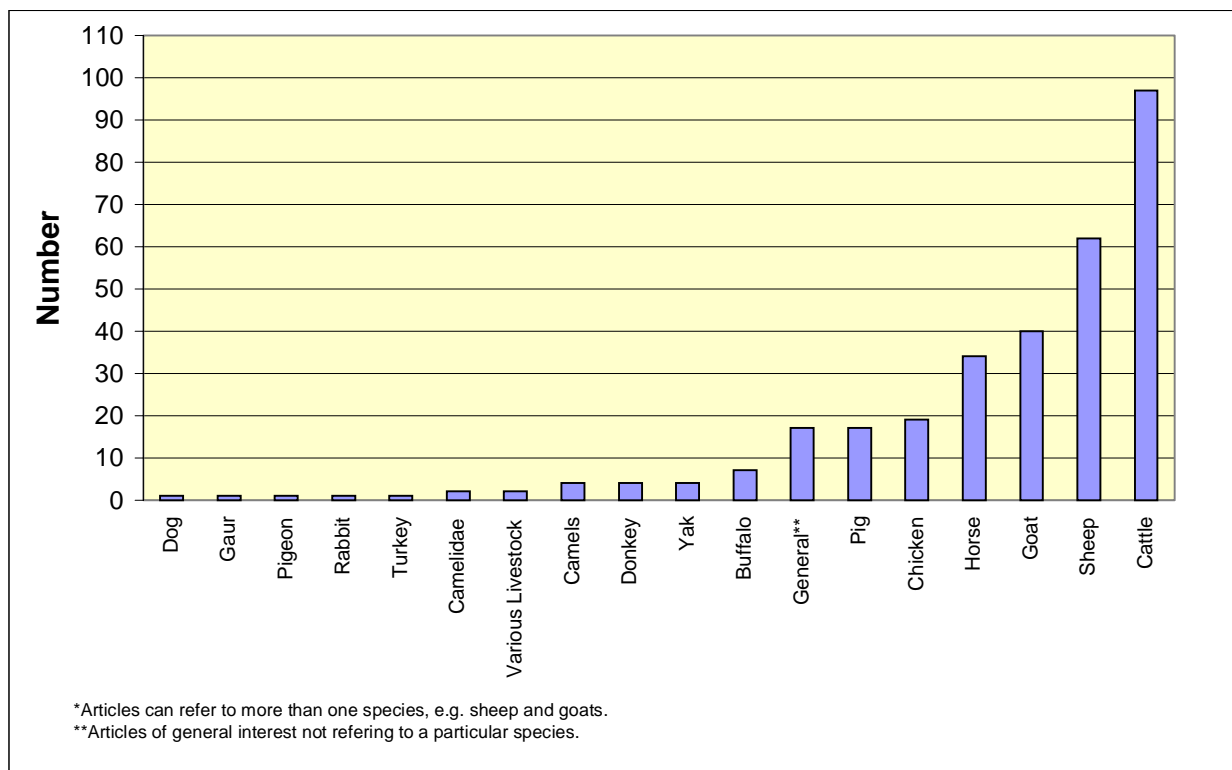
In 1997 we launched a questionnaire (see annex) to update our database and individual circulation list; this excluded libraries and

Table 1. Countries with only one paper published.

| | | | |
|-----------|---------------|---|-------------------------------------|
| Albania | Ivory Coast | Togo | European Nordic Countries* |
| Algeria | Korea | Uganda | Latin America* |
| Benin | Latin America | Ukraine | Mediterranean basin* |
| Bhutan | Malawi | Uruguay | North America* |
| Bulgaria | Malta | USA | Sub-Saharan francophone countries* |
| Burundi | Mauritania | USSR (Russia Fed.) | West and Central African countries* |
| Canada | Peru | Vietnam | |
| Chad | Russia | Yugoslavia | |
| Cile | Saudi Arabia | Zanzibar | |
| Cina | Scandinavia | Central and Eastern European countries* | |
| Denmark | Somalia | Central Asia* | |
| Estonia | South Africa | Europe* | |
| Germany | South America | | |
| Guatemala | Tanzania | | |

*Papers referring to subregions.

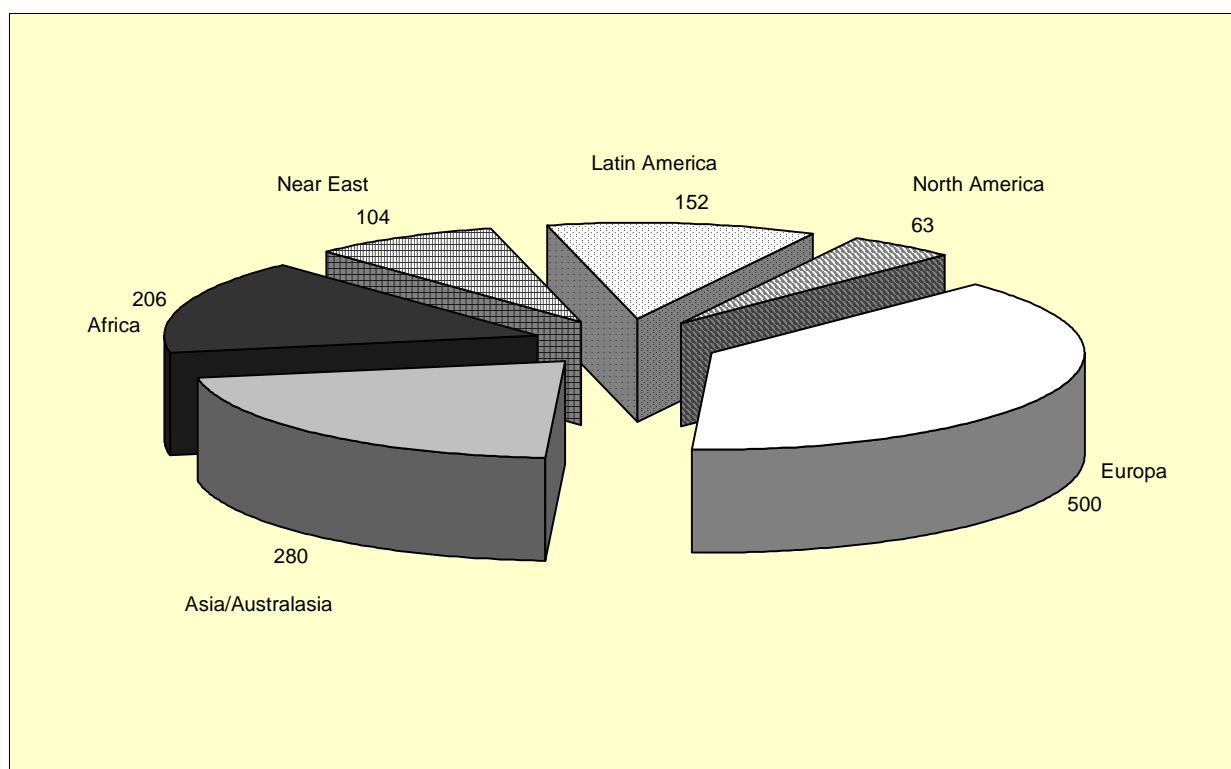
Figure 4. Number of articles published in AGRI, per species*.



institutions. We received 1 305 valid answers with an arbitrary division of 1/2 English, 1/4 French and 1/4 Spanish speaking *readers*. The geographic distribution of the readership that correctly answered the questionnaire (figure 5) was just over 38% from Europe (many from Eastern and Southern Europe), 21% from Asia/Australasia, 16% from Africa, 12% from Latin America, 8% from the Near East; less than 5% of the readership being from North America. Thus, a majority of the individual readers who personally receive AGRI are from less developed countries or countries in transition. Italy (table 2) has the largest national individual readership (136) followed by India (89) and Spain (65). The readers of AGRI are shown to be mainly involved with teaching, training and research (figure 6) while the main *fields* of interest are small ruminants (sheep and goats) and the cattle sector, followed by genetics and breeding (figure 7).

While defending the idea that this bulletin must accept all types of articles giving information on domesticated animal breeds, we realised very soon the need to standardise as much as possible the form and presentation to help the authors present better prepared articles and the readers to have more comprehensible material while giving the minimum necessary information relating to the breeds and line comparisons, within the framework of the locally prevailing production systems. Since the beginning we encouraged authors to include a maximum of photos and graphics which are more explicit than long discourse; maps, even sketchy ones, allowing a better localisation of the population described and its distribution than listing geographic names. This approach was received very favourably as shown by the growing number of interesting manuscripts received for the publication. This allowed to assure the regular publication of AGRI in the

Figure 5. Number of AGRI subscribers* in the different regions of the world.



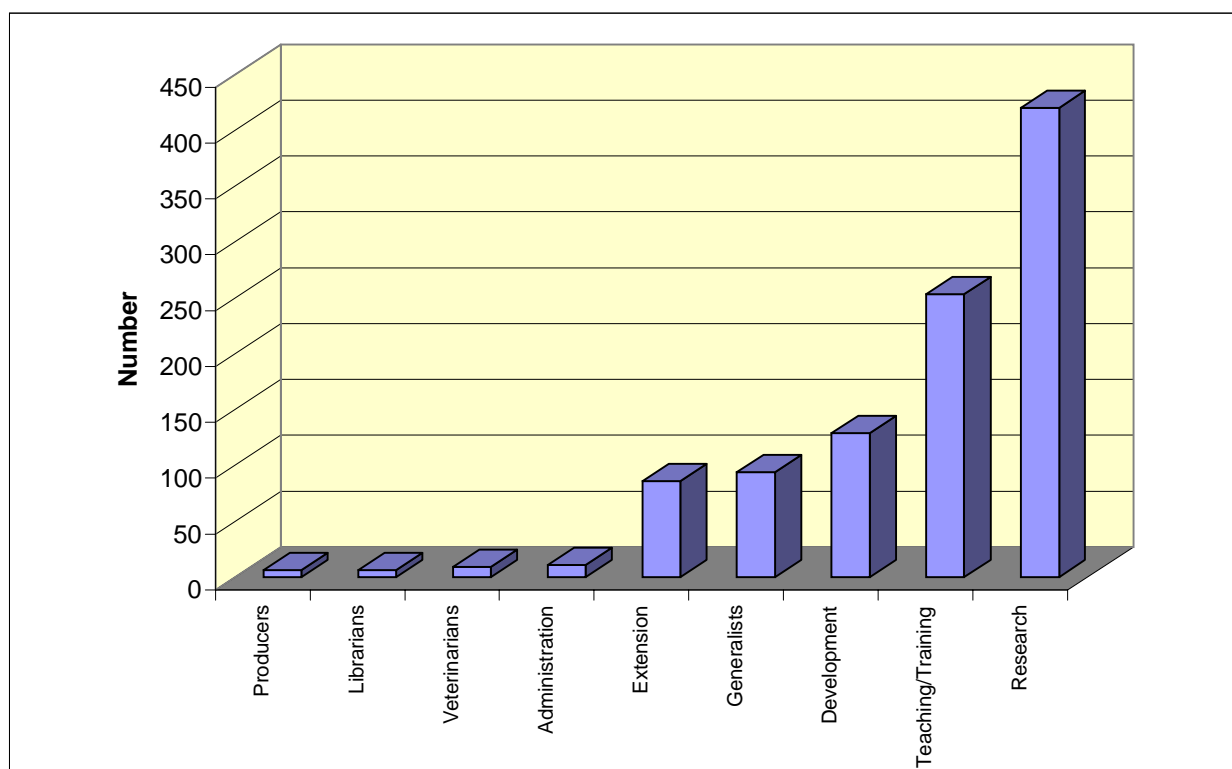
*This figure refers only to those *individual readers* who answered the 1997/1998 questionnaire.

Table 2. Individual country readership distribution of AGRI.

| Class | Country |
|---------------|--|
| 1-5 readers | Albania, Algeria, Angola, Antigua and Barb, Austria, Bahrain, Barbados, Benin, Bermuda, Bhutan, Bolivia, Bosnia and Herzegovina, Botswana, Bulgaria, Burundi, Cambodia, Cameroon, Cape Verde, Central African, Chad, Taiwan, Congo, Congo, Cook Islands, Costa Rica, Croatia, Cyprus, Czech Republic, Dominican Republic, Ecuador, El Salvador, Equatorial Guinea, Eritrea, Estonia, Falkland Islands, Fiji, Finland, French Guyana, Gambia, Guadeloupe (France), Guam (USA), Guatemala, Guinea, Guinea-Bissau, Guyana, Haiti, Honduras, Hungary, Iceland, Iran, Islamic Re, Ireland, Israel, Jamaica, Jordan, Kazakhstan, Korea, Kuwait, Kyrgyz Republic, Laos, Latvia, Lebanon, Lesotho, Liberia, Libya, Lithuania, Luxembourg, Macedonia, Madagascar, Maldives, Malta, Mauritania, Mauritius, Moldavia, Mongolia, Mozambique, Myanmar, Namibia, Netherlands Antilles, New Zealand, Nicaragua, Niger, Oman, Papua New Guinea, Puerto Rico (USA, Romania, Russian Federation, Rwanda, Samoa, Saudi Arabia Kingdom, Sierra Leone, Slovakia, Slovenia, Solomon Islands, Sri Lanka, Swaziland, Tanzania, Togo, Trinidad and Tobago, Tunisia, Uganda, Ukraine, United Arab Emirates, Vanuatu, Venezuela, Vietnam, Yemen, Yugoslavia, Zambia |
| 6-10 readers | Argentina, Bangladesh, Burkina Faso, Ivory Coast, Cuba, Denmark, Ethiopia, Ghana, Greece, Iraq, Malawi, Malaysia, Mali, Morocco, Nepal, Norway, Poland, Portugal, South Africa, Syria, Turkey, Uruguay, Zimbabwe |
| 11-15 readers | Colombia, Indonesia, Japan, Kenya, Peru, Senegal, Sudan, Sweden, Thailand |
| 16-20 readers | Australia, Chile, Mexico, Nigeria, Pakistan, Switzerland |
| 21-30 readers | Belgium, Canada, China, Netherlands, Philippines |
| 31-42 readers | Brazil, Egypt, France, Germany, United Kingdom, United States of America |
| 65 readers | Spain |
| 89 readers | India |
| 136 readers | Italy |

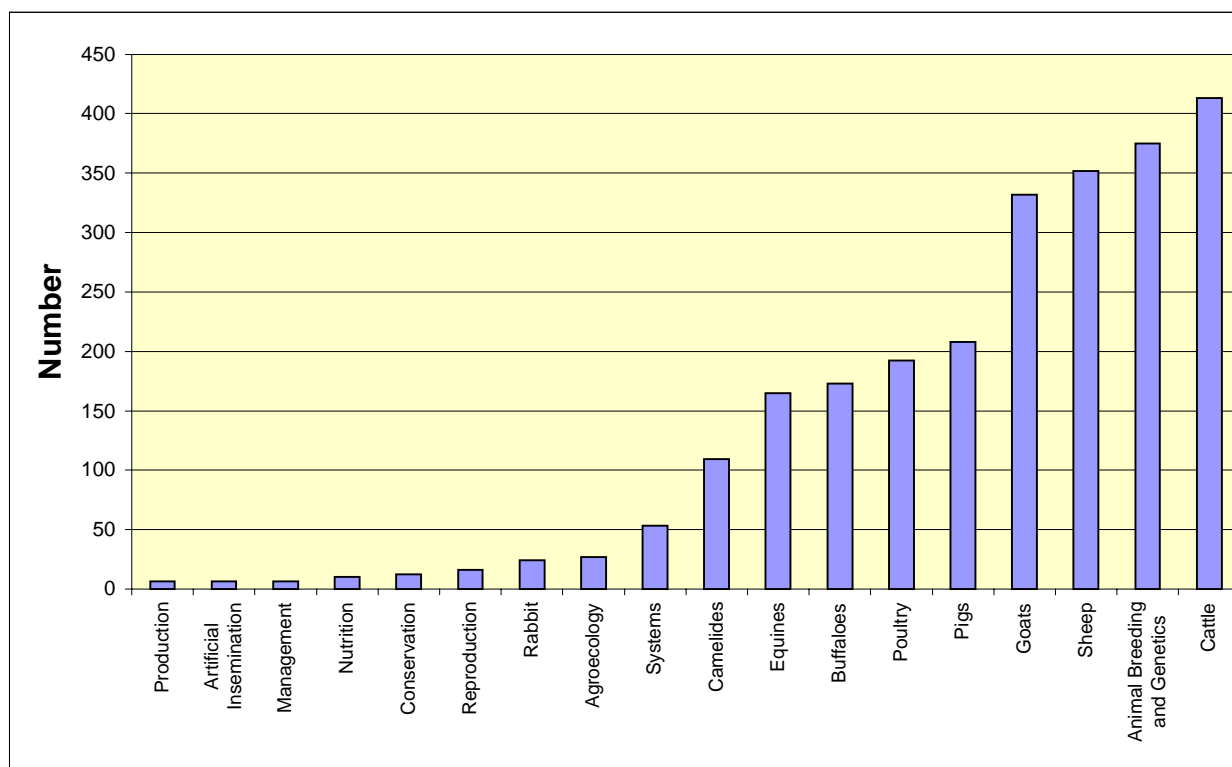
This table refers only to those *individual readers* who answered the 1997/1998 questionnaire, (1305 valid answers received).

Figure 6. Main responsibilities of AGRI readers*.



*Many readers may have two or more responsibilities, e.g. research and teaching.

Figure 7. Main interests of AGRI readers*.



*Many readers may have two or more interests, e.g. sheep and goats or reproduction and pigs.

.....

years 1992-1996 and also considerably increased the number of pages per issue. Evidently the regular flow of AGRI depends and will depend even more in the future on the availability of the necessary funds and the collaboration and voluntary input of all persons involved with the publication.

As the one that has been the longest involved voluntarily with the journal, I should underline that, in my humble opinion, a very important feature of AGRI is that it is one of the few remaining FAO periodicals offered free of cost to all those

requesting it in writing, in line with the strong recommendations made by the Country Expert Consultation in 1980. In this respect, the very meaningful financial and moral support of UNEP must be acknowledged. It is important also to refer here to the loyalty of this journal's readership, the great majority of which is in the developing world. The number of answers received (more than 1 500 of which 1 305 correctly filled in) to the 1997-98 questionnaire is an impressive showing of interest in the journal; over 75% of the questionnaires circulated to the global individual readership (some 2 000 addresses) has been duly filled-in and returned!

Jean Boyazoglu
Co-Editor of AGRI

.....



Animal Genetic Resources Information



S. Galal and J. Boyazoglu (Editors)

FAO, Animal Production and Health Division

Via delle Terme di Caracalla, 00100 Rome, Italy

Room C-579; tel.: +39-6-57056139; fax.: +39-6-57053927; Email: <Salah.Galal@fao.org>

Dear reader,

We are in the process of updating the AGRI mailing list. If you want to continue receiving AGRI, please complete and sign this form and mail/fax it to the address indicated above.

Name:
First Last Middle

Title: Ms. • Professor • Dr. • Mr. •

Address:
City Country Postal code

Fax: Telephone :
Number Country code / City code / Number Country code / City code /

Email:

Occupation: • Research • Teaching/training •
Development • Extension • Others (pls. specify)
.....

Specialization: • Animal breeding and genetics • Generalist • Systems
• Others (pls. specify)

Species of interest: • Buffalo • Camelides • Cattle • Equines (horse, ass, etc.)
• Goat • Sheep • Pig • Poultry
Others (specify):.....

Signature.....

Date.....



The state of African cattle genetic resources I. Classification framework and identification of threatened and extinct breeds

J.E.O. Rege

International Livestock Research Institute (ILRI), P.O. Box 5689, Addis Ababa, Ethiopia

Summary

A field and literature survey was conducted to determine the status of cattle genetic resources of sub-Saharan Africa and to identify cattle breeds at risk and those which may have become extinct over the last century. This paper - in two parts - summarises preliminary results of the survey. The survey revealed that sub-Saharan Africa is home to a total of 145 cattle breeds/strains comprising two taurine Longhorns, 15 taurine Shorthorns, 75 zebu (*Bos indicus*), 30 sanga, eight zenga (zebu-sanga), nine breeds derived from interbreeding of indigenous breeds/strains located in close proximity to each other, and six systematically created composite breeds. Out of the 145 breeds identified from the survey, 47 (about 32%) were considered to be at risk of extinction. Risk categories used were: Critical (most severe), Endangered, Vulnerable, and Rare (least severe). Of the breeds identified to be at risk of extinction, six were in the "Rare" category, 10 were "Vulnerable", another 10 were "Endangered" and 15 were in the "Critical" category. A total of 22 breeds (about 13%) previously recognised in the continent have become extinct in the last century. This number excludes some populations which have lost their individual identity due to admixtures involving two or more originally distinct breeds.

Resumen

Se llevó a cabo un estudio bibliográfico y de campo para determinar cuál era el estado de los recursos genéticos ganaderos del Africa subsahariana y para identificar las razas ganaderas en peligro así como aquellas que se hayan podido extinguir durante el último siglo. Este artículo, dividido en dos partes, resume los resultados preliminares del estudio. El estudio reveló que el Africa subsahariana alberga un total de 145 razas bovinas/estirpes, comprendiendo dos razas taurinas Longhorn, 15 Shorthorn, 75 zebu (*Bos indicus*), 30 sanga, 8 zenga (zebu-sanga), 9 razas derivadas de cruzamientos entre razas autóctonas y estirpes estrechamente relacionados y 6 razas compuestas creadas de forma sistemática.

De los 145 razas identificadas en el estudio, 47 (alrededor del 32%) se consideraron en peligro de extinción, las categorías de riesgo utilizadas siendo las siguientes: crítica (más severa), en peligro, vulnerable, y rara (menos severa). De las razas identificadas como en peligro de extinción, seis se encontraban en la categoría "rara", 10 eran "vulnerables", otras 10 estaban "amenazadas" y 15 estaban en la categoría "crítica". Un total de 22 razas (alrededor del 13%) previamente reconocidas en el continente se han extinguido en el último siglo. Esta cifra excluye algunas poblaciones que han perdido su identidad individual debido a mezclas entre dos razas originalmente distintas.

Key Words: *Breed classification, Cattle, Endangered breeds, Extinct breeds, Phenotypic diversity, Sub-Saharan Africa.*

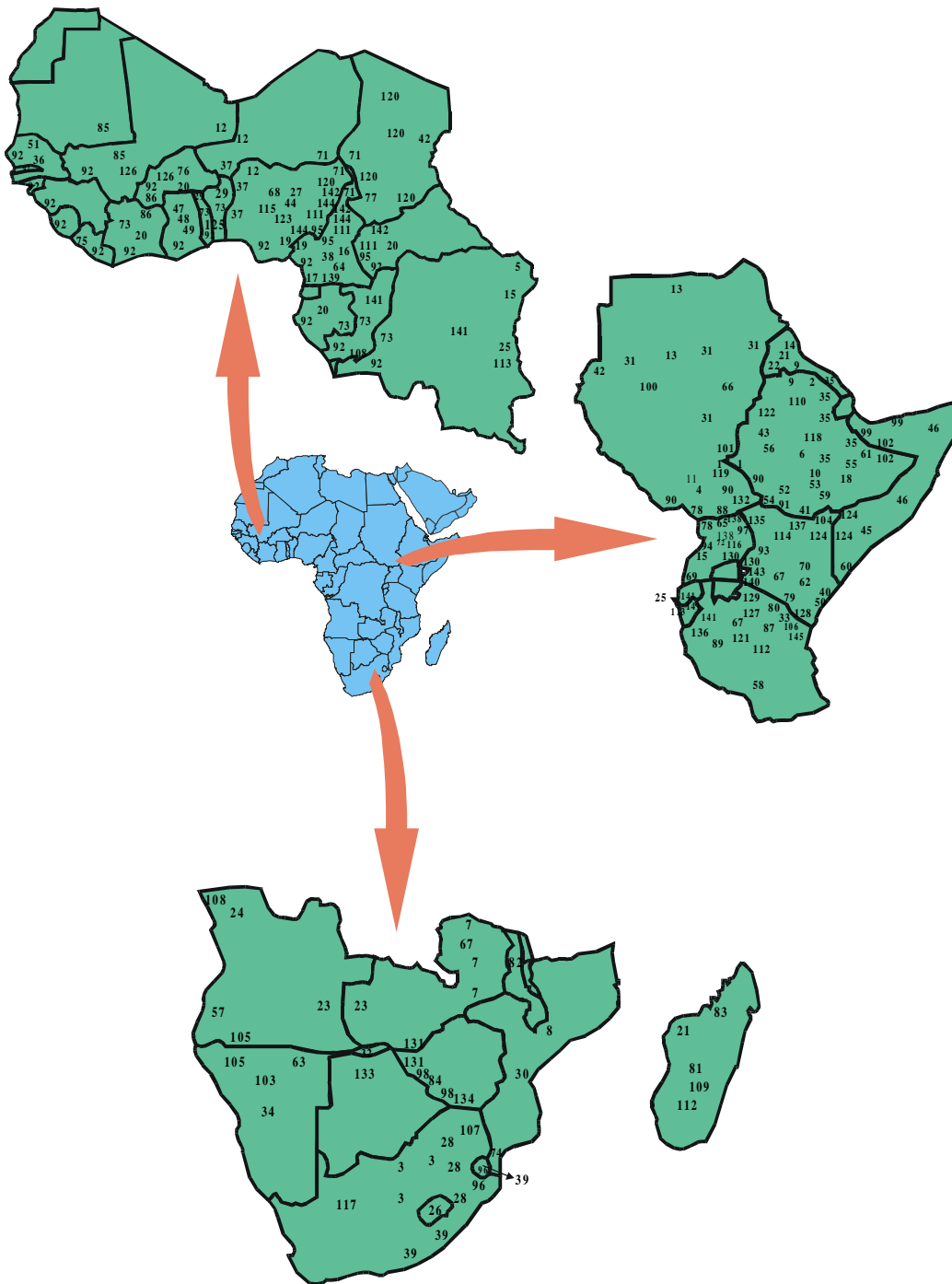


Figure 1. Distribution of cattle breeds in some African countries

Introduction

The origins of indigenous cattle of Africa still remain uncertain despite available archaeological, anthropological and historical evidence (Epstein and Mason, 1984; Blench,

1993). It is generally accepted that the African cattle populations arose from three main phases of introduction from Asia through the Nile valley in Egypt or via the Horn of Africa.

Subsequent migrations led to dense populations of cattle in the East African highlands, around the present-day Ethiopia

| List of breed codes reported in figure 1 | | |
|---|----------------------------|-------------------------------------|
| 1. Abigar | 48. Ghana Dwarf Muturu | 98. Nkone |
| 2. Adwa | 49. Ghana Shorthorn | 99. North Somali Zebu |
| 3. Afrikaner | 50. Giriama | 100. Nuba Mountain Zebu |
| 4. Aliab Dinka | 51. Gobra | 101. Nuer |
| 5. Alur (Nioka, Blukwa) | 52. Goffa | 102. Ogaden |
| 6. Ambo | 53. Guraghe | 103. Okavango |
| 7. Angoni | 54. Hammer | 104. Orma Boran |
| 8. Angonia (Angone) | 55. Harar | 105. Ovambo |
| 9. Arado | 56. Horro | 106. Pare |
| 10. Arsi | 57. Humbi | 107. Pedi |
| 11. Aweli Dinka | 58. Iringa Red | 108. Porto-Amboim |
| 12. Azaouak (Tuareg) | 59. Jem-Jem | 109. Rana (Omby Rana) |
| 13. Baggara | 60. Jiddu | 110. Raya-Azebo |
| 14. Baherie | 61. Jijiga | 111. Red Fulani |
| 15. Bahima | 62. Kamba | 112. Renitelo |
| 16. Bakosi | 63. Kaokoveld | 113. Ruzizi |
| 17. Bakweri | 64. Kapsiki | 114. Sabmuru |
| 18. Bale | 65. Karamajong zebu | 115. Savanna Muturu |
| 19. Banyo | 66. Kenana | 116. Serere |
| 20. Baoulé | 67. Kenya (Improved) Boran | 117. Shangan |
| 21. Baria | 68. Keteku | 118. Sheko |
| 22. Barka | 69. Kigezi | 119. Shilluk |
| 23. Barotse | 70. Kikuyu | 120. Shuwa |
| 24. Barra do Cuanzo | 71. Kuri | 121. Singida White |
| 25. Bashi | 72. Kyoga | 122. Smada |
| 26. Basuto | 73. Lagune | 123. Sokoto |
| 27. Biu | 74. Landim | 124. Somali Boran |
| 28. Bonsmara | 75. Liberia Dwarf Muturu | 125. Somba |
| 29. Borgou | 76. Lobi | 126. Sudanese Fulani (Peul Zebu) |
| 30. Bovines of Tete | 77. Logone | 127. Sukuma (Tinde) |
| 31. Butana | 78. Lugware | 128. Taita |
| 32. Caprivi | 79. Maasai (Kenya) | 129. Tarime (Shashi) |
| 33. Chagga (Wachagga) | 80. Maasai (Tanzania) | 130. Teso |
| 34. Damara (Herero) | 81. Madagascar zebu | 131. Tonga |
| 35. Danakil | 82. Malawi Zebu | 132. Toposa |
| 36. Djakore | 83. Manjan'i Boina | 133. Tswana |
| 37. Djelli (Diali) | 84. Mashona | 134. Tuli |
| 38. Doayo (Namichi) | 85. Maure | 135. Turkana |
| 39. Drakensberger | 86. Méré | 136. Ugogo Grey |
| 40. Duruma | 87. Mkalama Dun | 137. Unimproved Boran (Borana) |
| 41. Ethiopian Boran (Borana) | 88. Mongolla | 138. Usuk |
| 42. Fellata (Bororo) | 89. Mpwapwa | 139. Wakwa |
| 43. Fogera | 90. Murle | 140. Watende |
| 44. Forest Muturu | 91. Mursi | 141. Watusi |
| 45. Garre (Gherra) | 92. N'Dama | 142. White Fulani |
| 46. Gasara | 93. Nandi | 143. Winam (Kavirondo) |
| 47. Ghana "sanga" | 94. Nganda | 144. Yola |
| | 95. Ngaundere | 145. Zanzibar Zebu |
| | 96. Nguni | |
| | 97. Nkedi | |



Figure 2. Kuri cattle.

and Kenya. The humpless Longhorns arrived in the continent approximately 5000 BC, followed by the humpless Shorthorns around 2500 BC. Humpless Shorthorn cattle were historically distributed in almost all ecological zones. Isolated populations were recorded in Sudan and Central Tanzania in historical times and remnant populations still exist in Ethiopia. The humped zebu first arrived about 1500 BC and later in large numbers around 670 AD (Eptsein, 1971). The first zebu cattle in Ethiopia are thought to have been brought through Somalia by Semitic people from Arabia, and their subsequent interbreeding with the taurine Longhorns are considered to have produced the present-day sanga cattle, although recent (unpublished) evidence from ILRI work questions the "crossbred origin" of the sanga. The second introduction of zebu cattle led to a major replacement of sanga cattle from most parts of eastern Africa, emergence of intermediate zebu-sanga breeds in some cases, and separate evolution into different strains in the diverse ecological zones. The zebu cattle further spread

westwards and southwards to become the major group of cattle in the region bordered by latitude 20°N in northern Sudan, in the north, the western rain-forest barrier in the west and the Zambezi river, latitude 15°S, in the south.

Present-day African cattle can be classified into four broad categories: the humpless *Bos taurus*, widely distributed in West and Central Africa; the humped *Bos indicus* (zebu), distributed widely in eastern and the dry parts of West Africa; the sanga, found mainly in eastern and southern Africa; and sanga x zebu types ("zenga") found in eastern Africa. The taurine (humpless) type has two groups, Longhorns (*B. taurus longifrons*) and Shorthorns (*B. taurus brachyceros*), both of which are restricted to West and Central Africa.

In addition to the four broad groups of African cattle, there are more recent derivatives that have resulted either as a consequence of close proximity of two or more indigenous populations, sometimes accelerated by deliberate crossing (e.g. the Borgou, Méré and Ghana Sanga of West

Africa), or as a product of efforts to create composite commercial breeds (e.g. the Bonsmara of South Africa and the Mpwapwa of Tanzania).

There is very little information on the diversity in indigenous African livestock populations, both at phenotypic and genetic levels. Indeed, not even the number of breeds of the various species is known. Working estimates are: 100-150 indigenous cattle, 50-60 sheep and 45-50 goat breeds in sub Saharan Africa (Rege, 1998). Additionally, with very few exceptions, there is no data on the population sizes of the different breeds and their status, i.e. whether their numbers are decreasing, increasing or stable.

About the Survey

Starting in 1992, the International Livestock Centre for Africa (ILCA), now the International Livestock Research Institute (ILRI), initiated activities aimed at determining the status of, and compiling information on characteristics of, indigenous African domestic ruminant livestock, specifically cattle, sheep and goats. These activities consisted of:

- 1) sending out mail questionnaires to scientists in national programmes;
- 2) field visits to assess the status of individual breeds/populations identified from questionnaires;
- 3) requests for specific information from collaborating scientists;
- 4) conduct of rapid surveys in selected countries or of selected breeds/populations within a country; and
- 5) review of literature, especially the non-conventional (grey) publications such as annual reports, proceedings of national meetings (conferences, seminars), research project reports, etc.

In addition to these sources, a substantial amount of information was obtained from field expeditions conducted during a continent-wide exercise to collect blood samples for molecular characterisation, to quantify between- and within-population genetic diversity at the DNA level, which is part of ILRI's overall programme on animal

genetic resources. Field visits included interviews with government officials, national scientists and farmers. The aim of the interviews was to obtain, for each breed/population, indications of:

- 1) major uses;
- 2) trends (decreasing, increasing or stable) as perceived by local communities, extension personnel and scientists;
- 3) possible reasons for the perceived trends;
- 4) phenotypic description of the breed; and
- 5) levels of performance, where available.

Whereas the information on uses, trends and determinants of trends, as well as breed description, was mainly obtained from field visits and, to a lesser extent, mail questionnaires, most of the quantitative data on performance was obtained from comprehensive reviews of literature. Data on physical measurements (e.g. withers height) were, in a few cases, obtained by measuring representative animals on-farm or on-station during the field visits or through collaboration with national scientists.

This survey represents a first attempt to assess the status of the ruminant genetic resources of sub-Saharan Africa. It does not constitute the level of assessment required to make decisions on conservation and use. However, it provides a basis for additional, targeted surveys. As this was a very broad-brush survey, the information is incomplete, and may even contradict facts unknown to us. We are aware of this but feel the publication of these results will provoke action, particularly at national levels, that will help improve the information base on domestic African breeds/strains. In the meantime, efforts are underway to conduct in-depth breed surveys in pilot countries. One such survey, in Ghana, is already completed and will be published shortly. Additionally, activities on molecular characterisation of African cattle are well underway. Similar studies have also been initiated for sheep and goats.

Figure 1 shows the geographic distribution of cattle breeds in some African countries

Breed Groups and Clusters

As has been alluded to, the survey recognised four broad categories of African cattle: the humpless *Bos taurus*; the humped *Bos indicus* or zebu; the sanga; the zenga (sanga x zebu derivatives). Fifth and sixth categories included were the “recent derivatives” created from interbreeding amongst indigenous breeds, and the composite breeds developed from systematic crossing of two or more breeds for specific purposes. The taurine category was further subdivided into two groups – the Longhorns and the Shorthorns - while the zebu and the sanga were divided into several groups and clusters. A group is defined here as a collection of breeds or strains considered to have a common ancestry but not necessarily inhabiting the same geographical area. Cluster, on the other hand, is used to refer to a collection of breeds or strains which belong to the same category and, usually, group, and occupy the same geographical area e.g. a country and/or a defined ecozone within one or more countries. Thus, the sub-category “Small East African Zebu” was divided into groups represented by the Abyssinian Shorthorned Zebu, The Somali Zebu, The Teso Zebu, etc. Examples of clusters were the Kenya cluster of zebus, the Tanzania cluster of zebus and the Ovambo and southwestern cluster of sangas. Although the motivation behind the “clustering” was purely presentational convenience, an attempt was made to lump together only those breeds and strains which, because of geographical proximity have had a high likelihood of interbreeding and hence may have more common genetic background than other populations with similar evolutionary history but which have been more isolated. The term group is also used to refer to such broad categories as the humpless Longhorn and the humpless Shorthorn, members of which are considered to be genetically closely related and have not undergone the same differentiation as seen in the zebu or, to some extent, the sanga.

This paper, the first in a two-part series, provides a framework for the classification of African cattle breeds/strains on the bases of historical evidence, phenotypic appearance and geographical location, and identifies populations that may be threatened with extinction and those which may have become extinct in the last 100 years or so. The second paper (Rege and Tawah, 1999) describes the physical characteristics and status of each of the existing breeds in terms of breed development as well as their distribution, main uses and known or speculated evolutionary relationships among breeds.

Figures from 2 to 5 show some cattle breeds.

Classification of Breeds

The taurine cattle

The humpless Shorthorns and Longhorns of West and Central Africa have lived in their present niches for several millenia. The stringency of this environment has supposedly contributed to their small size and to the “low productivity” of these breeds compared to most zebus found in the more arid areas of the tropics. Nonetheless, they have acquired a hardiness to the harsh climatic conditions and resistance to the various diseases endemic to their environments. Notable among these adaptations is their tolerance to trypanosomosis, the major disease limiting introduction of non-native livestock into the vast humid and sub-humid areas of West and Central Africa. Aboagye *et al.* (1994) and Rege *et al.* (1994a; b; c) have reviewed the distribution and important characteristics of the Shorthorn cattle while ILCA (1979) summarised the attributes of the major trypanotolerant populations of West and Central Africa.

Table 1 summarises the classification of the humpless cattle of Africa. There are two humpless Longhorn breeds, the N'Dama and the Kuri, both found in West Africa and 14 humpless Shorthorn breeds widely distributed in the humid and sub-humid zones

Table 1. *Humpless cattle breeds of sub-Saharan Africa.*

| Group and Breed/Strain | Location/Country | Estimated ^a Population ^b | Mature weight (kg) | | Withers height (cm) | | Main uses (In order of importance) |
|---|--|---|-----------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 1. Humpless Longhorns | | | | | | | |
| 1. N'Dama | All coastal countries of West and Central Africa, plus Mali, B. Faso and C.A. Republic | 4 863 000 | 220-360 | 180-300 | 95-120 | 90-115 | Meat; Work; Manure |
| 2. Kuri | Chad; Niger; Nigeria; Cameroon | 110 000 | 500-750 | 360-450 | 140-180 | 126-145 | Meat; Milk; Work |
| 2. Humpless Shorthorns (West and Central Africa) | | | | | | | |
| 3. Ghana Shorthorn | | 788 000 | 190-395 | 163-280 | 105-117 | 99-110 | Meat; Work; Milk |
| 4. Baoulé | Central African Republic; Ivory Coast, Gabon, B. Faso | 1 082 000 | 160-300 | 150-240 | 100-106 | 90-103 | Milk; Meat |
| 5. Tobi | Burkina Faso | 490 000 | | | | | Meat; Milk |
| 6. Savanna Muturu | Nigeria | 58 000 | | 150-225 | | | Meat |
| 7. Liberia Dwarf Muturu | Liberia | 5 500 | | | 86-95 | 82-94 | Meat |
| 8. Bakwéni | Cameroon | 1 300 | | | | | Meat; Rituals |
| 9. Somba | Benin; Togo | 216 000 | 150-215 | 115-185 | 89-106 | 85-103 | Meat; Milk; Rituals |
| 10. Doayo (Naunchi) | Cameroon | 7 000 | | | 100-110 | 97-106 | Meat; Rituals |
| 11. Kapsiki | Cameroon | 3 300 | | | 105-117 | 100-109 | Meat; Rituals |
| 12. Ghana Dwarf Muturu | Ghana | 100 | | | | | Meat |
| 13. Bakosi | Cameroon | 1 300 | | | | | Meat; Rituals |
| 14. Lagune | Benin; Congo; Ivory Coast; Gabon; Togo; Zaire | 65 700 | 180-280 | 165-262 | 89-106 | 85-103 | Meat; Manure |
| 15. Logone | Chad | N.A | | | | | Meat |
| 16. Forest Muturu | Nigeria | 40 000 | | | 85-95 | 83-93 | Meat; Rituals |
| 3. Humpless Shorthorns (Eastern Africa) | | | | | | | |
| 17. Sheko | Ethiopia | 31 000 | | | | | Meat; Work |

^alatest available estimate; if multiple in same year, highest estimate used; combines estimates from different countries, if applicable.

Table 2. Zebu cattle breeds of sub-Saharan Africa.

| Group and Breed/Strain | Location/Country | Estimated ^{a)} population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|--|-------------------------|---------------------------------------|--------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 1. Large East African Zebu | | | | | | | |
| 1. Ethiopian Boran (Borana) | Ethiopia | 1 896 000 | 300-385 | 300-350 | | | Milk; Meat |
| 2. Kenya (Improved) Boran | Kenya, Tanzania, Zambia | 580 570 | 550-850 | 400-550 | | | Meat; Milk; Work |
| 3. Unimproved Boran (Borana) | Kenya | 1 882 000 | 255-395 | 250-355 | | | Milk; Meat |
| 4. Orma Boran | Kenya | 547 000 | | | | | Milk; Meat |
| 5. Somali Boran | Somalia | NA | 295-410 | 230-355 | | | Milk; Meat |
| 6. Karamajong zebu | Uganda | 510 000 | 320-490 | 300-410 | | | Milk; Meat |
| 7. Toposa | Sudan | 398 000 | 350-600 | 250-390 | 130-147 | 120-133 | Milk; Meat; Work |
| 8. Murle | Sudan, Ethiopia | NA | 300-410 | 220-320 | 128-134 | 100-122 | Milk; Meat; Work |
| 9. Butana | Sudan | 258 000 | 395-600 | 300-440 | 131-149 | 123-139 | Milk; Meat |
| 10. Kenana | Sudan | 1 670 000 | 400-610 | 300-435 | 132-148 | 124-138 | Milk; Meat |
| 11. Baggara | Sudan | 3 270 000 | 300-600 | 230-450 | | | Meat; Milk; Work |
| 12. Barka | Eritrea | 850 000 | 335-490 | 295-415 | 122-138 | 120-133 | Milk; Meat |
| 13. Turkana | Kenya | 621 750 | | | | | Milk; Meat |
| 2. Small East-African Zebu | | | | | | | |
| 2.1 Abyssinian Shorthorned Zebu | | | | | | | |
| 14. Arsi | Ethiopia | 2 012 000 | | | | | Work; Meat; Milk |
| 15. Adwa | Ethiopia | NA | | | | | Work; Meat; Milk |
| 16. Ambo | Ethiopia | NA | | | | | Work; Meat; Milk |
| 17. Bale | Ethiopia | 738 000 | | | | | Work; Meat; Milk |
| 18. Goffa | Ethiopia | NA | | | | | Work; Milk; Meat |

Continued

.... Table 2.

| Group and Breed/Strain | Location/Country | Estimated* population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|---|-----------------------|--------------------------|--------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 19. Guraghe | Ethiopia | NA | | | | | Work; Meat; Milk |
| 20. Harar | Ethiopia | NA | | | | | Work; Meat; Milk |
| 21. Jem-jem | Ethiopia | 434 000 | | | | | Work; Meat; Milk |
| 22. Snada | Ethiopia | NA | | | | | Work; Meat; Milk |
| 23. Mursi | Ethiopia | NA | | | | | Milk; Meat; Work |
| 24. Hammer | Ethiopia | NA | | | | | Milk; Work; Meat |
| 25. Jijiga | Ethiopia | 100 000 | | | | | Work; Milk; Meat |
| 26. Ogaden | Ethiopia; Somalia | NA | | | | | Milk; Meat; Work |
| 2.2 The cluster of southern Sudan and Vicinity | | | | | | | |
| 27. Lugware | Uganda, D.R. Congo | 196 200 | 295-355 | 230-260 | 102-111 | 98-109 | Work; Meat; Milk |
| 28. Mongolla | Sudan | 240 000 | | 130-225 | 101-110 | 100-105 | Milk; Meat |
| 29. Nkedi | Uganda | 752 000 | 240-450 | 270-325 | 102-121 | 99-107 | Work; Milk; Meat |
| 30. Nuba Mountain Zebu | Sudan | 44 000 | 200-225 | 175-225 | | | Meat (for ceremonies) |
| 2.3 The «Somali» group | | | | | | | |
| 31. Garre (Ghera) | Somalia | NA | 290-340 | 265-305 | 115-133 | 100-120 | Milk; Meat |
| 32. Gasara | Somalia | NA | 205-295 | 200-230 | 102-129 | 101-115 | Milk; Meat |
| 33. North Somali Zebu | Somalia | NA | | | | | Milk; Work |
| 34. Baherie | Eritrea | 10 000 | 190-285 | 170-220 | 109-124 | 98-120 | Milk; Meat |
| 2.4 The Kenya cluster | | | | | | | |
| 35. Kikuyu | Kenya | 89 500 | | | | | Meat |
| 36. Taika | Kenya | NA | 194-405 | 125-360 | 90-130 | 94-122 | Meat; Milk; Work |
| 37. Giriama | Kenya | NA | | | | | Meat; Milk; Work |
| 38. Duruma | Kenya | NA | | | | | Meat; Milk; Work |

Continued

.... Table 2.

| Group and Breed/Strain | Location/Country | Estimated ^{a)} population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|---------------------------------|---------------------|------------------------------------|--------------------|---------|---------------------|---------|------------------------------------|
| | | | Male | Female | Male | Female | |
| 39. Kauba | Kenya | 897 000 | | | | | Meat; Milk; Work |
| 40. Maasai | Kenya | 1 398 000 | 300-445 | 275-385 | 118-140 | 110-135 | Milk; Meat |
| 41. Winam (Kavirondo) | Kenya | 2 110 000 | 215-410 | 195-365 | 95-132 | 94-125 | Milk; Work; Meat |
| 42. Nandi | Kenya | 389 000 | 215-420 | 200-320 | 118-122 | 110-119 | Milk; Work; Meat |
| 43. Samburu | Kenya | 19 000 | | | | | Milk; Meat |
| 44. Watende | Kenya | NA | | | | | Milk; Meat; Work |
| 2.5 <i>The Teso group</i> | | | | | | | |
| 45. Teso | Kenya, Uganda | NA | | | | | Milk; Meat; Work |
| 46. Usuk | Uganda | NA | | | | | Milk; Meat; Work |
| 47. Kyoga | Uganda | NA | | | | | Milk; Meat; Work |
| 48. Serere | Uganda | NA | | | | | Milk; Meat; Work |
| 2.6 <i>The Tanzania cluster</i> | | | | | | | |
| 2.6.1 <i>Tanganyika</i> | | | | | | | |
| <i>Shorthorned group</i> | | | | | | | |
| 49. Iringa Red | Tanzania (Mainland) | NA | | | | | Milk; Meat |
| 50. Maasai | Tanzania (Mainland) | 100 000 | | | | | Milk; Meat |
| 51. Ugogo Grey | Tanzania (Mainland) | 100 000 | | | | | Milk; Meat |
| 52. Mkalama Dun | Tanzania (Mainland) | 1 000 | | | | | Milk; Meat; Work |
| 53. Singida White | Tanzania (Mainland) | 5 000 | | | | | Milk; Meat; Work |
| 54. Pare | Tanzania (Mainland) | 1 000 | | | | | Milk; Meat; Work |

Continued

.... Table 2.

| Group and Breed/Strain | Location/Country | Estimated ^(s) population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|------------------------------------|---|-------------------------------------|--------------------|---------|---------------------|---------|---------------------------------------|
| | | | Male | Female | Male | Female | |
| 55. Tarime (Shashi) | Tanzania (Mainland) | 100 000 | 160-210 | 150-190 | | | Milk; Meat; Work |
| 56. Chagga (Wachagga) | Tanzania (Mainland) | 1 000 | 170-240 | 155-190 | | | Milk; Meat; Work |
| 2.6.2 Zanzibar group | | | | | | | |
| 57. Zanzibar Zebu | Tanzania (Zanzibar & Pemba) | | 250-350 | 190-300 | | | Milk; Work; Meat |
| 2.7 The Angoni group | | | | | | | |
| 58. Angoni | Zambia | 300 000 | 270-730 | 180-480 | 122-127 | 119-122 | Meat; Work; Milk |
| 59. Malawi Zebu | Malawi | 796 700 | | 265-280 | | | Meat; Work; Milk |
| 60. Angonia (Angone) | Mozambique | 64 400 | 265-730 | 175-470 | 121-125 | 110-120 | Meat; Work; Milk |
| 2.8 The Madagascar group | | | | | | | |
| 61. Madagascar zebu | Madagascar | 7 000 000 | 350-450 | 250-350 | 117-140 | 110-135 | Meat; Milk; Work |
| 62. Baria | Madagascar | 500 | | | | | Not used (wild) |
| 3. West African Zebu | | | | | | | |
| 3.1 Gudali Group | | | | | | | |
| 3.1.1 The Sokoto sub-group | | | | | | | |
| 63. Sokoto | Nigeria | 4 352 000 | 495-660 | 240-355 | 130-138 | 116-132 | Milk; Meat; Work |
| 3.1.2 The Adamawa sub-group | | | | | | | |
| 64. Ngaundere | Cameroon; Nigeria; C. Africa Republic | 999 000 | 400-565 | 330-410 | 132-136 | | Milk; Meat; Work |
| 65. Banyo | Cameroon; Nigeria | N/A | 300-410 | 350-365 | | | Milk; Meat; Work |

Continued

.... *Table 2.*

| Group and Breed/Strain | Location/Country | Estimated ^{a)} Population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|------------------------------------|---|---------------------------------------|--------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 66. Yola | Cameroon; Nigeria | NA | 350-355 | 315-340 | | | Milk; Meat; Work |
| 3.2 Fulani Group | | | | | | | |
| 67. Gobra | Senegal | 1 300 000 | 300-350 | 250-300 | 130-144 | 124-140 | Milk; Meat; Work |
| 68. Sudanese Fulani (Peul Zebu) | Mali, B. Faso | 5 616 000 | 250-345 | 248-300 | 120-138 | 115-126 | Milk; Work; Meat |
| 69. White Fulani | Nigeria; Cameroon; C. Africa Republic | 9 645 000 | 425-665 | 250-380 | 130-152 | 1180138 | Milk; Meat; Work; Manure |
| 70. Red Fulani | Nigeria, Cameroon; C. Africa. Rep. | 4 924 000 | | | | | Milk; Meat; Work; Manure |
| 71. Fellata (Bororo) | Chad; Sudan | 50 000 in Sudan | 400-450 | 255-410 | | | Milk; Meat; Work |
| 72. Djelli (Diali) | Niger; Nigeria | NA | | | | | Milk; Meat; Work |
| 3.3 Other West African Zebu | | | | | | | |
| 73. Azacuak (Tuareg) | Mali; Nigeria; Niger | 506 000 | 350-500 | 300-410 | 128-135 | 122-130 | Meat; Work; Milk |
| 74. Shuwa | Chad; Nigeria; Cameroon | 4 554 000 | 350-475 | 250-300 | 135-140 | 125-128 | Work; Milk; Meat; |
| 75. Maure | Mauritania; Mali | 673 000 | 250-700 | 250-350 | 125-140 | 110-128 | Milk; Meat; Work |

NA = Not available.

^{a)}Latest available estimate; if multiple in same year, highest estimate used; combines estimates from different countries, if applicable.



Figure 3. Raya Azebo cattle.

of West and Central Africa. In addition there is one humpless Shorthorn breed in eastern Africa: The Sheko of Ethiopia.

The zebu cattle

Although zebu cattle are trypano-susceptible, in the absence of tsetse fly the large size and high production levels of many zebu breeds give them a competitive advantage over the trypanotolerant N'Dama Longhorn and Shorthorns (Rege *et al.*, 1994a;b). The zebu breeds or strains are abundant in the continent and exhibit a high level of resistance to harsh environmental conditions, making them the only type of cattle that can survive over a large part of Africa.

The zebu is the largest single cattle type in Africa. It is represented by some 75 breeds (Table 2). The highest concentration of the zebu is in eastern Africa and neighbouring countries in south-central Africa which, together, have 61 breeds. West Africa has only

13 zebu breeds, principally inhabiting the dry savanna and sahelian belts. The East African Zebu can be divided into two major sub-groups - the "Large" and the "Small" East African Zebu. The former has some 13 breeds, all restricted to the relatively drier parts of Sudan, Eritrea, Ethiopia, Somalia, Kenya, Tanzania and Uganda. The 49 Small East African zebu breeds are also principally found in the same countries as their "Large" counterparts. However, a small number inhabit south-central Africa (Zambia, Malawi, Mozambique) and Madagascar. Other than Zaire, the eastern border of which is inhabited by "spill-over" zebu from eastern Africa, the humid zone of Central Africa is devoid of zebu.

The Small East African zebu is further sub-divided into several *groups* or *clusters*: The Abyssinian Shorthorned Zebu (represented by 13 breeds/strains); the Cluster of southern Sudan and vicinity (4); the Somali group (4); the Kenya cluster (10); the Teso group (4); the Tanzania cluster consisting of the Tanganyika

Table 3. The sanga cattle breeds of sub-Saharan Africa.

| Group and Breed/Strain | Location/Country | Estimated ^(a) Population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|--|--|--|--------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 1. The sanga of eastern Africa | | | | | | | |
| 1.1 The Nilotic group | | | | | | | |
| 1. Abigar | Ethiopia | 548 600 | | | | | Milk; Meat; Work |
| 2. Aliab Dinka | Sudan | NA | | | | | Milk; Meat |
| 3. Aweil Dinka | Sudan | NA | | | | | Milk; Meat |
| 4. Nuer | Sudan | NA | | | | | Milk; Meat |
| 5. Shilluk | Sudan | NA | | | | | Milk; Meat |
| 1.2 The Abyssinian sanga | | | | | | | |
| 6. Danakil | Ethiopia, Eritrea | 680 500 | 250-380 | 200-305 | 130-145 | 120-125 | Milk; Meat; Work |
| 7. Raya-Azebo | Ethiopia | 521 000 | | | | | Work; Milk; Meat |
| 1.3 The Ankole group | | | | | | | |
| 8. Watusi | Uganda; Rwanda; Burundi; Tanzania; D.R.C | 1 600 000 | 350-425 | 290-350 | 132-135 | 110-127 | Milk; Meat; Work |
| 9. Bahima | Uganda; D.R.C. | * | | | | | Milk; Meat; Work |
| 10. Kigezi | Uganda | * | 220-380 | 200-330 | 112-132 | 108-120 | Milk; Meat; Work |
| 11. Bashi | D.R.C. | * | 235-400 | 220-340 | 115-135 | 110-124 | Milk; Meat; Work |
| 12. Ruzizi | DRC; Rwanda; Burundi | * | | | | | Milk; Meat; Work |
| 2. The sanga of southern Africa | | | | | | | |
| 2.1 The Shona group | | | | | | | |
| 13. Mashona | Zimbabwe | 500 000 | 350-635 | 260-410 | | | Meat; Work |
| 2.2 The Nguni group | | | | | | | |
| 14. Nguni | S. Africa; Swaziland | 2 156 000 | 400-680 | 225-450 | | | Meat; Work; Milk |

.... Continued...

.... Table 3.

| Group and Breed/Strain | Location/Country | Estimated ^{a)} Population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|---|------------------|---------------------------------------|--------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 15. Nkone | Zimbabwe | 400 | | 300-450 | | | Meat; Milk |
| 16. Pedi | S. Africa | 400 | | | | | Meat; Work; Milk |
| 17. Shangan | S. Africa | 600 | | | | | Meat; Work; Milk |
| 18. Landim | Mozambique | 536 400 | | | | | Meat; Work; Milk |
| 2.3 The Zambia/Angola cluster | | | | | | | |
| 19. Tonga | Zambia | 993 000 | 485-530 | 310-495 | 118-130 | | Milk; Meat; Work |
| 20. Porto-Amboim | Angola | NA | 400-530 | 350-425 | 120-140 | 118-129 | Meat |
| 2.4 Ovambo and south-western cluster | | | | | | | |
| 21. Ovambo | Namibia | NA | | | | | Meat; Work; Milk |
| 22. Kaokoveld | Namibia | NA | | | | | Meat; Work; Milk |
| 23. Okavango | Namibia | NA | | | | | Meat; Work; Milk |
| 24. Caprivi | Namibia | NA | | | | | Meat; Work; Milk |
| 25. Humbi | Angola | NA | | | | | Meat; Work; Milk |
| 2.5 The Setswana group | | | | | | | |
| 26. Barotse | Zambia; Angola | 793 000 | 255-700 | 240-455 | 120-137 | 114-129 | Milk; Meat; Work |
| 27. Damara (Herero) | Namibia | NA | | | | | Milk; Work; Meat |
| 28. Tswana | Botswana | 1 395 000 | 310-520 | 290-420 | 140 | 131 | Meat; Work; Milk |
| 29. Tuli | Zimbabwe | 3 300 | 450-820 | 360-570 | | | Meat |
| 2.6 The Afrikaner group | | | | | | | |
| 30. Afrikaner | South Africa | 302 000 | 450-950 | 360-555 | 130-150 | 128-140 | Meat; Work |

NA = Not available.

^{a)}Possibly included in Watutsi population figure.^{b)}Latest available estimate; if multiple in same year, highest estimate used; combines estimates from different countries, if applicable.

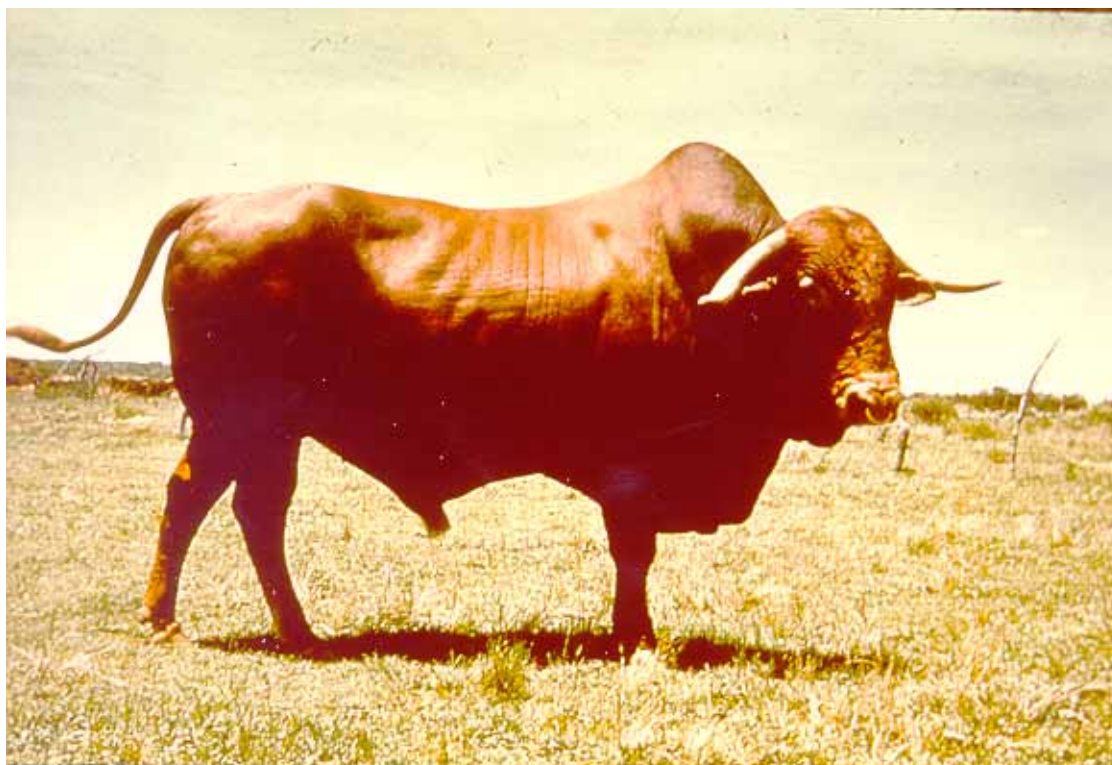


Figure 4. Afrikaner cattle.

group (8) and the Zanzibar Zebu (1); the Angoni group (3); and the Madagascar group (2).

The West African zebu consists of two main groups: The Gudali group represented by two sub-groups (Sokoto with only one breed, and Adamawa with 3 breeds/strains); and the Fulani group (with 6 breeds/strains). In addition, there are three other zebu breeds in West Africa, the Azoauak, Shuwa and Maure, which do not belong to either the Gudali or the Fulani groups.

The sanga cattle

Table 3 summarises the sanga cattle breeds/strains of Africa. There are 30 sanga breeds/strains sub-divided on the basis of location into the sanga of eastern (12 breeds/strains) and sanga of southern (18) Africa. The sanga of eastern Africa consist of three groups: Nilotic sanga of southern Sudan and south-western Ethiopia; the Abyssinian sanga of Ethiopia and Eritrea; and

the Ankole group with representatives in Uganda, Rwanda, Burundi, Tanzania and Democratic Republic of Congo (former Zaire).

The sanga of southern Africa are represented by six groups or clusters: the Shona represented by the *Mashona* of Zimbabwe; the Nguni group (5 breeds/strains); the Zambia/ Angola cluster (2); the Ovambo and south-western cluster (5); the Setswana group (4); and the Afrikaner group represented only by the *Afrikaner* breed.

The zenga cattle

Several breeds have supposedly resulted from crossbreeding between the zebu and sanga populations in the East African highlands where large concentrations of zebu (arriving from Asia) initially occurred, providing opportunity for admixture with sanga cattle, then already resident there. The resulting breeds have been classified into a separate category. The name "zenga" is suggested for this category. Exclusively located in eastern

Africa, some members are found in predominantly zebu habitat, others in sanga habitat. Indeed, the location of the zenga forms a natural division between the "zebu country" in the north and the predominantly "sanga country" in the south. Members of the zenga - a total of eight - are: the *Arado*, *Fogera*, and *Horro* (of Ethiopia); *Jiddu* (southern Somalia); *Alur*, also called *Nioka* (*Nyoka*) or *Blukwa* cattle (Democratic Republic of Congo); *Nganda* (Uganda); *Sukuma* (Tanzania); and *Bovines of Tete* (Mozambique). The zenga cattle are summarised in Table 4.

Recent derivatives

There are several cattle breeds which have been formed as a result of the coexistence of two or more breeds in close proximity to each other. In most cases this has been facilitated by

increased interaction among tribal groups and, sometimes, deliberate but non-systematic attempts to improve specific attributes. A good example of the process, but one which has not produced a recognised, distinct breed or strain, is the crossbreeding between the Kuri and zebu of the Lake Chad Basin (see Tawah *et al*, 1997) to produce a draught animal. Table 4 summarises breeds/strains in this category.

Commercial composites

Sub-Saharan Africa is home to, at least, six commercial composite breeds with varying proportions of exotic blood. Unfortunately, only two of them - the *Drakensberger* and the *Bonsmara*, both of South Africa - are secure in terms of numbers and existence of



Figure 5. Ankole cattle.

Table 4. Zenga (zebu-sanga) cattle, recent derivatives and synthetic breeds.

| Group and Breed/Strain | Location/Country | Estimated ^{a)} Population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|---|---------------------------|---------------------------------------|--------------------|---------|---------------------|---------|--|
| | | | Male | Female | Male | Female | |
| 1. Zenga (zebu x sanga) cattle | | | | | | | |
| 1. Arado | Eritrea; Ethiopia | 510 000 | 205-430 | 192-350 | 117-144 | 93-126 | Work; Milk; Meat |
| 2. Fogera | Ethiopia | 86 800 | | | 110-145 | 100-121 | Work; Milk; Meat |
| 3. Horro | Ethiopia | 3 286 000 | 320-480 | 210-400 | | | Work; Milk; Meat |
| 4. Jiddu | Somalia | NA | 340-590 | 325-430 | 109-133 | 108-124 | Milk; Meat |
| 5. Alur (Nioka, Blukwa) | D.R.C. | NA | 290-550 | 225-380 | 110-131 | 108-122 | Milk; Meat; Work |
| 6. Nganda | Uganda | 1 365 000 | 280-420 | 200-340 | 115-124 | 115-122 | Milk; Meat; Work |
| 7. Sukuma (Tinde) | Tanzania | 100 000 | 230-410 | 210-370 | 94-132 | 93-127 | Milk; Meat |
| 8. Bovines of Tete | Mozambique | NA | | | | | Meat; Milk; Work |
| 2. Recent derivatives | | | | | | | |
| 1. Borgou | Benin; Togo | 428 000 | 190-330 | 180-295 | | | Meat; Work; Manure |
| 2. Méré | B. Faso; Cote d'Ivoire | 693 000 | | | | | Meat; Work; Manure |
| 3. Ghana «sanga» | Ghana | 124 000 | | | | | Milk; Meat; Work; Manure |
| 4. Keteku | Nigeria | 293 000 | | | | | Meat; Work; Manure |
| 5. Djakore | Senegal | 350 000 | | | | | Milk; Meat; Work; Manure |
| 6. Bin | Nigeria | NA | | | | | Meat; Work; Manure |
| 7. Basuto | Lesotho | NA | | | | | Work; Meat; Milk |
| 8. Barra do Cuanzo | Angola | NA | 370-620 | 360-500 | | | Meat; Work; Milk |
| 9. Rana (Omby Rana) | Madagascar | 40 000-85 000 | | | | | Milk; Meat |

.... Continued

... Table 4.

| Group and Breed/Strain | Location/Country | Estimated ^{a)} Population | Mature weight (kg) | | Withers height (cm) | | Main uses (in order of importance) |
|--|----------------------------|---------------------------------------|--------------------|---------|---------------------|--------|--|
| | | | Male | Female | Male | Female | |
| 3. Systematic synthetics/composites | | | | | | | |
| 1. Drakensberger | South Africa; Swaziland | 149 000 | | | | | Meat; Work; Milk |
| 2. Bonstnara | South Africa | 143 000 | | | | | Meat |
| 3. Renitelo | Madagascar | 2 000-3 000 | 420-655 | 305-450 | | | Meat; Work |
| 4. Manjan'i Boina | Madagascar | 200 | | | | | Milk |
| 5. Mpwapwa | Tanzania | 1 500 | 360-620 | 290-455 | | | Milk |
| 6. Wakwa | Cameroon | NA | | | | | Meat |

NA = Not available.

^{a)}Latest available estimate; if multiple in same year, highest estimate used; combines estimates from different countries, if applicable.

programmes for genetic improvement. The other four are the *Mangan'i Boina* of Madagascar (a synthetic dairy breed), the *Renitelo*, also of Madagascar (beef), the *Mpwapwa* of Tanzania (dairy) and the *Wakwa* of Cameroon (beef). Details of these breeds are discussed in the second paper (Rege and Tawah, 1999).

Breeds at Risk

As has been alluded to, one of the objectives of the survey was to establish the status of sub-Saharan African cattle breeds and to identify those which are at risk as well as those which may have become extinct in the recent past. In the absence of time-trend census data, it was not possible to provide quantitative information on trends. However, the study results, mainly qualitative, together with an assessment of pressures currently affecting breeds, were used to draw conclusions regarding possible threat categories of different breeds considered to be at risk. A total of 47 (32%) breeds/strains were identified to be at risk (Table 5). Four risk categories were defined according to FAO (1992): Critical (most threatened), Endangered, Vulnerable, and Rare (least threatened). Out of the 47 breeds at risk, 15 (Pare, Mkalama Dun and Chagga of Tanzania; Bakweri, Kapsiki, Bakosi and Wakwa of Cameroon; Ghana Dwarf Muturu or Shorthorn; Nkone of Zimbabwe; Pedi and Shangan of South Africa; Renitelo and Baria of Madagascar; and Sengologa and Seshaga of Botswana) were classified in the "Critical" category, 10 were "Endangered", another 10 were "Vulnerable" and six were "Rare" (Table 5). The remaining six could not be definitively classified: Five were classified as lying somewhere between "Rare" and "Vulnerable" and one between "Endangered" and "Vulnerable".

Extinct Breeds

Starting with the 145 breeds identified in this survey and working backwards, a review of old literature dating as far back as 1902 was

Table 5. Breeds of sub-Saharan African cattle considered at risk.

| Risk status | Breed/strain | Principal location | Main causes of threat |
|-----------------------|------------------------|-----------------------------|--|
| Rare | Sekgatla | Botswana | Interbreeding with "Tswana"; Crossbreeding with Afrikaner |
| | Basuto | Lesotho | Crossbreeding |
| | Jiddu | Somalia | Neglect; Conflict |
| | Bovines of Tete | Mozambique | Conflict; Crossbreeding |
| | Biu | Nigeria | Interbreeding |
| Rare/Vulnerable | 6. Manjan'i Boina | Madagascar | Early stage of breed development |
| | Dongola | Northern Sudan | Interbreeding with Red Butana |
| | Shendi | Northern Sudan | Interbreeding with Red Butana |
| | Bambawa | Sudan (near Eritrea border) | Interbreeding with Red Butana |
| | Ingessana | Sudan-Ethiopia border | Conflict; interbreeding |
| | Kuri | Lake Chad Basin | Reduction in habitat; Crossbreeding; Conflict |
| | Vulnerable | Kyoga | Uganda |
| Usuk | | Uganda | Conflict; interbreeding |
| Singida White | | Tanzania | Interbreeding with neighbouring breeds |
| Tarime (Shashi) | | Tanzania | Interbreeding with neighbouring breeds |
| Watusi | | Rwanda, Burundi | Conflict, crossbreeding/interbreeding |
| Bahima | | Uganda | Conflict, crossbreeding/interbreeding |
| Bashi | | D.R. Congo | Conflict, crossbreeding/interbreeding |
| Ruzizi | | Burundi | Conflict, crossbreeding/interbreeding |
| Landim | | Mozambique | Conflict, Crossbreeding, Replacement |
| Rana | | Madagascar | Continued, unsystematic crossbreeding |
| Vulnerable/Endangered | Nigerian Forest Muturu | Nigeria | Neglect; Replacement |

.... Continued

.... Table 5.

| Risk status | Breed/ strain | Principal location | Main causes of threat |
|-------------|-------------------|--------------------|--|
| Endangered | Serere | Uganda | Conflict; interbreeding |
| | Watende | Kenya | Interbreeding with neighbouring breeds/strains |
| | Iringa Red | Tanzania | Interbreeding with Ugogo Grey |
| | Sheko | Ethiopia | Interbreeding with zebus |
| | Kikuyu Zebu | Kenya | Neglect; Crossbreeding |
| | Liberian Dwarf | Liberia | Neglect; Replacement; Conflict |
| | Muturu | Chad | Neglect |
| | Logone | Cameroon | Neglect |
| | Doayo (Nanchu) | Cameroon | Neglect |
| | Damara (Herero) | Namibia | Neglect; Crossbreeding, Replacement |
| | Mpwapwa | Tanzania | Absence of sustained dev. Programme |
| | Pare | Tanzania | Interbreeding |
| | Mklama Dun | Tanzania | Interbreeding |
| | Chagga (Wachagga) | Tanzania | Crossbreeding and interbreeding |
| Critical | Bakweri | Cameroon | Neglect |
| | Kapsiki | Cameroon | Neglect |
| | Bakosi | Cameroon | Neglect |
| | Ghana Dwarf | Ghana | Neglect |
| | Muturu/Shorthorn | Zimbabwe | Neglect |
| | Nkone | S. Africa | Interbreeding; Replacement |
| | Pedi | S. Africa | Interbreeding; Replacement |
| | Shangan | Botswana | Interbreeding with "Tswana"; Neglect |
| | Sengologa | Botswana | Interbreeding with "Tswana"; Neglect |
| | Seshaga | Botswana | Interbreeding with "Tswana"; Neglect |
| | Rruitelo | Madagascar | Interbreeding; Dermatophilosis |
| | Baria | Madagascar | Neglect; Lives in the wild |
| | Wakwa | Cameroon | Absence of sustained dev. programme |

*Crossbreeding as used here involves exotics; interbreeding involves other (neighbouring) indigenous breeds.

Table 6. Breeds of sub-Saharan African cattle considered to be extinct.

| Breed/strain | Class | Location | Last Report | Reference |
|--------------------------------------|--------------------|--|------------------------|--|
| 1. Nuba (Delami or Koalib) Shorthorn | Humpless shorthorn | Sudan | 1900s | Epstein (1971) |
| 2. Gimira («Kuri») | Humpless Longhorn | Ethiopia | 1929 | Encyclopædia Britannica (1929) |
| 3. Karagwe Shorthorn | Humpless Shorthorn | Uganda? | 1927 | Stuhlmann (1927) |
| 4. Sesse Shorthorn | Humpless Shorthorn | Sesse Islands, L. Victoria | 1909 | Epstein (1971) |
| 5. Kigezi Shorthorn | Humpless Shorthorn | Uganda | 1920s | Ford and Hall (1947) |
| 6. Socotra Shorthorn | Humpless Shorthorn | Socotra Islands, Off Horn of Africa | 1960s | Payne (1964) |
| 7. Unguja (Pemba or Mafia) Shorthorn | Humpless Shorthorn | Pemba, Mafia Islands | 1920s | Payne (1964) |
| 8. Manjaca | Humpless Shorthorn | Guinea Bissau | 1980s | JCA (1992) |
| 9. Bimal | Zebu | Somalia | 1955 | Rosetti and Congiu (1955) |
| 10. Singhi | Zebu | Somalia | 1953 | Bozzi and Triulzi (1953) |
| 11. Baria (not Madagascar Baria) | Zebu (zenga?) | Eritrea | 1929 | Marchi (1929) |
| 12. Ugogo | Zebu | Tanzania | 1920s | McCall (1928) |
| 13. Senegambia Shorthorn | Humpless Shorthorn | The Gambia; Southern Senegal (Casamance) | 1950s | Mason (1951); Epstein (1971) |
| 14. Arussi-Galla | Sanga | Ethiopia | 1900s | Lydekker (1912) |
| 15. Ugoi | Sanga | Tanzania | 1850s | Burton (1961) |
| 16. Ngami (Botswana) | Sanga | Botswana | 1900s | Lydekker (1912), Curson (1934) |
| 17. Nama | Sanga | Botswana | 1906 | Groenewald and Curson (1933) |
| 18. Bolowana | Sanga (?) | South Africa | Late 1890s | Thompson (1932) |
| 19. Sakalava | Sanga (?) | Madagascar | Late 1890s | Keller (1898); Lydekker (1912); Murdock (1959) |
| 20. Tottentot | Sanga (?) | S. Africa | Late 1800s/Early 1900s | Kolb (1719); Smith (1827); Jones (1953); Martinho (1955) |
| 21. Wadai Dinka | Sanga | D.R. Congo | 1936 | Curson and Thornton (1936) |
| 22. Mbulu | Zebu | Tanzania | 1953 | Jeffreys (1953) |

done. Each breed or population appearing in the old literature was followed progressively in subsequent publications. Of the breeds that could not be found in more recent literature, a large number was due to change in breed names over the years. However, some breeds which existed previously could not be found in recent literature under any name. To determine what may have happened to these "missing" breeds or strains, clarification was sought from the locations at which they were last reported. This process resolved some of the anomalies and helped to identify those populations which, on the basis of information available, had ceased to exist. Some of these were on record in the countries concerned as officially extinct while the rest could only be considered to be extinct because local officials and communities could not recall their previous existence or were certain that they had, through the years, disappeared as a result of one reason or another. A total of 22 breeds which existed at some point during this (20th) century could not be located and were considered extinct (Table 6). Of the extinct breeds, only one (*Gimira* of Ethiopia) was a humpless Longhorn, eight were humpless Shorthorns, four were zebu (one of them – the Baria of Eritrea – may have been a zenga), while the remaining seven were sanga, although the classification of three of these, Bolowana, Sakalava and Hottentot, as sanga was not certain.

Acknowledgements

This survey would not have been possible without the cooperation and, in several cases, direct involvement, of many persons and institutions. Our gratitude to the government ministries, universities, national research institutes, farmers organisations, including breed societies in Kenya, South Africa and Zimbabwe, individual farmers, scientists and extension staff and, last but not least, ILRI technical staff, particularly those who accepted the additional task of collecting data during blood sampling field expeditions. Special thanks to the several enumerators and research assistants who spent endless hours

extracting information from the grey literature. In the latter category I would like to thank, most especially, Yetnayet Mamo. This manuscript was diligently word processed by Wagaye Wolde Mariam.

References

- Aboagye, G.S., Tawah, C.L. & Rege, J.E.O.** 1994. Shorthorn cattle of West and Central Africa III. Physical, adaptive and special genetic characteristics. *World Anim. Rev.* 78 (1) 22-32.
- Blench, R.** 1993. Ethnographic and linguistic evidence for the prehistory of African ruminant livestock, horses and ponies. In: Shaw, T., Sinclair, P., Andah, B. and Okpoko, A (eds.). *The Archaeology of Africa*, Routledge, London, p.71-103.
- Bozzi, L. and Triulzi, G.A.** 1953. Osservazioni sugli animali domestici allevati in Somalia. *Riv. Agric. Subtrop.* 47:266-294.
- Burton, R.F.** 1961. *The Lake Regions of Central Africa*. 2 vols. New York.
- Curson, H.H.** 1934. Bechuana or Mangwato Cattle. *Farming in South Africa*. September 1934.
- Curson, H.H. and Thornton, R.W.** 1936. A contribution to the study of African native cattle. *Onderstepoort Journal of Veterinary Science and Animal Industry*. Vol. 7, No. 2, October 1936.
- Encyclopaedia Britannica.** 1929.
- Epstein, H.** 1971. *The origin of the domestic animals of Africa* Vol. 1 New York, USA, Africana Publishing Corporation.
- Epstein, H. and Mason, I.L.** 1984. Cattle. In: *Evolution of domesticated animals* (I.L. Mason, ed.), New York, USA, Longman, 6-27.

-
- FAO.** 1992. The management of global animal genetic resources. FAO Animal Production and health Paper No. 104. Food and Agriculture Organisation of the United Nations (FAO), Rome, Italy.
- Ford, H. and Hall, de Z.R.** 1947. The history of Karagwe, Bukoba District. Tanganyika Notes and Records 24:3
- Groenewald, J.W. and Curson, H.H.** 1933. A note on Ovambo cattle. Studies in native animal husbandry (6). Onderstepoort Journal of Veterinary Science and Animal Husbandry Vol 1, No. 2.
- ILCA** 1979. Trypanotolerant livestock in West and Central Africa. Vol. 1: General Studies and Vol. 2: Country Studies. International Livestock Centre for Africa (ILCA). Monograph No. 2, Addis Ababa, Ethiopia.
- ILCA.** 1992. Domestic Animal Genetic Resources Information Database (DAGRID). International Livestock Research Institute (ILRI), Animal Genetic Resources, Addis Ababa, Ethiopia.
- Jeffreys, M.D.W.** 1953. *Bos brachyceros* or dwarf cattle. Vet. Record 65: 393-396; 410-415.
- Jones, H.B.C.** 1955. Contribuicao para a Estudo da Origem do Bovino Africander. Boletim da Sociedade de Estudo de Moçambique No. 91.
- Keller, C.** 1898. Die Ostafrikanischen Inseln. Bibliothek der Landerkunde. Berlin.
- Kolb, P.** 1719. Caput Bonae Spei Hodiernum. Nuremberg.
- Lydekker, R.** 1912. The Ox and its Kindred. London.
- Marchi, E.** 1929. Studi sulla pastorizia della Colonia Eritrea (2nd ed.). Istituto Agricolo Coloniale Italiano, Florence.
- Martinho, J.P.** 1955. Contribuição para a Estudo da Origem do Bovino Africander. Boletim da Sociedade de Estudo de Moçambique No. 91.
- Mason, I.L.** 1951. *The Classification of West African Livestock*, Commonwealth Bureau of Animal Breeding and Genetics, Edinburgh, Technical Communication No. 7.
- McCall, F.J.** 1928. Annual Report of the Department of Veterinary Science and Animal Husbandry, 1926. Tanganyika Territory. London.
- Murdock, G.P.** 1959. Africa – Its Peoples and their Culture History. New York. Toronto. London.
- Payne, W.J.A.** 1964. The origin of domestic cattle in Africa. Emp. J. Exp. Agric. 32(126):97-113.
- Rege, J.E.O. and Tawah, L.C.** 1999. The state of African cattle genetic resources II. Geographical distribution, characteristics and uses of present-day breeds and strains. Animal Genetic Resources Bulletin (submitted).
- Rege, J.E.O.** 1998. The contribution of indigenous livestock to the economies of sub-Saharan African countries. In: Livestock Development Policies in Eastern and Southern Africa, Proceedings Regional Seminar, Mbabane, Swaziland, July 28 - August 1, 1997. Pub. CTA, 91-108.
- Rege, J.E.O., Aboagye, G.S. & Tawah, L.C.** 1994a. Shorthorn cattle of West and Central Africa I. Origin, distribution, classification and population statistics. World Anim. Rev. (FAO) 78 (1): 2-13.
- Rege, J.E.O., Aboagye, G.S. & Tawah, C.L.** 1994b. Shorthorn cattle of West and Central Africa II. Ecological settings, utility, management and production systems. World Animal Rev. (FAO) 78 (1): 14-21.
-

.....

Rege, J.E.O., Aboagye, G.S. & Tawah, C.L. 1994c. Shorthorn cattle of West and Central Africa III. Production characteristics. *World Animal Rev.* (FAO) 78 (1): 33-48.

Rossetti, G. and Congiu 1955. Ricerche zootecnico-veterinarie sugli animali domestici della Somalia. Ispettorato Veterinario, Amministrazione Fiduciaria Italiana della Somalia. Mogadishu. pp. 207.

Smith, H. 1827. *Griffith's Animal Kingdom*. London.

Stuhlmann, F. 1927. *Die Tagebücher von Dr. Emin Pascha, Vol IV*. Braunschweig, Berlin, Hamburg.

Tawah, C.L., Rege, J.E.O. & Aboagye, G.S. 1997. A close look at a rare African breed-the Kuri cattle of Lake Chad Basin: origin, distribution, production and adaptive characteristics. *S. African J. Anim. Sci.* 27 (2): 31-40.

Thompson, F.R.B. 1932. Studies in native animal husbandry V. Indigenous cattle in the Transkeian Territories. *Journal of the South African Veterinary Medical Association* 3: 190-193.

Characteristics of Munjal sheep

B.P. Kushwaha, Riyazuddin, R. N. Singh & S. Parthasarathy

Central Sheep and Wool Research Institute, Avikanagar Rajasthan, 304 501, India

Summary

Munjal is a mutton-type sheep found in some districts of the Haryana, Punjab and Rajasthan states. A sample survey was conducted in two districts of Rajasthan and one district of Punjab to record characteristics of Munjal sheep. Information was recorded on morphological characteristics, body measurements and body weight. Munjal sheep are quite big in size, tall, rectangular and massive with a dark brown face. Measurements were recorded of body length, height, heart girth, paunch girth, ear length and tail length. Adult body weights were also recorded. Wool samples were collected and analysed for their quality attributes. Wool from this breed is very coarse and hairy.

Résumé

La race Munjal est un type de mouton qui se trouve dans certains départements des états de Haryana, Punjab et Rajasthan. Une enquête sur échantillon a été conduite dans deux départements du Rajasthan et dans un du Punjab dans le but d'enregistrer les caractéristiques du mouton Munjal. On a noté les caractéristiques morphologiques, les mesures et le poids corporel. Le Munjal est un animal de grande taille, d'apparence rectangulaire et massif avec une tête de couleur brun foncé. Les mesures sur la longueur du corps, la hauteur, les circonférences du torax, la longueur des oreilles et de la queue ont été enregistrés, ainsi que le poids corporel adulte. On a ramassé des échantillons de laine qui ont été analysés pour évaluer leur qualité. La laine de cet animal est grossière et velue.

Key words: Munjal, Body weight, Wool quality, Punjab.

Introduction

The origins of Munjal sheep are not known exactly but it is supposed to have originated in India through the sheep breeders of Rajasthan, Punjab and Haryana breeding Nali with Lohi sheep (Basuthakur, 1988; Mason, 1988). The Munjal, a mutton-type sheep, is found in the Hissar, Ambala, Patiala, Karnal, Ganganagar and Bhatinda districts of the mentioned states. They are popular among the farmers for their heavy body weight. Farmers generally sell their surplus male lambs at the age of 8-10 months when weight is reported to be around 35 kg. Wool from this breed is very coarse and hairy. Traditional shepherds and landless farmers normally rear this type of sheep, which graze mostly in the outskirts of the villages, on stubble from the harvested crops, on canal banks and by the roadside. Information on population and distribution has not yet been documented and similarly, literature on Munjal sheep is scarce.

Materials and Methods

A sample survey was conducted on 76 farmers/sheep flocks of 24 villages representing five departments (tahsils) in two districts (Ganganagar and Hanumangarh) of Rajasthan and one district (Muktshar) of Punjab. In Rajasthan, a total of 66 farmers was surveyed, only 28 of whom kept Munjal sheep (mostly breeding rams) in their flocks. In the Muktshar district of Punjab all 10 farmers surveyed had Munjal sheep in



Figure 1. Munjal ram.

their flock with an average of 72 sheep per flock. Information related to morphological characters, body measurements and body weight was thus recorded from only 38 flocks, taking one or two animals from each flock. Wool samples were also collected and analysed for their quality attributes.

Results and Discussion

Morphological characters and body measurements

The animals of this breed are quite big in size, tall, rectangular and massive. They have a long head with a roman nose and narrow



Figure 2. Munjal ewe.

forehead. The face is generally tan or brown in colour which may extend up to the middle of the neck. Ears are long and leaf-like, hanging down beside the flat cheeks. Body fleece is generally white with a skin colour varying from pink to light blue. Legs are long and strong with brown to dark hooves. The tail is long. The udder is medium sized and well developed with medium sized teats. Both sexes are polled. The face is clear of wool and legs and belly are generally bare. Photographs of the animals of this breed taken during the survey are presented in figures 1, 2, 3 and 4.

Body measurements were recorded on 21 males and 49 females. Since most of the farmers keep only 1 or 2 rams in their flock

and rest of the males are disposed of at marketable age (8-10 months), males measured were mostly breeding rams.

Length, height, heart girth, paunch girth, ear length and tail length averaged 82.76 ± 1.07 , 78.71 ± 1.01 , 97.57 ± 1.51 , 99.47 ± 2.05 , 18.14 ± 0.64 and 44.85 ± 1.82 cm, respectively in males and 74.55 ± 0.83 , 68.73 ± 0.59 , 85.24 ± 0.72 , 89.20 ± 1.27 , 17.14 ± 0.44 and 39.13 ± 0.86 cm, respectively in females.

Body weight

Body weights were recorded on 15 males and 32 females. Adult body weight in males ranged from 50 to 83 kg with an average of 65.93 ± 2.43 kg. In females it ranged from 35 to 55 kg with an average of 45.37 ± 0.99 kg.

Table 1. Wool quality parameters

| Diameter (μ) | Hetero (%) | Hairy (%) | Medullation (%) | Staple length (cm) | Crimp (per cm) | Tenacity (g/tax) | Extension (%) |
|--------------------------|---------------------------|---------------------------|---------------------------|--------------------------|--------------------------|--------------------------|---------------------------|
| 51.18 ± 0.97 (90) | 11.71 ± 0.859 (90) | 52.65 ± 1.414 (90) | 64.37 ± 1.414 (90) | 9.05 ± 0.534 (90) | 0.47 ± 0.795 (89) | 4.68 ± 0.707 (82) | 13.97 ± 1.149 (81) |

The number of observations are between parenthesis.



Figure 3. Munjal lamb.



Figure 4. Munjal flock grazing at a roadside in Punjab.

Generally farmers sell their surplus ram lambs to the local butchers at the age of 8 to 10 months of age, body weight at this age varies from 30 to 40 kg.

Wool yield and quality

Flock owners were interviewed to collect information on wool production. Wool is coarse and hairy and is around 1.5 kg per shearing with two shearings a year in March/April and Sept./October. Wool samples were collected on 90 animals and were analysed for their quality attributes. Results are presented in table 1. The average fibre diameter, medullation and staple length of six-monthly growth were $51.18 \pm 0.97 \mu$, $64.37 \pm 1.41 \%$ and 9.05 ± 0.53 cm, respectively. Similar staple length was also reported by Basuthakur (1988). The diameter is very high, indicating that these fibres may produce only very coarse yarn. Medullation being very large it may be very difficult to produce even coarse yarn. However, these wools can be

blended with finer wools to improve the quality of the blends. Such blends will be useful for manufacturing carpet, dhurries, coarse blankets etc. The staple length is quite satisfactory or even better because of coarseness. Tenacity and extension are low because most fibres are of a hairy type. This was also observed when the fibres were combed to parallel them and weigh the fractured fibres after testing for tenacity fibres which were loosely packed or single rather than being a small cohesive mass. Crimps are low again due to the hairiness of the wool samples.

Reproduction and breeding

It is customary among the farmers to keep breeding rams in the flock all times. Rams are given utmost care and are supplemented with concentrate throughout the year. In the farmer's flock, age at first mating is reported to be around 12 to 15 months in females. Males are generally used for breeding at around 18 months of age. Ewes normally have single births and twinning is rare (about 5 %). Basuthakur (1988) reported that the animals of this breed can start breeding quite

early in life and are known for good rate of fertility and milk yield. Ewes are generally not milked.

Acknowledgments

We are thankful to the Director of the Sheep and Wool department of Rajasthan. District Sheep and Wool Officer of Suratgarh, Dr. R. K. Gandhi, Sheep and Wool extension Officer of Hanumangarh, Shri O. P. Kalwa, Sheep and wool extension supervisor, Sadulsahar, and Senior Veterinary officer Malout, Punjab for their help and cooperation during the survey.

References

Basuthakur, A. K. 1988; Sheep Research Production and Marketing in India. Inter-India publications, New Delhi, India , pp. 117.

Mason, I. L. 1988; World Dictionary of Livestock Breeds. C.A.B. International, pp. 297.

Conservation of livestock breed diversity

J.S.F. Barker

*Department of Animal Science, University of New
England, Armidale, NSW 2351, Australia*

Summary

Mankind uses some 40 species of animals as domestic livestock to meet our needs for food, clothing, power, etc. Within these species, there are in total some 4,500 breeds that are referred to as the global animal genetic resources. Each breed comprises a unique set of genes. More than 30% of breeds are estimated to be at risk of extinction, and many more, particularly in developing countries, are threatened by inefficient utilization. The Food and Agriculture Organization of the United Nations has been mandated by its member nations to manage the global animal genetic resources, and major progress has been made in the last few years. However, resources are limited, and priorities will have to be set for breed conservation, for breed development programmes and for evaluation studies. Breeds that are taxonomically distinct should be favoured for conservation, the objective being to maintain maximum genetic diversity of each livestock species. Genetic distances and phylogenetic diversity provide the best available objective criterion, and microsatellites are the current markers of choice for obtaining the genetic data. Microsatellite-based genetic distances will describe breed similarities due to common ancestry, but cannot account for consequences of artificial or natural selection. Phylogenetic trees for 11 water buffalo populations in southeast Asia, constructed using 25 polymorphic protein coding loci or 21 microsatellite loci, show differences in both topology and branch lengths, but the microsatellite tree is a better representation of the similarities due to common ancestry. Thus

phylogenetic diversity, based on microsatellite loci, should be used as an initial guide in making conservation decisions for livestock breeds.

Resumen

El Hombre utiliza unas 40 especies animales como ganado doméstico para satisfacer sus necesidades de alimentación, ropa, tracción, etc. Dentro de estas especies, existe un total de 4 500 razas conocidas como recursos genéticos animales globales. Cada raza comprende un grupo único de genes. Se estima que más del 30% de las razas están en peligro de extinción y muchas más, sobre todo en los países en vías de desarrollo, están amenazadas por una utilización ineficaz. La Organización para la Alimentación y la Agricultura de las Naciones Unidas ha sido encomendada por sus naciones miembros de gestionar los recursos genéticos animales globales. Se han hecho grandes progresos en este sentido en los últimos años, sin embargo, los recursos son limitados, y será necesario establecer prioridades para la conservación de razas, para programas de desarrollo de razas y para estudios de evaluación. Las razas taxonómicamente diferentes deberían ser favorecidas para la conservación, ya que el objetivo es de mantener la máxima diversidad genética de cada especie ganadera. Las distancias genéticas y la diversidad filogenética proporcionan el mejor criterio objetivo disponible y los microsatélites son actualmente los marcadores elegidos para obtener los datos genéticos. Las distancias genéticas basadas en microsatélites describirán las similitudes entre razas debido a sus antecedentes comunes pero no podrán

explicar las consecuencias de la selección artificial ni natural. Árboles filogenéticos para 11 poblaciones de búfalos de agua en el sudeste de Asia, elaborados utilizando 25 loci polimórficos codificadores de proteína o 21 loci de microsatélites, muestran diferencias tanto en topología como longitudes de ramas, pero el árbol de microsatélites es una mejor representación de las similitudes debidas a antecesores comunes. Por consiguiente, la diversidad filogenética, basada en loci de microsatélites, debería utilizarse como una guía inicial para tomar decisiones sobre la conservación de razas ganaderas.

Key words: Conservation priorities, Biodiversity measurement, Microsatellite, Phylogenesis.

Introduction

The total global biodiversity most likely includes tens of millions of species. Of this vast number, one species - our own, uses some 40 other animal species to meet our demands for food (meat, milk, eggs), clothing (wool and other fibres, skins), draft power and manure, to serve as a "bank" or hedge against hard times, and to satisfy various cultural, religious and recreational purposes. For these domestic livestock species, conservation at the species level (in the sense of preventing their loss) clearly is not an issue; what is of concern is conservation of diversity within each species.

The diversity within domestic livestock species is perceived generally in terms of differences among sub-groups that are referred to as breeds, where "breed" has been defined (Turton, 1974) as "a homogeneous, sub-specific group of domestic livestock with definable and identifiable external characters that enable it to be separated by visual appraisal from other similarly defined groups within the same species, or a homogeneous group where geographical separation from phenotypically similar groups has led to general acceptance of its separate identity". While generally appropriate, breeds often are not distinguished in the developing world. Local populations may have different names,

but without change in phenotype; a change in phenotype may occur without change in name; or all populations may have just one name and be phenotypically similar. In the broad context of global animal genetic resources, the term "breed" is used to include strains and populations, the members of which are distinguished from other such groups in local, national or regional usage. That is, a breed is a cultural entity, recognized as such by the community where it is found.

Animal Breeding and Conservation - A Conflict?

The aim of animal breeding is to change the genetic makeup of domestic animals so that they better meet our needs. Such improvement in production, product quality or the efficiency of production is sought by either or both of selection within breeds or use of differences among breeds through cross-breeding, grading-up to a superior breed by repeated back-crossing, or formation of a synthetic population. Thus future improvement is dependent on genetic variation - both the variation within breeds, and the variation between breeds, and loss of variation will restrict the options available to meet unpredictable future requirements.

Loss of variation within breeds is continually countered by the introduction of new variation through mutation (Franklin, 1981; Hill and Keightley, 1988), but the variation among breeds cannot be readily regenerated. Each breed is the product of mutation and genetic drift, as well as separate adaptation and evolution, with differing selection pressures imposed by climate, endemic parasites and diseases, available nutrition and criteria imposed by man. Each breed thus comprises a unique set of genes.

Yet many breeds have become extinct, and many more are at risk. The global animal genetic resources almost certainly comprise some 4 500 breeds, although the true number is not known. For seven mammalian species (ass, buffalo, cattle, goat, horse, pig and sheep), FAO (1995) lists 2 944 breeds as

Table 1. Numbers of breeds of each of seven major species of domestic livestock that are recorded in the FAO Global Databank for Animal Genetic Resources, and the numbers estimated to be at risk.

| Species | Numbers of breeds | | | Per cent at risk |
|--------------|-------------------|---------------------------------|------------|---------------------|
| | Recorded | With population size data | At risk* | |
| Ass | 77 | 24 | 9 | 37.5 |
| Buffalo | 72 | 55 | 2 | 3.6 |
| Cattle | 787 | 582 | 135 | 23.2 |
| Goat | 351 | 267 | 44 | 16.5 |
| Horse | 384 | 277 | 120 | 43.3 |
| Pig | 353 | 265 | 69 | 26.0 |
| Sheep | 920 | 656 | 119 | 18.1 |
| Total | 2 944 | 2 126 | 498 | 23.4 |

*Estimated from breeds with available population data

recorded in the FAO Global Databank for Animal Genetic Resources, of which 498 are considered to be at risk (Table 1). Extinctions of past breeds and this potential loss of more breeds is due to the demand for increased animal production, with economic pressures (primarily in developed countries) and political and social pressures (primarily in socialized economies and less-developed countries) causing some breeds to be considered unsuitable for today's needs or those of the immediately perceived future. Yet some of these breeds, and particularly those that have evolved in and become adapted to stressful environments, are likely to carry valuable genes and gene combinations controlling specific behavioural, physiological, and disease and parasite resistance traits. The genotypes of some of these breeds could be crucial to the development of sustainable animal production systems in the future.

Thus there is an apparent conflict, which will be countered only by active and effective management of all animal genetic resources.

Conservation History

The realization of the need for conservation of animal genetic resources is not new, and in fact has been on the international agenda for some 50 years (Barker, 1994). Translation of this realization into action has been slow, although there have been notable achievements and recent developments are encouraging. In the developed world, organizations such as the Rare Breed Survival Trust in the UK, Safeguard for Agricultural Varieties in Europe, and the American Livestock Breeds Conservancy in the USA have instituted effective programmes (see Alderson, 1990) for the conservation of rare and endangered breeds.

However, no such organizations and programmes exist, or are likely to be developed in the near future, in the developing world. In any case, the problems there are rather different. Many breeds are endangered, but others that are not numerically small and that are being used for production, are threatened - primarily by being cross-bred with imported breeds that are perceived (often wrongly) to be superior. But these native breeds are likely to be

well-adapted to the traditional husbandry systems. Thus primary emphasis needs to be given to more effective use and genetic improvement of these breeds within the prevailing and generally sustainable production systems.

FAO has for some 25 years taken a major role in promoting awareness of the significance to mankind of the global animal genetic resources (e.g. FAO, 1984, 1992). There is now a clear recognition (FAO, 1993) that "in the global management of animal genetic resources, the fundamental distinction is **not** between those breeds that are endangered and those that are not, but between those that are perceived to have little or no current utility and those which do have current utility or seem likely to have in the immediate future. For each of these latter categories, the necessary actions are then **preservation** (as live animals or frozen storage of embryos or semen) or **utilization** (including development of breeding programmes for genetic improvement)." Given this recognition, the apparent conflict between animal breeding and conservation is resolved; successful management of animal genetic resources must incorporate both utilization and conservation.

In November 1995, the FAO Conference of member governments made two major decisions in relation to animal genetic resources: (i) it provided an intergovernmental mechanism for animal genetic resources by broadening FAO's long-established Commission on Plant Genetic Resources to a Commission on Genetic Resources for Food and Agriculture, and (ii) it supported as a priority activity for FAO a Global Strategy for the Management of Farm Animal Genetic Resources. Most importantly, the Global Strategy is being designed to harmonize fully with the UN Convention on Biological Diversity, the international law which is now ratified for use by more than 170 countries. FAO is funding the essential core activities of the Strategy (coordinating and facilitating regional and national programmes, maintaining the Global Databank and early warning system,

developing technical guidelines for use by countries in establishing cost-effective action and reporting on all activities), although very substantial funding from sources external to FAO will be required for full implementation of all aspects. Nevertheless, with the direct involvement of individual countries, other international agencies, including the International Agricultural Research Centres, and non-governmental organizations including farmer associations, implementation of the Strategy has been initiated. Its success will be vital to the future of livestock breed diversity.

Defining the Problem

Ideally, consideration of the conservation of animal genetic resources would start with complete information on all existing breeds - numbers, distribution and population structure, trends in numbers (increasing, stable or decreasing), productive performance and adaptive characters. That ideal is not even within reach - the best information available is that there are some 4 500 breeds, of which some 30% are estimated to be at risk of loss (FAO, 1995). For most breeds, even basic data on population numbers and trends are not available. Of the 2 944 breeds of seven mammalian species that are listed in the FAO Global Databank, 818 (28%) do not have any population data. This proportion of 28% is certainly an underestimate of the global situation, as breeds not yet listed are primarily from developing countries where census data is less likely to be available (FAO, 1995).

Clearly the first problem is lack of information, so the identification and characterization of all breeds of livestock has high priority in the FAO global strategy, and must also have high priority for all national livestock development programmes (Barker, 1992). Once available, this documentation of existing resources would identify those breeds at risk of extinction. However, census data alone will not provide a basis for the rational choice of breeds for development programmes. Identification at the global level

Table 2. *F*-statistics for swamp and river buffalo estimated using 25 polymorphic protein coding loci and 21 microsatellite loci.

| | F_{IS} | F_{ST} | F_{IT} |
|------------------------|-------------|------------|------------|
| <u>Protein Coding</u> | | | |
| Swamp | -.004(.062) | .182(.041) | .181(.083) |
| River | -.068(.048) | .108(.036) | .048(.059) |
| <u>Microsatellites</u> | | | |
| Swamp | .047(.027) | .168(.018) | .207(.034) |
| River | .031(.028) | .038(.008) | .068(.029) |

Table 3. Correlation coefficients among genetic distances estimated from data on 21 microsatellite loci or 25 protein coding loci.

| | Microsatellites | | | Protein Coding | | |
|----------|-----------------|-------|----------|----------------|-------|-------|
| | Nei D | D_A | Delta mu | Reynolds | Nei D | D_A |
| Reynolds | .921 | .911 | .748 | .831 | .773 | .798 |
| Nei D | | .991 | .890 | .754 | .774 | .772 |
| D_A | | | .897 | .748 | .771 | .783 |
| Delta mu | | | | .738 | .797 | .772 |
| Reynolds | | | | | .929 | .965 |
| Nei D | | | | | | .937 |

All significant, $P < 0.001$

of priority breeds for immediate development was one of the major questions addressed by an FAO Expert Consultation (FAO, 1992). The criteria for choice of such breeds were specified as:

1. the breed possesses one or more highly desirable attributes in terms of productivity and/or adaptation,
2. the breed is endangered, or is not being utilized efficiently,
3. the breed should be one whose improvement could have the potential to influence large populations, either of the same breed in one or more countries, or other very similar breed types.

Again, there is an assumption of information being available with regard to "desirable attributes", but for many breeds, particularly in developing regions, this will be true, and pragmatic and subjective decisions will have to be made.

In relation to the total global biodiversity, animal production encompasses a small and finite universe - only some 4 500 breeds across some 40 species, and the total genetic variation within each species. There is a critical need to develop breeding programmes to improve the production and productivity of a selected set of breeds that have major utility now or which seem likely to have in the immediate future. As already noted, selection of these breeds must be somewhat *ad hoc*, and the best choices may not be made. Thus breed evaluation studies are needed to compare performance, and the underlying genetics and biology, so that future choices will be more soundly based. Further, with increasing emphasis on sustainable production systems, on animal welfare and on pollution, future breeding objectives may differ quite dramatically from those now

considered most important. Thus there is the need to prevent loss of breed diversity (i.e. conservation of endangered breeds).

But given all of these needs, the relatively small and finite universe of animal production is simply not manageable. On both country and global scales, financial and other resources are limited; it will not be possible to maintain every breed that is in danger of extinction, nor to develop breeding programmes for all breeds considered suitable for improvement, nor to set up evaluation studies of the many breeds that might contribute to future breeding programmes.

For each of these different aspects of animal genetic resources conservation, priorities will have to be defined in choosing those breeds to be included.

Setting Priorities for Conservation

In the context of setting priorities for conservation, Miller (1977) suggested that the only rational criteria are the likelihood of extinction of a breed and the degree to which it possesses unique genes. Further, he suggested that immunogenetic and other biochemical polymorphism data should provide the basis for determining whether a breed is sufficiently unique (i.e. sufficiently different) from other breeds.

With emphasis on the need for planning of comparative evaluation studies, Barker (1980, 1985) suggested that the genetic relationships among breeds within each species of livestock should be determined, so that the breeds could be grouped into sets that are genetically similar, with one representative from each set then included in an evaluation study.

Both of these suggestions relate to different aspects of what is now broadly interpreted as conservation, and the concepts of genetic uniqueness and genetic relationships have become accepted as the basis for setting priorities for conservation (FAO, 1993; Barker, 1994). Inherent in this is the practical realization that breeds that are taxonomically

distinct should be favoured for conservation, and that the objective is to minimize loss of genetic diversity.

An objective quantification of the magnitude of the genetic differences among a set of breeds can be obtained from allele frequency data for each breed, estimating genetic distances between each pair of breeds, and by constructing from these pair-wise distances a diagram that best represents all relationships among the populations, i.e. a phylogenetic tree. The topology of the tree shows the patterns of relationships, while branch lengths indicate the magnitude of the differences between breeds. The problem then is how to use this information to ensure maintenance of maximum genetic diversity, or realistically to ensure minimum loss of genetic diversity, given economic, social and other constraints. For non-domestic species, the analogous problem of optimizing reserve selection so that the maximum species diversity is preserved has been considered recently by several workers (see Witting and Loeschcke, 1995). Methods based on phylogenetic relationships, on genetic divergence or on both have been developed, and these have been compared by Krajewski (1994). He shows that the different measures of taxonomic diversity agree in identifying the species that contribute the most and the least to overall diversity, but disagree at intermediate levels. Although no consensus has emerged as to the relative merits of these measures, some of the complications in dealing with sets of species that are outlined by Krajewski (1994) are not relevant to the breed conservation issue. Here all breeds in one species is the set to be considered, and the diversity is specified in terms of genetic differences, i.e. allele frequencies translated to genetic distances. Thus a method combining topology and divergence information is likely to be preferred. In addition, as the objective is to minimize loss of genetic diversity in each species, within-breed genetic variation must be considered, perhaps by being incorporated into a diversity measure as suggested by

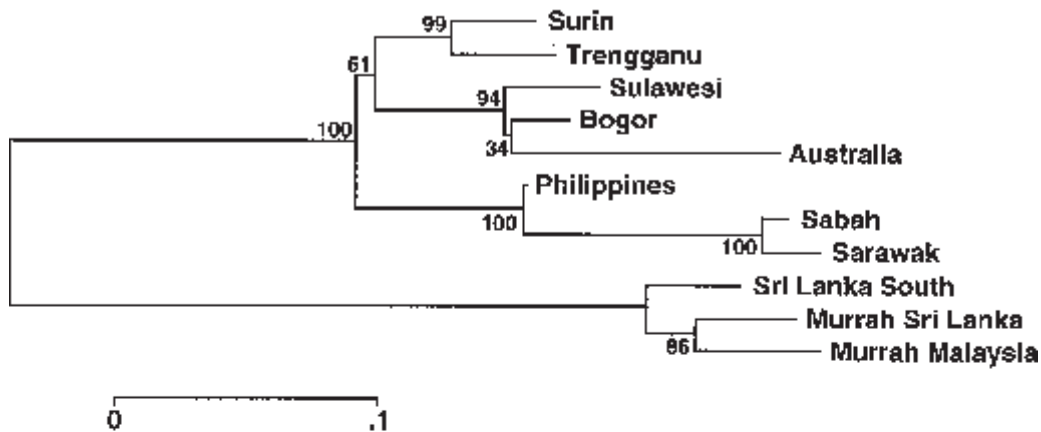
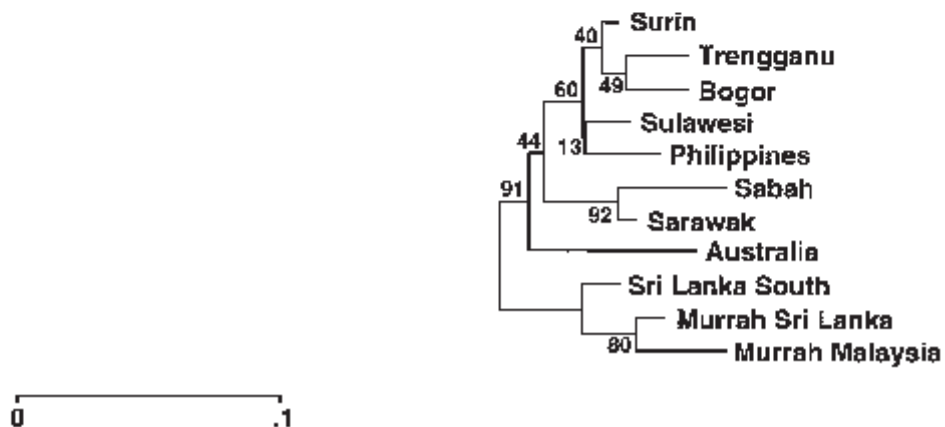
(A) Microsatellites**(B) Protein coding loci**

Figure 1. Neighbour-joining trees constructed using the DA distance measure for (a) microsatellite loci and (b) protein coding loci.

Krajewski (1994). However, even without such incorporation, the breed with highest average heterozygosity should be preferred in choosing among breeds that otherwise have equal priority.

In setting conservation priorities for livestock breeds, genetic distances and phylogenetic diversity will provide the best objective criterion. However, distance measures cannot account for consequences of

artificial selection on morphological or economic traits, nor for natural selection on fitness, and thus will give only the first (albeit essential) guide in making conservation decisions. Where decisions regarding conservation are to be made among a set of rare or endangered breeds, phylogenetic diversity will be the major criterion. In the choice of breeds for development programmes or for comparative evaluation studies, final decisions should also take into account any available data on traits of

economic value, specific adaptive features, presence of unique genes or genotypes, local or regional importance of a breed in production systems, and availability of resources and infrastructure in the region where a breed is located (FAO, 1993).

However, before any priorities can be set, we first need the distance estimates. Some 100 studies of genetic distances among livestock breeds have been made (Barker, 1994), but these are not sufficient to provide a global overview that would be a basis for conservation decisions. Hence FAO, as part of its Strategy for the Management of Farm Animal Genetic Resources, has planned a global research initiative to characterize genetic diversity in each livestock species.

In 1993, an FAO Working Group concluded that a global project for estimating genetic distances among the breeds of each species of domestic livestock was feasible, and the design and procedures were outlined (FAO, 1993). Another FAO Working Group, supported by some 20 international experts, has now developed the details of this global project for the measurement of domestic animal genetic diversity, referred to as the MoDAD project (FAO, 1996). All 4 500 breeds clearly cannot be included, and a two-phase strategy is proposed, with appropriate selection of breeds in phase 1 to evaluate the range of diversity within each species. Phase 2 for each species would depend on results obtained in phase 1, but could for example, include analyses of additional breeds from regions showing the greatest diversity.

Measuring Diversity

Many phylogenetic studies have used allele frequencies of biochemical polymorphisms (e.g. Nei, 1987 - Table 9.3). However, microsatellite markers are being used increasingly in population and evolutionary genetics (Bruford and Wayne, 1993). They have essentially replaced biochemical polymorphisms as the markers of choice for

such studies, because their higher average per locus heterozygosity and potentially many more loci are expected to provide higher resolution discrimination among closely related populations of a species.

Microsatellites are the markers of choice for gene mapping studies in livestock species, so that many loci will be available for use in phylogenetic studies, and some studies of relationships among livestock breeds already have used microsatellite markers, e.g. cattle - MacHugh *et al.* (1994), sheep - Buchanan *et al.* (1994), pigs - van Zeveren *et al.* (1995) and buffalo - Barker *et al.* (1997b). They are to be used in the FAO MoDAD project.

While microsatellites have apparent advantages as markers for genetic distance studies (Meghen *et al.*, 1994; Hall and Bradley, 1995), our knowledge of their evolution is inadequate to be certain that they are representative of the genome. Most are in non-coding regions, and subject to genetic turnover mechanisms (Amos and Hoelzel, 1992), mutation mechanisms are not clear and may differ for different types of microsatellites (Estoup *et al.*, 1995), loci with large numbers of alleles may be subject to high mutation rates and show departures from Mendelian segregation, while undetected non-amplifying (null) alleles (Pemberton *et al.*, 1995) would cause errors in allele frequency estimates and hence in estimated genetic distances.

The question then is whether microsatellite markers will give the same phylogeny as a set of real genes. In particular, would they give the same phylogeny as one (impossible to obtain!) based on allele frequencies at the larger set of genes controlling important productive, reproductive and adaptive traits? In fact, they may not, and it should not be expected that they would because two different questions are being addressed. When genetic distances are estimated and used to construct a phylogeny of breed relationships, the aim is to describe similarities that are due to common ancestry. Thus the markers used should be selectively neutral. In contrast, the genes controlling traits of interest to animal

breeders will not be selectively neutral, and breeds may be genetically similar at these loci because of convergence due to similar selection pressures, or dissimilar because of differential selection. As emphasized earlier, distance measures describing similarities due to common ancestry should be used only as an initial criterion in making breed conservation decisions.

An Example

Nevertheless, it is of interest to determine if microsatellite markers will give the same phylogeny as a set of real genes. A possible approach to this is to compare phylogenies based on biochemical and microsatellite markers, as has been done for water buffalo populations in southeast Asia (Barker *et al.*, 1997a,b). Seventeen populations (12 swamp type, 2 river (Murrah breed) and 3 Lankan buffalo (genetically river type)) were sampled and a total of 801 animals assayed for 53 protein coding loci, 25 of which were polymorphic. A subset of 261 animals from 11 of these populations were genotyped for 21 polymorphic microsatellite loci. Only the results for the 11 populations that were assayed for both protein coding and microsatellite loci will be discussed here, with results based only on the 25 polymorphic loci for the former.

F-statistics estimates (Table 2) show no significant departures from Hardy Weinberg expectation for either buffalo type, whether based on protein coding loci or on microsatellites. All estimates of F_{ST} (population differentiation), however, were significantly greater than zero, and for swamp buffalo (8 populations), estimates from protein coding and microsatellite loci were very similar. For river buffalo, the F_{ST} estimated from protein coding loci was about three times that from microsatellites, but not significantly different.

For both protein coding and microsatellite loci, the standard genetic distance of Nei (1978), the D_A distance of Nei *et al.* (1983) and Reynolds' distance (Reynolds *et al.*, 1983) were estimated, while for

microsatellite loci only, the $(\delta\mu)^2$ distance of Goldstein *et al.* (1995) also was estimated. Correlation coefficients among these measures (Table 3) were all highly significant ($P < 0.001$), although the correlations among measures using microsatellites only or protein coding loci only were higher than those between microsatellites and protein coding loci.

Neighbour-joining trees (Saitou and Nei, 1987), constructed using the D_A distance measure for each of microsatellites and protein coding loci (Figure 1), show differences in both topology and branch lengths. In both trees, some of the nodes are not strongly supported and although the numbers of loci used are not small (21 microsatellite and 25 protein coding), assay of larger numbers of loci could well change the patterns of relationships. Clearly it is not possible to confirm which tree is a better representation of the true genetic relationships among these populations, but the microsatellite tree does accord better with the geography of the populations and the known history of the Australian population, which descends from a small number of animals imported from Timor some 160 years ago. The microsatellite tree is most likely a better representation of the similarities due to common ancestry, while the protein coding loci tree is distorted, reflecting additional effects due to bottlenecks in some populations and selection at some loci.

Thus these results provide empirical support for the view that phylogenetic diversity based on microsatellite loci, as in the proposed FAO MoDAD project, will provide the best objective criterion for making initial conservation decisions for livestock breeds.

Acknowledgements

The water buffalo studies were supported by the Australian Centre for International Agricultural Research (Project No. 8364, and a Small Grant for the microsatellite analyses), and could not have been done without the active collaboration of many colleagues,

particularly Dr. S.G. Tan, Mr. O.S. Selvaraj, Professor T.K. Mukherjee, Dr. S.S. Moore and Dr. D.J.S. Hetzel.

References

- Alderson L.** (Ed.) 1990. Genetic Conservation of Domestic Livestock. CAB International, Wallingford.
- Amos B. & Hoelzel A.R.** 1992. Applications of molecular genetic techniques to the conservation of small populations. *Biol. Conserv.*, 61: 133-144.
- Barker J.S.F.** 1980. Animal genetic resources in Asia and Oceania - The perspective, pp. 13-19 in Proc. SABRAO Workshop on Animal Genetic Resources in Asia and Oceania. Tropical Agriculture Research Center, Tsukuba, Japan.
- Barker J.S.F.** 1985. Identifying the breeds to be evaluated, in Evaluation of Large Ruminants for the Tropics : proceedings of an International Workshop held at CSIRO, Rockhampton, Qld, Australia, 19-23 March, 1984, edited by J.W. Copland. ACIAR Proceedings, 5: 161-166.
- Barker J.S.F.** 1992. Proposals for an action program for animal genetic resources in the AAAP region. Proc. 6th AAAP Animal Science Congress, 1: 229-238.
- Barker J.S.F.** 1994. Animal breeding and conservation genetics, pp. 381-395 in Conservation Genetics, edited by V. Loeschcke, J. Tomiuk and S.K. Jain. Birkhäuser Verlag, Basel.
- Barker J.S.F., Tan S.G., Selvaraj O.S. & Mukherjee T.K.** 1997a. Genetic variation within and relationships among populations of Asian water buffalo (*Bubalus bubalis*). *Animal Genetics*, 28: 1-13.
- Barker J.S.F., Moore S.S., Hetzel D.J.S., Evans D., Tan, S.G. & Byrne, K.** 1997b. Genetic diversity of Asian water buffalo (*Bubalus bubalis*): Microsatellite variation and a comparison with protein-coding loci. *Animal Genetics*, 28: 103-115.
- Bruford M.W. & Wayne R.K.** 1993. Microsatellites and their application to population genetic studies. *Current Opinion in Genetics and Development*, 3: 939-943.
- Buchanan F.C., Adams L.J., Littlejohn R.P., Maddox J.F. & Crawford A.M.** 1994. Determination of evolutionary relationships among sheep breeds using microsatellites. *Genomics*, 22: 397-403.
- Estoup A., Garnery L., Solignac M. & Cornuet J-M.** 1995. Microsatellite variation in honey bee (*Apis mellifera* L.) populations: hierarchical genetic structure and test of the infinite allele and stepwise mutation models. *Genetics*, 140: 679-695.
- FAO** 1984. Animal genetic resources conservation by management, data banks and training. FAO Animal Production and Health Paper 44/1. FAO, Rome. pp. 186.
- FAO** 1992. The management of global animal genetic resources. FAO Animal Production and Health Paper 104. FAO, Rome. pp. 309.
- FAO** 1993. An integrated global programme to establish the genetic relationships among the breeds of each domestic animal species. FAO Division of Animal Production and Health, Report of a Working Group. Mimeo, pp. 32.
- FAO** 1995. World Watch List for Domestic Animal Diversity. 2nd Ed. FAO, Rome.
- FAO** 1996. Global project for the measurement of domestic animal diversity (MoDAD). FAO Animal Production and Health Paper, Rome.

- Franklin I.R.** 1981. Population size and the genetic improvement of animals. In *Future Developments in the Genetic Improvement of Animals*, edited by J.S.F. Barker, K. Hammond and A.E. McClintock. Academic Press Australia, Sydney, 181-196.
- Goldstein D.B., Linares A.R., Cavalli-Sforza L.L. & Feldman M.W.** 1995. An evaluation of genetic distances for use with microsatellite loci. *Genetics*, 139: 463-471.
- Hall S.J.G. & Bradley D.G.** 1995. Conserving livestock breed biodiversity. *Trends in Ecology and Evolution*, 10: 267-270.
- Hill W.G., & Keightley P.D.** 1988. Interrelations of mutation, population size, artificial and natural selection. In *Proc. Second Internat. Conf. Quant. Genet.*, edited by B.S. Weir, E.J. Eisen, M.M. Goodman and G. Namkoong. Sinauer, Sunderland, 57-70.
- Krajewski C.** 1994. Phylogenetic measures of biodiversity: a comparison and critique. *Biol. Conserv.*, 69: 33-39.
- MacHugh D.E., Loftus R.T., Bradley D.G., Sharp P.M. & Cunningham P.** 1994. Microsatellite DNA variation within and among European cattle breeds. *Proc. Roy. Soc. Lond. B*, 256: 25-31.
- Meghen C., MacHugh D.E. & Bradley D.G.** 1994. Genetic characterization and West African cattle. *World Anim. Rev.*, 78: 59-66.
- Miller R.H.** 1977. The need for and potential application of germplasm preservation in cattle. *J. Hered.*, 68: 365-374.
- Nei M.** 1978. Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics*, 89: 583-590.
- Nei M.** 1987. *Molecular Evolutionary Genetics*. Columbia University Press, New York.
- Nei M., Tajima F. & Tateno Y.** 1983. Accuracy of estimated phylogenetic trees from molecular data II. Gene frequency data. *J. Mol. Evol.*, 19: 153-170.
- Pemberton J.M., Slate J., Bancroft D.R., & Barrett J.A.** 1995. Non-amplifying alleles at microsatellite loci: a caution for parentage and population studies. *Molec. Ecol.*, 4: 249-252.
- Reynolds J., Weir B.S. & Cockerham C.C.** 1983. Estimation of the co-ancestry coefficient: basis for a short term genetic distance. *Genetics*, 105: 767-779.
- Saitou N. & Nei M.** 1987. The neighbour-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.*, 4: 406-425.
- Turton J.** 1974. The collection, storage and dissemination of information on breeds of livestock. *Proc. 1st Wld Cong. Genet. Appl. Livestock Prod.*, 2: 61-74.
- Witting L. & Loeschcke V.** 1995. The optimization of biodiversity conservation. *Biol. Conserv.*, 71: 205-207.
- van Zeveren A., Peelman L., van de Weghe A. & Bouquet Y.** 1995. A genetic study of four Belgian pig populations by means of seven microsatellite loci. *J. Anim. Breed. Genet.*, 112: 191-204.

The development of a system of linear measurements to provide an assessment of type and function of beef cattle

G.L.H. Alderson

Rare Breeds International, Avenue Q, N.A.C., Stoneleigh Park, Warks CV8 2LG, UK

Summary

Linear measurements have been used as indicators of weight in cattle. Withers height has been used most commonly, but measurements of White Park cattle show that it is of limited value. This paper explores alternative measurements and indices, not only to provide a superior guide to weight, but also as indicators of type and function of beef cattle. A complex index is recommended for breed characterisation, while hip width or rump length are shown to be the preferred single measurements for evaluation of individual animals or within-herd groups.

Resumen

Las medidas lineales se han usado como indicadores del peso en el vacuno. La alzada a la cruz ha sido la variable utilizada más comunmente, aunque sus valores en la raza White Park Cattle se han mostrado de utilidad limitada. Este artículo explora variables alternativas e índices, no sólo para proveernos de una mejor guía del peso, sino también de un indicador del tipo y de la función del vacuno de carne. Se recomienda un índice complejo para la caracterización racial, mientras que la anchura de la cadera y la longitud de la grupa se muestran como las variables simples más adecuadas para la evaluación de animales individuales o de grupos dentro del rebaño.

Key words: *Body measurement, Linear measurement, Cumulative index, White Park cattle.*

Introduction

Morphological descriptions of beef cattle have been used either to indicate breed origins and relationships through the medium of head measurements (Jewell 1963), or to indicate size and weight through measurement of height at withers. EAAP and FAO used withers height as a prime indicator of type (Simon and Buchenauer 1993). More recently rump height has been preferred to withers height, especially for describing cattle in beef show classes. In addition, girth circumference has been used as a basis for the calculation of weight. Since 1994, a group within the International Committee for Animal Recording has been established to study conformation recording, but its work has been directed mainly at dairy cattle (Stoll *et al.*, 1996).

Height, at either withers or rump, has limited value as an indicator of weight, and it was evaluated as a preliminary exercise in this study. Similarly, the accuracy of girth as an indicator of weight is low, and Schwabe used a more complex formula involving girth, height and body length measurements (Schwabe and Hall 1989). The limited value of both height and girth as single measurements, and the lack of a tested empirical alternative, has restricted the ability of breeders to assess type in beef cattle and their value as potential breeding stock, and has limited the opportunity to provide potential purchasers with a reliable evaluation of animals. This paper is concerned with measurements associated with production characteristics. The development of a new system has been explored to enable earlier assessment of breeding animals and, by

establishing a standard format, to enable comparison of individual animals against a breed standard. The study determined the value of each measurement, and devised indices of type and function.

Method

Eight separate measurements were used for females, and nine for males. The common measurements were:

Height at withers, height at rump (tuber coxae), body length (withers to pins), length from withers to hips (tuber coxae), rump length from hips (tuber coxae) to pins (tuber ischii), width of hips (tuber coxae), width of pins (tuber ischii), width of chest (widest point of shoulders) and depth of chest (thorax) immediately behind forelegs.

The extra measurement for bulls was scrotal circumference.

The equipment was a measuring stick (as used for horses) to measure withers height, rump height, body length and length from withers to hip; a measuring stick modified with sliding caliper to measure hip width, pin width, chest width, chest depth and rump length; and an adapted metal tape to measure scrotal circumference.

The measurements were taken by the same fieldsman using a standard form; a separate form was prepared for each animal. The measurements from the standard form were transferred to a master sheet which calculated the appropriate indices for each animal and the mean, SD and coefficient of variation for the group of animals included in the study. Two measurements were not used in subsequent analyses. Length from withers to hip was intended initially as a check on body length and rump length measurements, but it proved unreliable. Width of pins was measured on the points of the bone rather than the outer dimension, and again this proved unreliable.

The calculated indices were: weight, height slope, length index (2 indices), rump length index, balance, width slope, depth index, foreleg length and cumulative index (3 indices). In all cases the results for individual animals were indexed against the average for the group. The calculations are reported in table 1.

The system was tested on White Park cattle in the United Kingdom in 1994-5, and was repeated in 1997. Both sexes were included, comprising 315 cow records and 29 bull records. Some animals were included in both phases of the study. The ages of the females ranged from 15 months to 201

Table 1. Results of the calculations for individual animals indexed against the average for the group.

-
- Weight: $\text{Body length} \times \text{girth depth} \times [(\text{hip width} + \text{chest width})/2]/1050$
 - Height slope: $\text{rump height} - \text{withers height}$
 - Length index ⁽¹⁾: $\text{body length}/\text{chest (thorax) depth}$
 - Length index ⁽²⁾: $\text{body length}/\text{withers height}$
 - Rump length index: $\text{rump length}/\text{withers to hip length}$
 - Balance: $(\text{hip width} \times \text{rump length})/(\text{chest depth} \times \text{chest width})$
 - Width slope: $\text{hip width} - \text{chest width}$
 - Depth index: $\text{chest depth}/\text{withers height}$
 - Foreleg length: $\text{withers height} - \text{chest depth}$
 - Cumulative ⁽¹⁾ index : $(\text{weight}/\text{breed average weight}) + [(\text{length index 1} + \text{balance})/2]$
 - Cumulative ⁽²⁾ index : $(\text{weight}/\text{breed average weight}) + [(\text{length index 2} + \text{balance})/2]$
 - Cumulative ⁽³⁾ index : $(\text{weight}/\text{breed average weight}) + \text{length index 2} + \text{balance}$
-

months, and of the males from 12 months to 133 months; almost all animals were in unfitted (i.e. not Show) condition.

Results

The results were expressed as:

1. mean, SD, coefficient of variation and range for each linear measurement
2. graphs to show the development with age for each linear measurement
3. coefficient of correlation matrix for linear measurements, weight and cumulative index
4. histogram profile charts for individual animals and for individual animals with sire and dam for linear measurements, weight and cumulative index.

Evaluation of measurement of height at withers and rump was undertaken as a preliminary analysis. The measurements were easy to apply and showed high repeatability. Height at withers was correlated most closely with length of foreleg (0.760), but both height at withers and rump had a lower correlation with weight and cumulative(3) index than any of the other linear measurements. Height was an unreliable guide to weight, and lacked value as an indicator of type or functionality, except possibly at extremes of the frequency

distribution curve. It was determined that the identification and assessment of alternative indicators was necessary.

The results indicate that White Park cows reach their maximum weight at approximately nine years of age, but that they reach 98-99% of maximum size (linear measurements) by five years of age. The results for mature cows were calculated from data relating to animals of 60 months of age or more. The measurements (Table 2) and indices (Table 3) show the averages for the breed and indicate the typical structure of White Park cattle.

Each measurement developed at a different rate at different ages (Table 4). Rump height developed most rapidly, reaching 93-94% of full size by 24 months of age. Chest depth developed more slowly, reaching 97.8% of full size by 60 months of age. Most other measures progressed at a similar rate (86-90% at 24 months, 92.5-94.5% by 36 months, circa 96% by 42 months, and 99% by 60 months). The development of hip width was unusual. It developed most slowly to 24 months of age, but reached 99.4% by 60 months of age. Leg length (calculated by deducting chest depth from withers height) reached full size by 12 months of age. Chest width was the most variable measurement

Table 2. Linear measurements of mature cows over 60 months of age.

| Measurement | Mean (cm) | S.D. | Cof. V. | Max (cm) | Min (cm) |
|----------------|-----------|------|---------|----------|----------|
| Withers height | 131.13 | 5.31 | 4.05 | 145 | 121 |
| Rump height | 131.91 | 5.12 | 3.88 | 146 | 120 |
| Length of body | 147.57 | 6.62 | 4.49 | 169.5 | 131 |
| Rump length | 55.41 | 3.57 | 6.44 | 68 | 47 |
| Width of hips | 59.66 | 3.81 | 6.39 | 72 | 51 |
| Width of chest | 51.37 | 5.94 | 11.57 | 71 | 41 |
| Depth of chest | 75.36 | 3.46 | 4.59 | 88 | 62 |

Table 3. Indices of mature cows over 60 months of age.

| Index | Mean | SD | Cof.V. | Max | Min |
|---------------------|--------|-------|--------|------|------|
| Weight (kg) | 619.43 | 96.13 | 15.52 | 957 | 399 |
| Length 1 | 1.97 | 0.09 | 4.36 | 2.08 | 1.72 |
| Length 2 | 1.11 | 0.05 | 4.31 | 1.26 | 0.96 |
| Rump length | 0.59 | 0.05 | 7.73 | 0.83 | 0.49 |
| Balance | 0.86 | 0.08 | 9.08 | 1.11 | 0.66 |
| Depth | 0.56 | 0.03 | 4.72 | 0.64 | 0.49 |
| Foreleg length (cm) | 56.31 | 4.09 | 7.26 | 66 | 45 |
| Cumulative 1 | 2.47 | 0.15 | 6.28 | 2.89 | 2.09 |
| Cumulative 2 | 2.04 | 0.16 | 7.84 | 2.55 | 1.69 |
| Cumulative 3 | 3.88 | 0.17 | 4.45 | 4.37 | 3.38 |

Table 4. Development of cows at 42 and 60 months of age relative to full size measured by two methods (a. and b.)

| Measurement | 42 months of age | | | 60 months of age | | |
|---------------|------------------|------|-------|------------------|------|-------|
| | a. | b. | av. | a. | b. | av. |
| Wither height | 96.2 | 96.9 | 96.55 | 98.8 | 99.1 | 98.95 |
| Rump height | 98.1 | 98.1 | 98.10 | 99.4 | 99.3 | 99.35 |
| Body length | 95.8 | 94.2 | 95.00 | 99.1 | 97.4 | 98.25 |
| Rump length | 96.1 | 95.2 | 95.65 | 99.5 | 97.4 | 98.45 |
| Hip width | 95.5 | 93.9 | 94.70 | 99.4 | 99.3 | 99.35 |
| Chest width | 95.4 | 92.1 | 93.75 | 98.9 | 97.9 | 98.40 |
| Chest depth | 94.5 | 93.9 | 94.20 | 97.8 | 98.2 | 98.00 |
| Weight | 86.1 | 82.2 | 84.15 | 95.2 | 94.4 | 94.80 |

a) total sample development curves

b) age group direct comparisons

Table 5. Development of cows at 42 and 60 months of age relative to conformation of mature cows.

| Index | 42 months of age | | | 60 months of age | | |
|----------------|------------------|-------|--------|------------------|-------|--------|
| | a | b | av | a | b | av |
| Weight | 102.8 | 98.2 | 100.50 | 101.6 | 100.9 | 101.25 |
| Length 1 | 101.4 | 100.3 | 100.85 | 100.8 | 99.3 | 100.05 |
| Length 2 | 98.7 | 97.2 | 97.95 | 99.4 | 98.3 | 98.85 |
| Rump length | 102.5 | 101.7 | 102.10 | 101.3 | 99.2 | 100.25 |
| Balance | 103.4 | 103.2 | 103.30 | 101.9 | 100.8 | 101.35 |
| Depth | 95.9 | 96.9 | 96.40 | 98.9 | 98.9 | 98.90 |
| Foreleg length | 100.2 | 100.9 | 100.55 | 100.1 | 100.6 | 100.35 |
| Cumulative 1 | 101.6 | 99.9 | 100.75 | 101.1 | 100.3 | 100.70 |
| Cumulative 2 | 100.5 | 98.9 | 99.70 | 100.1 | 100.2 | 100.15 |
| Cumulative 3 | 101.1 | 100.4 | 100.75 | 100.5 | 100.1 | 100.30 |

(coefficient of variation of 11.57%) and this reflected the sensitive response of chest width to changes in the fitness (condition) of an animal. Other measurements had a coefficient of variation between 3.88% and 6.44%.

The linear measurements were used to calculate indices, which show the structure and proportions of each animal. The indices for all females in the study (Table 3) show the highest coefficient of variation for balance, rump length index, and foreleg length, and the lowest for both length indices, depth index and cumulative(3) index. Weight was markedly variable (coefficient of variation 15.52%) and this again reflected the variety of environmental effects and the condition of each animal. Indices remained relatively constant at each stage of development from 12 months of age (younger animals were not included in the study). A detailed comparison of indices (Table 5) in 42 month and 60 month animals, compared with fully mature animals, showed that only length(2) index and depth index varied with age, and this resulted from the slower development of chest depth (Table 3). All other indices were within 1.7% of the expected norm at 60 months of age and within 4.1% at 42 months of age.

Both linear measurements and indices for different age groups (Tables 4 and 5) were calculated by two methods. The first (a.) was derived from a best curve based on data of the total study sample; the second (b.) was calculated from averages for each age group (42 month - 40-44 months; 60 month - 58-62 months). The results for indices were tested against coefficient of regression data. In all cases the slope of the regression line was negligible (<0.06% per month of age). The steepest slope was for depth index, followed by length(2) index. Cumulative(3) index and balance showed no regression.

The coefficient of correlation was calculated between each linear measurement and weight and cumulative(3) index (Table 6).

The results were also produced on histogram profile charts, either for individual animals or for an animal in conjunction with

its sire and dam, incorporating linear measurements and weight and cumulative(3) index.

Discussion

The linear measurements and, more particularly, the indices, established norms for breed type and conformation. They showed that White Park cows are short-legged animals with a good girth; long-bodied with good rump length; and are active, grazing beef animals. They have the proportions of beef animals (Table 7), although not typical of either the traditional British beef animal such as the Beef Shorthorn or Aberdeen Angus, or continental breeds such as the Charolais or Limousin. They are efficient grazing animals, adapted to rough grazing, and the non-breeding animals produce high quality beef, with excellent flavour, low fat and good marbling (Alderson 1997).

It was not possible to demonstrate the degree of variation within breed due to genetic effects. Blott (1997) indicated a relatively high level of homozygosity in the breed, but Royle (1983) showed relatively high heterozygosity, and anecdotal opinion suggests different types in the breed. For example, breeders claim that the Dynevor type based on Dynevor Raven is long-bodied and 'scopy', while the Chartley type based on Dynevor Samson is more compact, and the linear assessment results do confirm some differences. There were specific differences - for example mature Dynevor type cows stand as high or higher at the rump than at the withers, whereas mature Chartley type cows are the opposite (Table 8) - but other indices, such as depth, width slope and balance, which seem to indicate variation, probably are more a reflection of environmental effects and the more fertile grazing available to the Chartley herd. Indices for length did not show significant variation. An undefined proportion of the variation for each measurement or index was due to human error or to environmental effects of management systems.



Figure 1. These two cows are the same weight and have the same chest depth, but the difference in withers height is 11 cm.

Table 6. Correlation matrix for primary measurements, weight and cumulative(3) index: results for all cows above diagonal, and for cows over 60 months of age below diagonal.

| | Weight | Withers ht | Rump ht | length | Rump lg | Hip width | Chest wi | Chest de | cum ind |
|----------------|--------|---------------|------------|--------|------------|--------------|-------------|-------------|------------|
| Weight | | 0.693 | 0.661 | 0.862 | 0.815 | 0.918 | 0.849 | 0.924 | 0.6 |
| Withers ht | 0.566 | | 0.902 | 0.721 | 0.627 | 0.644 | 0.399 | 0.707 | 0.4 |
| Rump ht | 0.621 | 0.903 | | 0.652 | 0.632 | 0.613 | 0.431 | 0.655 | 0.4 |
| Length | 0.726 | 0.616 | 0.569 | | 0.708 | 0.771 | 0.551 | 0.771 | 0.6 |
| Rump Length | 0.745 | 0.469 | 0.535 | 0.522 | | 0.779 | 0.636 | 0.795 | 0.6 |
| Hip Width | 0.861 | 0.424 | 0.496 | 0.535 | 0.654 | | 0.751 | 0.837 | 0.6 |
| Chest Width | 0.864 | 0.259 | 0.369 | 0.396 | 0.632 | 0.761 | | 0.711 | 0.5 |
| Chest Depth | 0.861 | 0.594 | 0.628 | 0.528 | 0.683 | 0.644 | 0.668 | | 0.5 |
| Cum 3 Index | 0.818 | 0.446 | 0.481 | 0.805 | 0.781 | 0.809 | 0.561 | 0.611 | |

Cumulative(3) index is the preferred measure of the overall value of an animal, although it does not distinguish between different types of conformation. It has a neutral correlation with age, and thus can be used as a measure in young animals to predict mature rating. It is a superior measure to weight because it incorporates measures of desirable conformation, namely length and balance. The results of coefficient of regression calculations reinforced the value of cumulative(3) index on young animals as an indicator of mature type and conformation. It is influenced to some degree by management, but to a much smaller degree than weight.

However, it is a complex index requiring five measurements on each animal. Simpler alternatives are provided by single measurements. Results for cows (Table 6) show that body length, rump length and hip width have the highest correlation with cumulative(3) index. Correlation with weight shows that hip width and chest depth are the best measures. Thus, hip width and/or rump length are the preferred measures. The results

(Table 9) suggest that rump length and hip width in particular are more difficult to measure than withers height, but the differences are small (3% and 2.2% respectively). Both measurements are influenced by management of young cattle. Analyses of within-herd results confirm the superiority of these measures compared with height, but indicate that hip width is more reliable than rump length.

A limited study of bulls showed a different pattern of results. In a very small sample of mature bulls, body length was significantly the best indicator of cumulative(3) index, followed by rump length and scrotal circumference, while chest depth was the best indicator of weight. In the full sample of bulls, rump length and chest depth were the best indicators of cumulative(3) index, while all linear measures had a high correlation with weight. Results for linear measurements of young bulls, together with weight and cumulative(3) index, were presented in histogram profile charts, and these were used



Figure 2. This heifer was reared on a low-input, non-intensive system. Linear assessment identified her type and quality at a young age.

in conjunction with GCI (effective founder number) data (Alderson 1992) to identify superior bulls. The results for each bull were also presented with the results of their sire and dam.

Limitations

Cumulative(3) index is the best measure of functional value of an animal, but it requires five linear measurements. It has value in detailed studies and analyses of breeds of cattle, but it is too complicated to operate on a routine herd basis. The measurements of hip width or rump length, ideally combined in a rump area index, offer simpler alternatives, and are superior to height as indicators of cumulative(3) index or weight.

Consistency of measurements was influenced to some degree by human error. There seemed to be greater difficulty with some measurements; but all were within a 5%

limit of variation. The employment of one operator was designed to minimise this problem.

Environmental influence also confused the genetic effect, especially in young animals. For example, sire referencing demonstrated that animals raised on Salisbury Plain varied significantly from their half-sibs on Oxfordshire pastures, and the effect was noted even on indices, particularly weight, length(2) index, balance and depth index (table 10) in these two herds. A wider study of five herds confirmed the environmental effect on weight, length(2) index and depth index, but not the effect on balance. All the linear measurements except withers height and rump height were susceptible to environmental effect.

Different calculations are necessary for different breeds, and this is indicative of varying conformation between breeds. These differences were not tested fully, but

Table 7. Indices of conformation in different types and breeds of cattle.

| Breed | Type | Index | |
|---------------|--------------|---------------|--------------|
| | | Length/height | Length/girth |
| White Park | Beef | 1.095 | 0.72 |
| British White | Beef | 1.089 | 0.72 |
| Gloucester | Dual-purpose | 1.056 | 0.71 |
| Ayrshire | Dairy | 1.053 | 0.68 |
| Friesian | Dairy | 1.039 | 0.68 |

Table 8. Indices of conformation in mature cows in two herds of White Park cattle

| Herd | Ht slope | Index | | | |
|----------|----------|--------|---------|-------|---------|
| | | Length | Rump lg | Depth | Balance |
| Dynevor | 2.07 | 1.99 | 0.58 | 0.55 | 0.9 |
| Chartley | -1.51 | 1.98 | 0.57 | 0.57 | 0.88 |

Table 9. Variation of measurements compared with previous measurement on same cow by same operator.

| Cow | Age (m) | % change | | | | | | |
|---------|---------|------------|---------|--------|---------|-----------|----------|----------|
| | | Withers ht | Rump ht | length | Rump lg | Hip width | Chest wi | Chest de |
| A | 144/180 | 1.21 | 1.21 | 2.17 | 3.81 | 3.69 | 2.38 | 2.82 |
| B | 132/156 | 1.55 | 0.76 | 1.36 | 0.91 | 6.03 | 10.42 | 2.67 |
| C | 111/144 | 2.86 | 1.57 | 3.89 | 3.51 | 0 | 2.68 | 5.63 |
| D | 108/132 | 1.53 | 1.52 | 1.02 | 1.96 | 5.26 | 7.45 | 2.61 |
| E | 96/132 | 0 | 2.99 | 1.36 | 9.43 | 0 | 9.78 | 4.11 |
| F | 91/118 | 1.45 | 1.76 | 4.05 | 1.82 | 6.67 | 2.04 | 1.32 |
| G | 86/122 | 1.18 | 1.61 | 4.12 | 9.57 | 3.71 | 2.33 | 0 |
| Average | | 1.397 | 1.631 | 2.567 | 4.43 | 3.623 | 5.297 | 2.737 |

Table 10. Progeny results for a bull in two herds under contrasting systems of management. Results expressed as % difference from breed average.

| Herd | Index | | | | | | |
|-------------------|----------------|--------------------|--------------------|---------------|---------|---------------|-----------------|
| | Weight (kg) | Length(1) index | Length(2) index | Rump index | Balance | Leg length | Cum(3) index |
| 1 (17 daughters) | -11.3 | 3.2 | -0.1 | -3.5 | 0.2 | 5.6 | 0.5 |
| 2 (6 daughters) | 9.8 | 1.3 | 1.6 | 1.4 | 4.2 | 3.9 | 4.2 |
| Sire's own rating | 12.2 | 2.1 | 9.1 | -10.8 | 1.4 | -0.1 | 4.6 |

preliminary results were obtained from Red Poll, Irish Moiled and Traditional Hereford cattle.

Linear assessment is only one factor in selection procedures. It takes no account of maternal qualities, and the use of linear assessment in the absence of breeding records or evaluation of temperament could be misleading.

Conclusions

Height at either withers or rump has limited value as an indicator of weight, and negligible value as an indicator of type and function. The value of weight is limited without some qualification of associated type and conformation. Cumulative(3) index is a useful indicator of overall morphological merit, as it combines values of weight and structure, and provides an accurate portrait of typical breed type. It is relatively constant throughout the life of an animal and can be used on young animals to predict mature merit. It is influenced by environmental effect (although significantly less so than weight), and it is complicated, requiring five linear measurements. Thus cumulative(3) index has potential application in breed studies to establish type and function, although its application by individual breeders is less attractive.

Single linear measurements are more relevant for on-farm within-herd use. Hip width or rump length, individually or

combined, are the preferred measurements. They have a high correlation with cumulative(3) index and weight, and are significantly superior to height in both cases. They are influenced by environmental effect, and are of limited value in between-herd comparisons.

The study demonstrated the value of linear assessment in general, and specific measurements in particular, with White Park females, and identified further potential areas of study to evaluate and refine the proposed system, namely:

1. repeat the study with a significant sample of bulls to assess the validity of indices for both sexes of White Park cattle
2. relate linear measurements to production characteristics other than weight (e.g. meat quality, KO%, maternal qualities, longevity)
3. study the effect of compensatory growth on linear assessment in animals reared under different systems of management
4. apply the system to other breeds to compare them with the White Park, and to test cumulative(3) index as an indicator of function in different breeds.

References

Alderson, G.L.H. 1992. A System to Maximise the Maintenance of Genetic Variability in Small Populations; In L.Alderson and I.Bodo (eds) Genetic Conservation of Domestic Livestock (Vol. II), CAB International, Wallingford

Alderson, L. 1997. A Breed of Distinction; Countrywide Livestock Ltd, Shrewsbury

Blott, S.C. 1997. Genetic variation within and between European cattle breeds; University of Edinburgh

Jewell, P.A. 1963. Cattle from British archaeological sites: In A.E.Mourant and F.E.Zeuner (eds) Man and Cattle, Royal Anthropological Institute, London

Royle, N.J. 1983. Polymorphisms of Rare Breeds of Cattle; University of Reading

Schwabe, A.E. & S.J.G. Hall. 1989. Dystocia in nine British breeds of cattle and its relationship to the dimensions of the dam and calf; *The Veterinary Record*, 125, 636-639

Simon, D.L. & D. Buchenauer. 1993. Genetic diversity of European livestock breeds; Wageningen Pers.

Stoll, J., J.F. Duckerts, F. de Graaf, L. Journeaux & E. Rehben. 1996. Report of the ICAR Working Group on conformation recording; In J.Renaud and J.van Gelder (eds) Performance recording of animals (EAAP Publication No. 87), Wageningen Pers.

A programme for the inventory, characterisation, evaluation, conservation and utilisation of European rabbit (*Oryctolagus cuniculus*) genetic resources

G. Bolet¹, M. Monnerot², C. Arnal³, J. Arnold⁴, D. Bell⁵, G. Bergoglio¹⁴, U. Besenfelder⁶, S. Bosze⁷, S. Boucher⁴, J.M. Brun¹, N. Chanteloup⁸, M.C. Ducourouble⁹, M. Durand-Tardif¹⁰, P.J. Esteves¹¹, N. Ferrand¹¹, G. Hewitt⁵, T. Joly¹², P.F. Koehl¹³, M. Laube⁹, S. Lechevestrier⁹, M. Lopez³, G. Masoero¹⁴, R. Piccinin¹⁵, G. Queney², G. Saleil¹, A. Surridge⁵, W. van der Loo¹⁶, J. Vanhomerig¹⁵, J.S. Vicente¹⁷, G. Virag¹⁸ & J.M. Zimmermann⁴

¹Institut National de la Recherche Agronomique, S.A.G.A., P.O. Box 27, F31326 **Castanet-Tolosan** Cedex, France

²Centre National de la Recherche Scientifique, C.G.M., Avenue de la Terrasse, F91198 **Gif sur Yvette** Cedex, France

³Universidad de Zaragoza, Facultad de Veterinaria, Miguel Servet, 177. 50013 **Zaragoza**, Spain

⁴Fédération Française de Cuniculture, 28 rue du Rocher. F75008 **Paris**, France

⁵University of East Anglia, School of Biological Sciences, NR4 7TJ. **Norwich**, Great Britain

⁶Interuniversitäre Forschungsinstitut Für Agrarbiotechnologie, Konrad Lorenz Strasse 20, A3430 **Tulln**, Austria

⁷Agricultural Biotechnology Center, Institute for Animal Science, PO Box 411, **Gödöllő** H2101, Hungary

⁸Institut National de la Recherche Agronomique, P.A.P., F37380 **Nouzilly**, France

⁹Centre d'Enseignement Zootechnique, Ecole d'Aviculture, Parc du Chateau, F78120 **Rambouillet**, France

¹⁰Bureau des Ressources Génétiques, 16 rue Claude Bernard, F75231 **Paris** Cedex 05, France

¹¹Faculdade de Ciências do Porto, Departamento de Zoologia-Antropologia, Praça Gomes Teixeira, P4050 **Porto**, Portugal

¹²Institut Supérieur d'Agriculture Rhône Alpes, 31 Place Bellecour, F69288 **Lyon** Cedex 02, France

¹³Institut Technique de l'Aviculture, 28 rue du Rocher. F75008 **Paris**, France

¹⁴Istituto Sperimentale per la Zootecnia, Via Pianezza, 115. 10151 **Torino**, Italy

¹⁵Entente Européenne d'Aviculture et de Cuniculture, Rabbit Section, Wilhelminastraat 8. 6351 GN **Bocholtz**, The Netherlands

Summary

In Europe, more than 60 breeds are described by the national associations of rabbit breeders. However, these breeds are scarcely used in the commercial production of rabbit meat in Europe, which is based mainly on commercial strains. A European programme, coordinated by the I.N.R.A., has been initiated to realise the inventory of all these breeds and to evaluate the zootechnical value and the genetic characteristics of some of them.

Through the European association of rabbit breeders and the FAO national focal points, all the European countries have been asked to fill out a questionnaire describing their populations of rabbits. A data bank is being set up, which will be included in the FAO (DAD-IS) and EAAP data banks. A sample of 10 breeds has been chosen (Flemish Giant, French Lop, Belgian Hare, Vienna White, Champagne Argente, Thuringer, Fauve de Bourgogne, Chinchilla, Himalayan, British). Their zootechnical value (reproduction, growth and carcass traits) is being evaluated



Figure 1. Flemish Giant rabbit.

on three experimental farms, in comparison with a control breed. At the same time, their genetic polymorphism and the genetic distances between these 10 breeds are calculated on the basis of microsatellites, mitochondrial DNA, other genetic markers and protein polymorphism. Finally, a bank of frozen embryos from these 10 breeds is being constituted.

Résumé

En Europe, plus de 60 races de lapins sont décrites dans les standards des Associations nationales d'éleveurs. Cependant, ces races sont très peu utilisées pour la production de viande, qui est basée essentiellement sur des animaux croisés provenant d'un faible nombre de races ou souches. Un programme européen, coordonné par l'INRA, a été mis en place pour recenser ces races et évaluer le potentiel zootechnique et les caractéristiques génétiques de certaines d'entre elles. Par la voix de l'Entente Européenne de Cuniculture et des points focaux nationaux de la F.A. O.,

tous les pays d'Europe ont reçu un questionnaire destiné à décrire leurs races de lapins. Une banque de données est en cours de réalisation, elle sera ultérieurement intégrée dans les banques de données de la F.A.O. (DAD-IS) et de la F.E.Z. Un échantillon de 10 races a été choisi (Géant des Flandres, Bélier Français, Lièvre Belge, Blanc de Vienne, Argenté de Champagne, Chamois de Thuringe, Fauve de Bourgogne, Chinchilla, Russe, Papillon Anglais). Leurs performances zootechniques (reproduction, croissance et caractéristiques de la carcasse) sont évaluées dans 3 fermes expérimentales, en comparaison avec une souche témoin unique. A partir du polymorphisme de microsatellites, de l'ADN mitochondrial, de marqueurs génétiques et de protéines, la variabilité génétique au sein de ces races et les distances génétiques entre elles sont calculées. Enfin, une banque d'embryons congelés est en cours de réalisation.

Key words: Conservation, Rabbit data bank, Breed characteristics, Reproductive traits, Genetic diversity.

Introduction

Western Europe contributes some 60 per cent of the world's commercial production of rabbit meat. This production is sometimes based on pure-bred animals, but mainly on cross-bred animals derived from very few specialised strains disseminated through pyramidal systems. Pure-breeding is decreasing and local populations, used in traditional backyard farming, are currently nearly extinct. Additionally, these populations have hardly been surveyed (Arnold and Rochambeau, 1983). Depending on the extent of selection, one can distinguish local populations or breeds, defined by standard (more than 60 in Europe) or commercial strains, extracted from a few breeds and strongly selected for production traits. Besides rabbit breeders for meat production, there are fancy breeders. The former are predominant in France, Spain, Italy, Hungary,... where rabbit meat consumption is popular. The latter are particularly active in Germany and Switzerland, but also in France, Italy, Belgium

and the United Kingdom; most of their national associations are federated into a European Association (Entente Européenne d'Aviculture et de Cuniculture), including 15 countries and 66 breeds, most of them being present in various countries.

At the present time, profits obtained from rabbit husbandry are insufficient to encourage the development of this industry. An inventory of existing genetic resources could allow a better knowledge of the abilities of breeds rarely used today because of their low level of production under standard conditions. It could lead to the development of new activities with higher profits for breeders. Owing to the great variability in body size, diversification of carcass weight is one approach. The patterns of production could be diversified, and, with more adapted breeds, lead to a more extensive way of rearing rabbits for a part of the production. This inventory could also lead to the utilisation of breeds showing a good resistance to some diseases, which would reduce the use of chemicals to the benefit of the breeder and the consumer. It could also lead to the utilisation of animals with a better



Figure 2. French Lop rabbit.



Figure 3. Belgian Hare rabbit.

feed efficiency which would help decrease effluents produced by animal husbandry (particularly the nitrogenous waste), and hence help protect the environment. Finally, a more thorough knowledge of the different breeds, particularly from an immunological point of view, would develop the production of rabbits for laboratories, producing more homogeneous or more sensitive groups of animals better suited to users' needs.

Characterisation and Conservation: Current Status

Some efforts have already been made to characterise European rabbit breeds.

- Most national associations of fancy rabbit breeders have made a primary phenotypic characterisation of the breeds they manage (national breed standards). The European Association, mentioned above, has published a European standard of breeds (Anonymous, 1995), which tries to synthesise the national standards. It gives

a detailed description of the shape, conformation, size, fur coloration for a total of 66 breeds.

- A European network of interested laboratories has been working together on the characterisation of genetic polymorphism in several wild and domestic populations from Portugal, Spain and France (Monnerot *et al.*, 1994; Van der Loo *et al.*, 1991, 1999; Ferrand, 1995).
- A research network on rabbit production in the Mediterranean area was founded in 1987 with the support of the International Centre for Advanced Mediterranean Agronomic Studies (CIHEAM). It includes 8 countries (including Spain, Italy and France) and is working on the identification and characterisation of local populations and breeds (Rouvier, 1994; Khalil, 1998). So far, mainly southern Mediterranean populations have been considered.
- A European concerted action ("Germplasm banking"), in which the rabbit was used as a model animal, allowed the defining of the technical basis for cryoconservation of rabbit embryos. Embryos from some commercial cross-bred strains and from 3 small-size endangered strains were collected and frozen (Joly *et al.*, 1994, 1996).

In spite of these efforts, important points are lacking concerning European breeds :

- The rabbit is missing from the FAO / DAD-IS, <<http://www.fao.org/dad-is/>> and the European Association for Animal Production (EAAP) (Hannover) data banks on domestic animal breeds.
- Most previous work was devoted to selected strains or cross-breeding (see Rochambeau, 1988, for a review), and comparative studies of native breeds are badly lacking. Only some zootechnical evaluations so far have concerned a few breeds treated separately (Pilandon *et al.*, 1986; Chevalier *et al.*, 1986; Lopez *et al.*, 1992, Pagano Toscano *et al.*, 1992, Koehl and Van Der Horst, 1998), and some

cross-bred products of these breeds were compared (Ouhayoun and Poujardieu, 1978).

- There is no systematic collection of rabbit genetic resources, as exists in most domestic animal species. Their preservation is currently performed *in situ* by either professional or fancy breeders, but without any consistent programme such as that of the "Conservatoire National des Animaux de Basse-Cour" which worked in France between 1978 and 1985 (Arnold and Rochambeau, 1983) and no longer exists.

So far, different parts of a conservation programme have developed more or less independently, and do not constitute an integrated and coherent project. For these reasons, an ambitious programme was proposed and approved within an EC framework (Regulation No 1467/94 on the conservation, characterisation, collection and utilisation of genetic resources in agriculture). The objective of this paper is to describe in detail this programme, which began in 1997.

Creation of a Data Bank

The first phase involves the definition of a rabbit data bank. The rabbit has not, to date, been included in the FAO and EAAP data banks. Our project will allow a data bank to be created with information from the European countries involved in the project. The first source of data will be the review of 66 breeds registered by the above mentioned European Association of Rabbit Breeders which proposes only a phenotypic description of the breeds and very little zootechnical data. Detailed propositions for the organisation of a rabbit data bank have already been made by Lukefahr (1988) and by Khalil (1993). On the basis of these different sources, we assembled a questionnaire (Annex 1) which was sent to national rabbit breeders associations and to FAO European national focal points. This questionnaire is available in English, French, Spanish, Italian, German and Russian. So far, more than 100 national breeds were registered, from twelve countries. The questionnaire may be obtained from the first

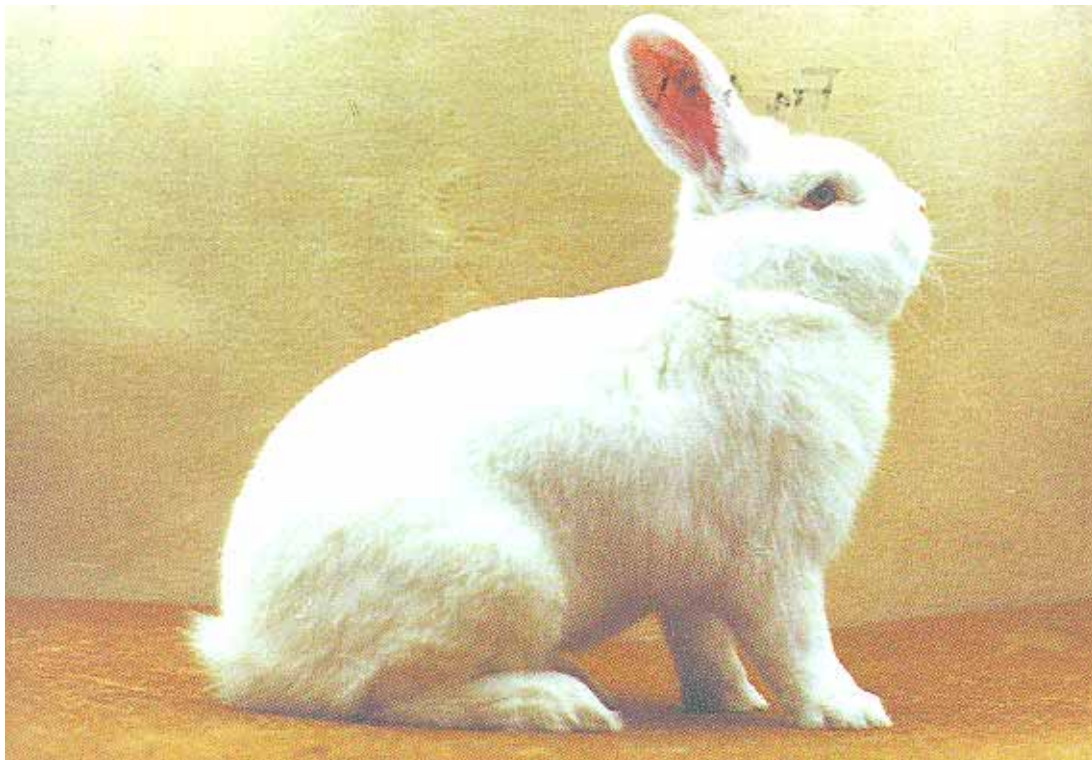


Figure 4. Vienna White rabbit.

author of this paper. The construction of the computerised data bank is now in progress and a first draft will be available in 1999 (Ducourouble *et al.*, 1999). It will be maintained by participants and incorporated simultaneously, in FAO and EAAP data banks, as soon as possible.

Detailed Characterisation of a Sample of 10 Breeds

Choice of the 10 breeds

As it is not possible to make a detailed characterisation of all the censed breeds, for economical reasons, we decided to work on a sample of 10 breeds. Figures from 1 to 10 show these breeds.

These breeds were chosen on three criteria:

- They are presumed to be among the oldest; some breeds which are known to originate from recent cross-breeding of the oldest ones.
- They are present in various European countries; some breeds, which are present in only one country and whose

preservation is relevant of a national programme, were discarded in spite of their potential interest.

- They have a potential zootechnical interest. The 10 chosen breeds were :
- Two heavy breeds : Flemish Giant and French Lop
- Five average-sized breeds : Belgian Hare, Vienna White, Champagne Argente, Thuringer, Fauve de Bourgogne
- Three small-sized breeds : Chinchilla, Himalayan, British (Papillon).

Zootechnical characterisation

The evaluation of the breeds takes into account the following points :

- On-farm characterisation concerning reproductive performances and genealogies.
- Zootechnical evaluation, in controlled management and environmental conditions including growth, feed efficiency, carcass traits and muscular fibre characterisation.

For the latter, the average size of a sample from a population is expected to be around 50 females and 20 males whose dam and sire are known, as unrelated as possible. Young are bought in farms at weaning or at birth. When bought at birth, they have to be nursed



Figure 5. Champagne Argente rabbit.



Figure 6. Thuringer rabbit.

by a synchronised female on the experimental farm. This technique is now well known and used for introduction of rabbits in farms.

On each experimental farm, comparison is made between a control strain and one or more breeds at a time, according to the number of available cages. The control strain will be the 9077, from INRA (France), which has been kept without selection, as a control line, for 20 years. Conditions will be standardised as much as possible, but in agreement with biological abilities of each breed.

The parameters which will be measured are the following.

Reproductive traits

Reproductive performances of females will be studied during a period of 6 months to one year, i.e. an average of 4 litters, according to the reproduction rhythm chosen (in rabbits, mating is performed either the day after littering, 5 to 12 days after, or even after weaning in traditional systems). Ovulation of

does is induced by coitus (or by an injection of exogenous gonadotrophins in artificial insemination) (Bolet *et al.*, 1992).

In all cases, fertility rate, litter size and weight of young at birth will be recorded. In some cases, components of litter size, measured by laparoscopy, namely number of ova shed and number of embryos alive, will also be recorded. Both are easily counted by laparoscopy at 12 days of gestation without any damage to the female or the foetuses (Santacreu *et al.*, 1990). Male semen composition and quality will be recorded by semen collection using an artificial vagina, a technique routinely used in rabbits.

Growth and feed efficiency

Adult body weight differs dramatically between rabbit breeds, but little is known about growth rates, except in "standard" strains. First and second litters of does compared for reproductive traits will be used to establish growth curves: a sample of them will be weighed from birth to adult weight.



Figure 7. Fauve de Bourgogne rabbit.

To measure appetite and feed efficiency, animals will be fed ad libitum during growth, and feed consumption of groups of siblings after weaning will be measured.

Carcass traits

To study the carcass traits, a sample of young from 3rd and 4th littering will be slaughtered around 2, 2.3 or 2.6 kg. These weights correspond to classical slaughter weights and to potentially heavier weights more adapted to cutting. The carcass composition will be studied according to classical criteria defined and normalised by Blasco *et al.* (1993), including a reference carcass dissection and prediction of carcass composition (via meat/bone ratio estimated by hind leg dissection). Components of quality such as muscular pH and colour will be measured. When possible, organoleptic qualities of meat will be analysed by trained panels.

The above assessment of quality will be completed by further analysis of muscular fibres using established techniques (the

avidin-biotin complex method and the ATP-ase method). On samples from the same animals, lipids, energy, protein and magnesium content of muscle will be determined using a near infrared spectroscopy (NIRS) method on freeze-dried samples of muscle (Masoero *et al.*, 1994).

Detailed Characterisation of Genetic Diversity

A number of different markers will be used to evaluate different breeds and different geographical populations within each breed, to establish diversity within and between breeds. All of them have proved to be useful criteria for this purpose in preliminary works. The final objective is to measure the degree of specificity of each breed. This will also allow definition of the best possible sampling strategy for measuring and conserving biodiversity at all levels.

Diversity in maternal lineage: mitochondrial DNA characterisation

Mitochondrial DNA (mtDNA), which is almost exclusively maternally inherited, is an essential tool to assign animals to a precise maternal lineage. Early studies on several natural populations and three breeds has led to the recognition of 22 types organised within two well differentiated and geographically structured maternal lineages (Monnerot *et al.*, 1994). Sequence analysis of various domains of mtDNA belonging to these different types has allowed the genetic characterisation of animals whatever their degree of relationship. Two regions of mtDNA are assessed: part of cytochrome b gene for distantly related animals and part of the main non-coding domain for closely related animals. Various pairs of primers specific for these two regions define mtDNA portions of different lengths and phylogenetic signals. Once amplified by PCR, these portions are analysed either by sequencing or by RFLP (Hardy *et al.*, 1995; Monnerot *et al.*,

1996; Mougel, 1997). Comparisons with data previously collected will then allow the animal under study to be assigned to one of the 2 mtDNA groups already described.

Allelic diversity of immunoglobulin loci

The antibody diversity in the rabbit is fundamentally different from that of other species. The rabbit Ig loci are highly polymorphic (with alleles that show up to 40 % amino acid differences). Most polymorphic are the a locus encoding the predominantly expressed variable region gene and the b locus encoding the constant region of the Kappa light chain. There is strong evidence that in the rabbit species, allele diversity at the Ig loci is a crucial component of population fitness (Van der Loo and Verdoodt 1992, Van der Loo 1993). While contributing to the characterisation of populations, the monitoring of the Ig genotypes is important to avoid accidental loss of allele diversity in races.



Figure 8. Chinchilla rabbit.



Figure 9. Himalayan rabbit.

Haplotypic diversity of the Major Histocompatibility Complex (MHC)

In all species, MHC is the best 'identity card' of individuals, possibly playing a role in individual recognition. Rabbit class I and class II MHC gene haplotypes were described (Marche *et al.*, 1989). One class I and 4 class II loci (DQ, DR, DP, DZ/DO) are identified and used in this programme.

Casein polymorphism

In milk the four major caseins, namely β -, α s1-, α s2- and κ -casein exist in the form of casein micelles. Studies on the physicochemical characteristics of bovine, goat and ovine milk have shown that the genotype has a significant effect on casein, fat content and micelle characteristics. Whereas the bovine, ovine and caprine are of major importance to the dairy industry, rabbits are principally bred as meat animals. Nevertheless, the level of lactation in rabbit does is important since doe milk is the only

food available to pups up to the age of 21 days. RFLP of the rabbit κ -casein gene has been observed in DNA isolated from New Zealand rabbits and digested with Hind III (Baranyi *et al.*, 1996). It was shown that in this breed the κ -casein B allele is less frequent than the κ -casein A allele and that differences between the two alleles include part of the LINE sequences in the first and the fourth introns (Hiripi *et al.*, 1998). The polymorphism of κ -casein gene will be evaluated after DNA amplification of the first intron, using primers located in the bordering exons. The diversity of α s2 casein will be examined with isoelectric focusing of milk samples (Virág *et al.*, 1996).

Diversity of various proteins

Genetic variability between populations will be estimated from the analysis of protein polymorphism. Around 25 variable loci will be studied. The following established techniques will be used: conventional electrophoresis, electrophoresis in denaturing conditions, isoelectric focusing in pH gradients, isoelectric focusing in immobilised pH gradients, isoelectric focusing followed by immunoblotting or enzyme blotting. Enzymes will be visualised with specific staining methods and non enzymatic proteins with current staining methods. (Ferrand, 1995).

Diversity of alleles at microsatellite loci

Microsatellites occur commonly in eukaryotic genomes and consist of stretches of mono, di or tri nucleotide motifs which are randomly repeated, dispersed throughout the whole genome and usually flanked by unique sequences. They are becoming the markers of choice for molecular population genetics because they represent an important source of polymorphic genetic markers which are easily and rapidly scored. Differentiation of very closely related populations is possible based

on allele frequencies as well as diagnostic alleles. A set of 20 to 30 microsatellite loci already identified and available will be used (Rico *et al.*, 1994; Mougel *et al.*, 1997; Surridge *et al.*, 1997; Van Haeringen *et al.*, 1997). Some of them have already been shown to be useful in characterising wild and domesticated populations (Vachot, 1996; Mougel, 1997; Surridge, 1997). They will provide an essential tool for the identification of breeds and testing individual genetic relatedness in combination with the other markers presented above. In the future all the nuclear markers could be useful in identifying quantitative trait loci (QTL) for breeding for zootechnical characteristics.

Conservation

The conservation actions will be as follow.

Constitution of a bank of frozen embryos

It is our intention that embryos and possibly semen of each breed should be stored in at least 2 places for a matter of security, according to recommendations concerning management of frozen germplasm. When sampling and collecting this material, the objectives are to avoid any shift in gene frequencies between original population and cryoconserved sample population. To reduce the initial genetic drift, three variables must be taken into account: sample size (Joly *et al.*, 1999), choice of animals and breeding system. The efficiency of the method has already been investigated (Joly *et al.*, 1996).

In situ preservation by breeding the selected populations in farms



Figure 10. British rabbit.

Following the data produced by this project, *in situ* preservation of breeds of reduced genetic variation or at particular risk of extinction will be proposed in private farms. These will be supported technically by breeders organisations and supported financially, in order to manage their stock according to some defined rules (mating plan, number of bucks and does, exchange between farms,...).

Conclusion

The situation of European rabbit breeds among domestic mammals is unique: there is still a great diversity of breeds, but they are mainly owned by fancy breeders who have more interest in their phenotype than their zootechnical and economical value. These genetic resources may be important for rabbit meat production if rabbit breeders can take advantage of this diversity. The inventory of all the breeds and the detailed characterisation of a sample of 10 breeds will provide valuable information on economical value of this diversity. This programme will collect and disseminate such vital information, and may have important consequences for the management and conservation of European rabbit genetic resources for the future.

References

- Anonymous.** 1995. Standard 95 pour l'appréciation des races de lapins reconnues par l'Entente Européenne. Pfister U. and Piccinin R. ed.
- Arnold J. & Rochambeau H. de.** 1983. Inventaire des races menacées et des actions de conservation : les animaux de basse-cour. Ethnozootechnie n° 33.
- Baranyi M., Aszodi A., Devinoy E., Fontaine M-L., Houdebine M-L. & Bösze S.** 1996. Structure of the rabbit κ -casein encoding gene: expression of the cloned gene in the mammary gland of transgenic mice. *Gene* 174. 27-34.
- Blasco A., Ouhayoun J. & Masoero G.** 1993. Harmonisation of criteria and terminology in rabbit meat research. *World Rabbit Science*, 3, 3-10.
- Bolet G., Garcia-Ximenez F. & Vicente J.S.** 1992. Criteria and methodology used to characterise reproductive abilities of pure and crossbred rabbits in comparative studies. *Options Méditerranéennes*. n° 17, 95-104
- Chevalier C., Perrier G., Ouhayoun J., de la Foye F., Matter P. & Charrier E.** 1986. Potentialités zootechniques de la race Argente de Champagne. 4èmes Journées de la recherche cunicole en France, Paris, 10-11 décembre 1986, vol 2., paper 27. INRA-ITAVI
- Ducourouble M.C., Lechevestrier S., Arnold J., Bolet G., Boucher S., Brun J.M., Durand-Tardif M., Koehl P.F., Menigoz J.J., Piccinin R., Saleil G.** 1999. Réalisation d'une base de données informatique sur les races de lapins en Europe. 8èmes Journées de la Recherche Cunicole, Paris, 9-10 juin 1999, INRA-ITAVI
- Ferrand N.** 1995. Variação genética de proteínas em populações de coelho (*Oryctolagus cuniculus*). Unpublished doctoral thesis, Porto University.
- Hardy C., Callou C., Vigne J.D., Casane D., Dennebouy N., Mounolou J.C. & Monnerot M.** 1995. Rabbit mitochondrial DNA diversity from prehistoric to modern times. *J. Mol. Evol.*, 40, 227-237.
- Hiripi L., Devinoy E., Rat P., Baranyi M., Fontaine M.L., Bösze Z.** 1998. Polymorphic insertions/deletions of both 1550 nt and 100 nt in two microsatellite-containing, LINE-related intronic regions of the rabbit κ -casein gene. *Gene* 213. 23-30.

Joly T., Theau-Clement M., Drouet-Viard F., de Rochambeau H. & Renard J.P. 1994. Application de la cryoconservation des embryons à la protection des ressources génétiques chez le lapin. *Genet. Sel. Evol.*, 26, supp. 1, 267-278.

Joly T., Vicente J., Theau-Clement M., Garcia-Ximenez F., Besenfelder U. & Renard J.P. 1996. Cryopreservation of genetic resources in rabbit species: practical applications. 6th World Rabbit Congress, 9-12 July 1996, Toulouse, France.

Joly T., Renard J.P., Rochambeau H. de & Baselga M. 1999. Size of the sample to store for re-establishing a breed from frozen embryos in domestic rabbit. *Livest. Prod. Sci.* (submitted).

Khalil M.H. 1993. Descriptive model for rabbit genetic resources data bank. *World Rabbit Science*, 1, 113-118.

Khalil M.H. 1998. Model for the description of rabbit genetic resources in Mediterranean countries. Application to the Egyptian breeds Giza White and Baladi. CIHEAM ed.

Koehl P.F. & Van Der Horst F. 1998. Performances zootechniques d'un troupeau de lapins de race Normande. 7èmes Journées de la recherche cunicole en France, Lyon, 13-14 May 1998, 41-44, ITAVI, Paris.

Lopez M., Sierra I. & Lite M.J. 1992. Carcass quality in Gigante de España purebred and commercial crossbred rabbits. *Options Méditerranéennes, Série A: Séminaires Méditerranéens*, 17, 75-80.

Lukefahr S.D. 1988. Conservation of global rabbit germplasm resources. 4th Congress of the World Rabbit Science Association, 10-14 October 1988, Budapest (Hungary). Vol "Genetics and Physiology", pp. 129-136 (S. Holdas ed.).

Marche P.N., Rebiere M.C., Laverriere A., English D.W., Leguern C. & Kindt T.J. 1989. Definition of rabbit class I and class II MHC gene haplotypes using molecular typing procedures. *Immunogenetics*, 29, 273-276.

Masoero G., Dalle Zotte A., Parigi-Bini R., Xiccato G. & Bergoglio G. 1994. Utilisation de la spectroscopie dans le proche infrarouge (NIRS) dans l'évaluation de la qualité de la carcasse et de la viande de lapins soumis au transport pre-abattage. 6èmes Journées de la recherche cunicole en France, La Rochelle, 16-17 December 1994, vol. 2., 435-439, ITAVI, Paris

Monnerot M., Vigne J.D., Biju-Duval C., Casane D., Callou C., Hardy C., Mougel F., Soriguer R., Dennebouy N. & Mounolou J.C. 1994. Rabbit and man: genetic and historic approach. *Genet. Sel. Evol.*, 26, supp.1, 167-182.

Monnerot M., Loreille O., Mougel F., Vachot A.M., Dennebouy N., Callou C., Vigne J.D. & Mounolou J.C. 1996. The European rabbit: wild population evolution and domestication. 6th Congress of the WRSA, 9-12 July 1996, vol 2, 331-334.

Mougel F., Mounolou J.C. & Monnerot M. 1997. Nine polymorphic microsatellite loci in the rabbit, *Oryctolagus cuniculus*. *Animal Genetics*, 28, 58-71.

Mougel F. 1997. Variations de trois types de marqueurs génétiques dans l'évolution de l'espèce *Oryctolagus cuniculus*: aspects moléculaires et relations avec la biologie et la structure des populations. PhD Thesis, University of Paris-Sud.

Ouhayoun J. & Poujardieu B. 1986. Etude comparative de races de lapins en croisement. Relations interraciales et intraraciales entre caractères des produits terminaux. 4èmes Journées de la recherche cunicole en France, Paris, 10-11 décembre 1986, vol 2., paper 25. INRA-ITAVI

- Pagano-Toscano G., Lazzaroni C., Zoccarato I. & Benatti G.** 1992. Conservation and improvement of the Carmagnola Grey Rabbit. 5th Congress of the WRSA, 25-30 July 1992, Corvallis (USA). *J. Appl. Rabbit. Res.*, 15, 240-246.
- Pilandon M.T., Henaff R. & Ponsot J.F.** 1986. Potentialités zootechniques d'une souche de lapins Gris du Bourbonnais. Bilan de 5 années d'observations. 4^{èmes} Journées de la recherche cunicole en France, Paris, 10-11 December 1986, vol. 2., paper 26., INRA-ITAVI
- Rico, C., Rico, I., Webb, N., Smith, S., Bell, D. & Hewitt, G.** 1994. Four polymorphic microsatellite loci for the European wild rabbit, *Oryctolagus cuniculus*. *Animal Genetics* 25, 367.
- Rochambeau H. de.** 1988. Genetics of the rabbit for wool and meat production (1984-1987). 4th Congress of the World Rabbit Science Association, 10-14 October 1988, Budapest (Hungary). Vol. "Genetics and Physiology", pp. 1-68 (S. Holdas ed.).
- Rouvier R.** 1994. Les travaux du groupe "Réseau de recherches sur la production de lapin dans les conditions méditerranéennes". First international conference on rabbit production in hot climates, 6-8 September 1994, Cairo (Egypt). *Cahiers Options Méditerranéennes*, 8, 27-32.
- Santacreu, M.A., Viudes, M.P. & Blasco, A.** 1990. Evaluation par coelioscopie des corps jaunes et des embryons. Influence sur la taille de portée chez la lapine. *Reprod. Nutr. Dev.*, 30, 583.
- SurrIDGE, A. K.** 1997. Genetic variation and population structure of European wild rabbits (*Oryctolagus cuniculus*). PhD Thesis, University of East Anglia.
- SurrIDGE, A. K., Bell, D. J., Rico, C. & Hewitt, G. M.** 1997. Polymorphic microsatellite loci in the European rabbit (*Oryctolagus cuniculus*) are also amplified in other lagomorph species. *Animal Genetics* 28, 302-305.
- Vachot A.M.** 1997. Homologies et singularités interspécifiques (Ranides) et intraspécifiques (Lapins). PhD Thesis, University of Paris-Sud.
- Van Der Loo W., Ferrand N. & Soriguer R.C.** 1991. Estimation of gene diversity at the b-locus of the constant region of the immunoglobulin light chain in natural populations of European Rabbit (*Oryctolagus cuniculus*) in Portugal, Andalusia and on the Azorean Islands. *Genetics*, 127, 789-799.
- Van der Loo W. & Verdoodt B.** 1992. Patterns of interallelic divergence at the Rabbit b-locus of the Immunoglobulin Light chain constant region (c_{κ1}) are in agreement with population genetic evidence for overdominant selection. *Genetics* 132, pp. 1105-1117.
- Van der Loo W.** 1993. Variance analysis of immunoglobulin alleles in natural populations of rabbit (*Oryctolagus cuniculus* L.): the extensive interallelic divergence at the b-locus could be the outcome of overdominant-type selection. *Genetics* 134: 171-187.
- Van der Loo, W. F. Mougél, Ch. Bouton, M. S. Sánchez, & M. Monnerot.** 1999. The allotypic patchwork pattern of the rabbit *IGKC1* allele b5wf: genetic exchange or common ancestry? *Immunogenetics*, 49, 7-14.
- Van Haeringen, W. A., Den Biemen, M., Van Zutphen, L. F. M. & Van Lith, H. A.** 1997. Polymorphic microsatellite DNA markers in the rabbit (*Oryctolagus cuniculus*). *J. Experimental Animal Science* 38, 49-57.
- Virág Gy., Baranyi M., Bősze Zs. & Devinoy E.** 1996. Variability of α s2 casein phenotypes in a New Zealand White rabbit breeding stock in Hungary. *World Rabbit Science* 4, 84.

The Peel-Neck chicken of Belize and Guatemala, Central America

J.G. Mallia

Department of Population Medicine, Ontario Veterinary College, University of Guelph,
Guelph, Ontario, Canada N1G 2W1

Summary

A previously undescribed heavy, soft-feathered population of chickens characterised by a partially featherless portion of the neck known in Belize as the "Peel-Neck" was studied. It was also widely present in Guatemala. This population of chickens was said to have a dual purpose, raised for meat and eggs. The eggs were tinted, and the hens were sitters, also caring for the young until independent. The birds were active and independent foragers. The plumage colour was variable, but body weight and conformation were consistent across all specimens, as was the comb type and dimensions, and location of the featherless area on the neck. These chickens were kept under backyard rearing systems in both countries, although usually they were allowed to scavenge, at which they were said to excel. This population was not considered threatened, although apparently no attempt was made to breed pure specimens.

Key words: Genetic resources, Chicken, Belize, Guatemala.

Résumé

Nous décrivons une population nouvelle de poule Bélizienne de grande taille au plumage doux, mais chez laquelle le cou est dépourvu de plumes, soit au "cou pelé". Cette population se retrouve aussi au Guatemala. C'est une race à viande et à production d'oeufs teintés brun clair. Les poules sont

couveuses, protégeant leurs poussins jusqu'à leur indépendance. Elles sont aussi d'excellentes fourrageuses. Leur plumage est de couleur variable, mais leur masse, leur conformation, la forme de leur crête et l'emplacement de la partie du cou dépourvu de plumes sont semblables chez tous les spécimens. Ces poules sont élevées dans un système au sol. La population n'est pas menacée; par contre il n'y a aucun effort pour assurer l'élevage de race pure.

Introduction

The naked neck (Na) is an autosomal dominant gene responsible for the complete absence of feathers and down in the neck region of chickens (Davenport, 1914; Warren, 1933; Hertwig, 1933), mapped on chromosome 1 (Stevens, 1991). Several breeds exhibiting the naked neck trait have been described, including the "Cou Nu du Forez" from France, the "Malay Game" from Malaysia, the "Shingangadi" from Zaire, and the "Transylvanian Naked Neck" from Hungary and Romania (Merat, 1986). The "Transylvanian Naked Neck" is said to have been brought over into Europe during the Turkish occupation (Bodo *et al.*, 1990). Breed characteristics and production data are available for the European populations of chickens with the naked neck trait, for example France (Zein-el-Dein *et al.*, 1984; Pesti *et al.*, 1996), and Hungary (Bodo *et al.*, 1990). The naked neck trait was also said to be present in several regions of the world, generally those with a hot and humid climate (Merat, 1986), although breed descriptions

were not available. The purpose of this report is to bring attention to, and provide a description of a population of chickens in Belize and Guatemala exhibiting the naked neck trait.

Materials and Methods

Participants included local inhabitants raising and selling fowl in Belize and Guatemala in January 1994, and January 1998, identified by visiting the towns on their market day, and by the presence of free-ranging fowl, and chickens kept in enclosures adjacent to homes. A questionnaire was administered to the participants in English (Belize) or Spanish (Guatemala). Information on population data, description of the breed, its uses, management conditions and performance were collected. A version in English of the questionnaire is reported in table 1. Photographs were also taken to show body conformation, face and neck characteristics, and plumage.

Breed Description and Uses

The Peel Neck chicken was present throughout Belize and most of Guatemala, usually in mixed flocks also having chickens with normally feathered necks. Information on the Peel Neck was collected on 38 males and 114 females examined by the author, and additional information was gathered by means of a breed-information questionnaire based on that used by Mallia (1998). The body was large and compact with a "heavy breed" conformation, superficially similar to the Rhode Island Red or Sussex. The back was broad and flat, and the breast was prominent, broad and square (prevalence, $P=100\%$). On average, males weighed 3.95 kg (standard error, 0.10 kg), and females weighed 3.1 kg (standard error, 0.06 kg). The legs of the Peel Neck were proportionately longer than the fore-mentioned heavy breeds, devoid of feathering, and with prominent well spread toes ($P=100\%$). The wings were of medium size, carried high, horizontally, and close to the body, and in the male the wing tips were

Table 1. Peel Neck Fowl: questionnaire for growers and breeders in Belize and Guatemala (adapted from Mallia, 1998).

-
- Q1. How many birds, males, females, young, do you have, and where they kept ?
 - Q2. How many females are ideally kept for each male ?
 - Q3. Do they forage, do you feed them, or both ?
 - Q4. From where do you obtain new stock ?
 - Q5. Do you keep the chickens with ducks or turkeys ?
 - Q6. If Q5.is in the affirmative, do you perceive any problems ?
 - Q7. At what age do the males and females mature ?
 - Q8. What weight do the males and females have at maturity ?
 - Q9. What colour are the eggs, does the hen incubate the eggs, how many does she lay annually ?
 - Q10. What is the morbidity and mortality of the young, and if so, mainly at what age ?
 - Q11. What is the morbidity and mortality of the adults?
 - Q12 What are the main uses of rearing chickens ?
 - Q13. Is there a particular feather colour preference for Peel Necks ?
 - Q14. Do you breed Peel Necks with other fowl ?
 - Q15.What are the main uses of rearing Peel Neck chickens ?
 - Q16. Any management or production advantages of the Peel Necks with respect to other fowl ?
-



Figure 1. Male (to the left) and female (to the right) Peel Neck; (Belize).

usually obscured by the abundant shackle (P=100%). The tail was small or medium sized, carried upright at an angle of around 45° (P=73.4 %). The sickles in the male were only occasionally well-developed; usually they extended to only slightly beyond the rest of the tail, and were not markedly curved (P=73.4 %). Some males, particularly those in Guatemala, however, had an upright, well-developed tail with prominent sickles (P=26.6%). The head was of medium size with prominent, alert amber eyes and a medium-sized curved beak. There was no presence of white skin in the face or ear lobes (P=100%). The rest of the head was also devoid of feathering, aside from a small, well-defined tuft at the ears, and a “cap” of feathers extending around the blade of the comb (P=100%). The unfeathered area extended to about half way down the neck; an area with sparse feathering was present, corresponding to the anterior extremity of the keel; however, the rest of the body was covered with normal feathering (P =100%).

The plumage-type was soft-feathered in all specimens observed; feather colouring was very diverse: white, black, brown, red, partridge, speckled, silver cuckoo and wheaten colour phases were observed. The skin on the featherless areas was bright red, and usually with a fairly irregular texture (P=83%). Folds of skin were also occasionally present, particularly at the base of the posterior aspect of the wattles, giving a “double-wattled” appearance (P=78%). Despite the irregular appearance of the skin of the featherless areas, no evidence of follicles was present (P=100%). The comb in all specimens was single, upright, well-developed, deeply serrated and with 5-6 points in both the male and female; the free part of the blade was usually well-distanced from the contour of the skull (P =100%). Many of the physical characteristics of the Peel Neck chicken are illustrated in figures 1-3. The birds were very gregarious, and interacted well with other fowl and also humans. They actively foraged

around houses, gardens, ditches and fields during most of the daylight hours, roaming considerable distances. The hens laid tinted or light brown eggs ($P = 100\%$), and were easily trained to deposit their eggs in a convenient location for collection. They were also said to be excellent sitters, and several hens with broods were observed the hens being cautious and defensive when with young. The young were also very alert and independent, exploring various sources of food, for example ground coconut left out to dry. The long-legged appearance, bare head and neck areas of the young mirror those of the adults. The dorsal parts of the body were unevenly feathered or naked, from a young age, however the ventral parts, especially the abdomen, were thickly covered with feathers.

The questionnaire showed that despite their markedly wide distribution, there was no indication given by poultry growers of the Peel Neck having superior carcass or

egg-laying traits. Although they were considered to be good foragers, and the hens good sitters and mothers, they were not considered better than the fully-feathered varieties. Hens commenced laying eggs between seven and eight months, and laid between 40-50 eggs per year; these values are comparable to those found by Cook *et al.* (1997) for other backyard poultry raised in the Yucatan, Mexico. When reared in enclosed backyard compounds, Peel Necks were said to be about as productive as chickens with feathered necks: production of eggs was said to be linked to ration quality and quantity, rather than presence or absence of neck feathers, despite the predominantly hot and humid climate. The prevalence of Peel Neck chickens per backyard within a village varied substantially (e.g. from 0-80% in Monkey River Town, Belize). When present in a village, the overall prevalence of Peel Neck chickens was usually of 30-50%, although



Figure 2. Female Peel Neck with young; (Belize).



Figure 3. Female Peel Neck scavenging in village; (Guatemala).

these estimates are only approximate due to the difficulty in identifying all chickens within a population.

Discussion and Conclusion

It has long been hypothesised that chickens with the naked neck trait have better production traits compared to normal feathered chicks when raised in hot climates

(> 30°C); an excellent review is given by Merat (1986), and studies examining the possible production advantages of naked neck trait chickens are still ongoing (Pesti *et al.*, 1996). This study has not specifically attempted to assess the productivity of the Peel Neck chicken. However, it is interesting to note that although no advantage of raising Peel Necks was identified by those who raise them, these chickens are widely kept in both nations, and possibly also other Central American countries.

Some authors have suggested that the Na gene is incompletely dominant, and homozygous chickens (Na Na) can be distinguished from the heterozygous chickens (Na na) because the former genotype has markedly reduced or no feathers on the front of the neck (Crawford, 1976; Scott and Crawford, 1977), and lack of feathers on the ventral face of the thighs and breast (Merat, 1986). The featherless areas in the chickens observed in this study were fairly consistent across the population, consistent with the Na gene being fully dominant as stated by Stevens (1991). The Na gene is on chromosome 1, which also carries

the gene for pea comb (P), and blue egg (O), however, no significant linkage was said to occur (Bitgood *et al.*, 1980). In this study, Peel Neck chickens consistently had single combs and tinted or light brown eggs. Most known populations of fowl having the naked neck trait are present in parts of the world that have a hot and humid climate (Merat, 1986). The Peel Neck is another example of fowl with this trait being raised under tropical conditions. Further studies are necessary to ascertain whether these chickens have any production-trait advantages over normally feathered chickens when kept in hot and humid conditions. This is of particular

relevance for chickens that are raised under challenging situations, for example as scavengers or in backyard systems.

Acknowledgements

I would like to thank Dominique Charron for her useful contributions.

References

- Bitgood, J.J., Shoffner, R.N., Otis, J.S. & Briles, W.E.** 1980. Mapping of the gene for pea comb, blue egg, barring, silver, and blood groups. *Poultry Science*, 59, 1686-1693.
- Bodo, I., Kovacs, G. & Ludrovsky, F.** 1990. The Naked Neck fowl. *Animal Genetic Resource Information*, 7, 91-97.
- Cook, A., Rivera, T., Moguel, J., Keane, B., & Ferraes, N.** 1997. Possible strategies for reducing mortality amongst backyard poultry in Yucatan, Mexico. *Proceedings of the 8th International Symposium on Veterinary Epidemiology and Economics; Epidemiol. Santé Anim*, 31-32:02.A16.
- Crawford, R.D.** 1976. Incomplete dominance of the gene for Naked Neck in the domestic fowl. *Poultry Science*, 55, 820-822.
- Davenport, C.B.** 1944. The bare necks. *Journal of Heredity*, 5, 374.
- Hertwig, P.** 1933. Geschlechtsgebundene und autosomale Koppelungen bei Hühnern. *Verhandlungen des deutschen Zoologischen Gesellschaft* 112, 118.
- Mallia, J.G.** 1998. Indigenous domestic turkeys of Oaxaca and Quintana Roo, Mexico. *Animal Genetic Resource Information*, 23, 69-78.
- Merat, P.** 1986. Potential usefulness of the Na (Naked Neck) gene in poultry production. *World's Poultry Science Journal*, 42, 124-142.
- Pesti, G.M., Leclercq, B., Chagneau, A-M & Cochard, T.** 1996. Effects of the Naked Neck (Na) gene on the sulfur-containing amino acid requirements of broilers. *Poultry Science*, 75, 375-380.
- Scott, T. & Crawford, R.D.** 1977. Feather number and distribution in the throat tuft of Naked Neck chicks. *Poultry Science*, 56, 686-688.
- Stevens, L.** 1991. Genetics and evolution of the domestic fowl. Cambridge University Press, Cambridge, U.K.; pp. 306.
- Warren, D.C.** 1933. Nine independently inherited autosomal factors in the domestic fowl. *Genetics*, 18, 68-81.
- Zein-El-Dein, A., Merat, P. & Bordas, A.** 1984. Composition corporelle de poulets "Cou nu" ou normalment emplumes selon le taux proteique de la ration. *Genet. Sel. Evol.*, 16, 491-502.

European approaches to conservation of farm animal genetic resources

D.L. Simon

*Institute of Animal Breeding and Genetics, School of Veterinary Medicine,
Buenteweg 17p, 30559 Hanover, Germany*

Summary

Based on several sources of information an overview has been given on the development, present situation and problems of conservation of animal genetic resources in Europe.

Presently, 1 029 breeds of cattle, sheep, goats, pigs, horses and asses are registered by the EAAP-Animal Genetic Data Bank, Hanover. 42.8 % of the breeds are classified as being 'at risk'. More than 360 conservation programmes are underway, which, however, in many cases seem to be operated independently of the status of endangerment and of similar breeds in other countries.

The primary objectives of conservation in Europe, i.e. 'conservation for potential use, later' and 'conservation for cultural reasons', are different from the objective 'conservation for sustainable use, now', which is primarily expressed for developing countries. Different objectives call for different answers to questions, such as: are breeds appropriate units of genetic diversity, how should endangerment be defined, what should be conserved and is incrossing and selection compatible with conservation?

In view of the large number of breeds 'at risk' and of similar breeds existing in different countries as well as the high costs of conservation it is concluded that characterisation of breeds for genetic uniqueness is presently the most urgent task in conservation. This requires effective co-operation across national borders in Europe.

Résumé

Sur la base de différentes sources d'information on présente une révision sur le développement, la situation actuelle et les problèmes de la conservation des ressources génétiques animales en Europe.

Actuellement 1 029 races de bovins, ovins, caprins, porcins et équins sont enregistrés dans la Base de Données sur les Ressources Génétiques Animales de la FEZ à Hannover. Parmi ces races, 42,8% sont classées dans la catégorie "à risque". En ce moment plus de 360 programmes de conservation sont en oeuvre, cependant, dans plusieurs des cas, ils semblent opérer indépendamment du niveau de danger de disparition et de la présence des mêmes races dans d'autres pays.

Les objectifs principaux de la conservation en Europe, tels que "la conservation pour l'utilisation potentielle future" et "la conservation pour des raisons culturelles", sont bien différents des objectifs de "conservation pour l'utilisation durable actuellement" présentés comme prioritaires par les pays en voie de développement. Les différents objectifs mènent à différentes réponses aux questions telles que: Est-ce que les races sont des unités appropriées pour la diversité génétique? Comment pourrait-on définir le niveau de danger? Que devrait-on conserver et est-ce que le croisement et la sélection sont compatibles avec la conservation?

Etant donné le nombre de races "à risque" et la présence de ces mêmes races dans d'autres pays, ainsi que le coût élevé de la conservation, on conclut que la caractérisation des races dans le seul but génétique est

actuellement le thème plus urgent de la conservation. Ceci nécessite d'une coopération effective entre tous les pays européens.

Key words: *Genetic diversity, Farm animals, Endangerment, Conservation programmes, Objectives, Europe*

Introduction

Europe is a heterogeneous region of the world with 46 or so countries, several supra-national institutions and many Non-Governmental Organisations (NGOs). As a result approaches to conservation of farm animal genetic resources (FAGR) differ not only in the observed objectives and species but also in length of involvement and, possibly, in the effectiveness of actions in this field.

According to FAO's World Watch List for domestic animal diversity (1995), about 34 per cent of the so far listed breeds of the major farm animal species; cattle, goats, horses, pigs and sheep are bred in Europe and about 69 per cent of the active conservation programmes of breeds at risk of the mentioned species are underway in this region. Obviously, Europe can play an important part in the maintenance of the world's farm animal genetic resources.

The activities in Europe started some time before the UNCED Rio Conference in 1992 formulated the Convention on Biological Diversity. One significant milestone in the process of growing awareness of conservation was the informal and later formal foundation of the Rare Breeds Survival Trust (RBST) in 1968 and 1973, respectively, and the foundation of the Rare Breeds Farm Park, 1969, at Warwickshire in the United Kingdom. Subsequent milestones were:

1969 Discussion of 'Needs and methods of gene conservation in animal breeding' in the Genetics Commission of the European Association for Animal Production (EAAP), in Helsinki (Maijala, 1970).

- 1970 Use of markers to estimate genetic distances among breeds (Kidd and Pirchner, 1971).
- 1979 Animal breeding scientists propose a definition of the status of endangerment of breeds and criteria for conservation of endangered breeds (Deutsche Gesellschaft für Züchtungskunde, 1979).
- 1979 Foundation of the Nordic Working Party on Animal Gene Banks for the Scandinavian countries (Maijala *et al.*, 1992).
- 1980 Set-up of the Working Group on Animal Genetic Resources (EAAP-WGAGR) by the Commission on Animal Genetics of the European Association for Animal Production.
- 1983 Survey by the EAAP-WGAGR on breeds and country populations in Europe (Maijala *et al.*, 1984).
- 1987 Set-up of the EAAP Animal Genetic Data Bank (EAAP-AGDB) at the Institute of Animal Breeding and Genetics, Hanover (Simon, 1990).
- 1992 Commission of European Communities 'Workshop and Training Course on data collection, conservation and use of animal genetic resources' in Hanover.
- 1993 EAAP-publication No. 66 'Genetic diversity of European livestock breeds', with status of endangerment and formation of groups of similar breeds.
- 1994 Nomination of National Focal Points in FAO Member-Countries of Europe as national co-ordinators for conservation of FAGR.
- 1996 INTERNET presentation of information of European breeds by the EAAP-Animal Genetic Data Bank, Hanover, and INTERNET-presentation of information of the FAO Domestic Animal Information System DAD-IS, Rome.

Conservation of farm animal genetic resources is a continuous process, which in Europe is taking place on several levels and with different kinds of actions. Earlier reports

on the situation in Europe were given by Maijala *et al.* (1984), as a report of the EAAP-Working Group on Animal Genetic Resources, Simon and Buchenauer (1993), as a report of the EAAP Animal Genetic Data Bank, and by Ollivier *et al.* (1994), with concluding remarks on the situation in different regions of Europe and on urgent tasks. Since then the nomination of National Co-ordinators for conservation in European countries and the forthcoming of the EC-Regulations 2078/92 and 1467/94 have given additional strength to the idea of conservation of FAGR in this region.

For the preparation of this paper, information of the following sources could be used: From the EAAP-AGDB, Hanover, information on breeds of the major farm animal species; cattle, sheep, goats, pigs, horses and asses, as registered until February 1997; in addition information from the National Organisations of EAAP, the National Focal Points for Animal Genetic Resources of European countries, the European Commission in Brussels, from many NGOs and from individual experts who are active in this field.

The intention of this paper is to collect present information of various sources and form one integrated picture of approaches to conservation of farm animal genetic resources in Europe and to come to conclusions on priorities of actions in this field.

Quantity of Farm Animal Genetic Resources, Number of Breeds

Following the practice of the EAAP-WGAGR, the quantity of farm animal genetic resources is measured in terms of breeds. This term is used for a group of similar interbreeding animals within a country, which, according to the people who work with them, should be regarded as a breed. The term 'breed' includes groups of animals, which by other people may be called strain, variety or line. A

synonymous term could be 'country population' as explained by Maijala *et al.* (1984).

The number of breeds of the major farm animal species; cattle, sheep, goats, pigs, horses and asses which have been recorded by the EAAP-AGDB until 1997 is presented in table 1.

In 36 countries a total of 1 029 breeds is recorded. Only breeds are listed for which a reasonable amount of information could be obtained. The number of breeds of the individual species; cattle, sheep, goats, pigs, horses and asses is 311, 338, 101, 134, 139 and 6, respectively. These numbers are quite impressive; however, due to the recording systems within countries they may include several breeds with the same or similar genetic background. This can be a problem if decisions have to be made, such as which one out of several endangered breeds should be conserved and which not. It will be of interest to know how many of these breeds are considered to be endangered.

Status of Endangerment of Breeds

The question which criteria should be used to define the status of endangerment of a breed is not settled. Several systems have been proposed (Deutsche Gesellschaft für Züchtungskunde DGfZ, 1979; Maijala *et al.*, 1984; European Commission 1992, 1994; Bodo, 1995; FAO, 1995; Simon, 1995; and others). The procedure applied here is basically the same as that proposed by Simon and Buchenauer (1993). It considers four conditions which represent danger for the continuance of the present genetic makeup of a breed:

- Low number of breeding herds and decreasing population size, each as an indicator of imminent danger of the loss of the breed in the near future,
- 'immigration' or use of animals of other breed(s) for reproduction, as a factor in the genetic change of the breed,

Table 1. Number of breeds in six farm animal species in European countries (EAAP-Animal Genetic Data Bank, 2/1997).

| Country | Species | | | | | | Total |
|----------------|---------|-------|-------|------|--------|-------|-------|
| | Cattle | Sheep | Goats | Pigs | Horses | Asses | |
| Albania | 5 | 8 | 8 | 4 | 4 | 0 | 29 |
| Austria | 8 | 2 | 0 | 0 | 2 | 0 | 12 |
| Belgium | 10 | 6 | 3 | 3 | 4 | 0 | 26 |
| Bulgaria | 2 | 0 | 0 | 0 | 0 | 0 | 2 |
| Croatia | 5 | 4 | 1 | 1 | 4 | 0 | 15 |
| Cyprus | 2 | 2 | 2 | 3 | 0 | 0 | 9 |
| Czech Republic | 1 | 1 | 0 | 1 | 1 | 0 | 4 |
| Denmark | 5 | 4 | 1 | 2 | 2 | 0 | 14 |
| Estonia | 3 | 0 | 0 | 1 | 1 | 0 | 5 |
| Finland | 4 | 2 | 1 | 2 | 11 | 0 | 20 |
| France | 44 | 55 | 6 | 17 | 31 | 0 | 153 |
| Germany | 27 | 26 | 5 | 16 | 11 | 0 | 85 |
| Greece | 3 | 12 | 2 | 0 | 2 | 0 | 19 |
| Georgia | 3 | 0 | 0 | 1 | 0 | 0 | 4 |
| Hungary | 1 | 3 | 0 | 1 | 6 | 0 | 11 |
| Iceland | 2 | 2 | 1 | 0 | 1 | 0 | 6 |
| Ireland | 12 | 5 | 1 | 2 | 5 | 0 | 25 |
| Italy | 31 | 54 | 29 | 9 | 20 | 5 | 148 |
| Latvia | 1 | 0 | 0 | 1 | 0 | 0 | 2 |
| Lithuania | 2 | 0 | 0 | 1 | 0 | 0 | 3 |
| Luxembourg | 4 | 1 | 0 | 1 | 3 | 0 | 9 |
| Netherlands | 4 | 11 | 2 | 3 | 3 | 0 | 23 |
| Norway | 3 | 6 | 1 | 2 | 4 | 0 | 16 |
| Poland | 4 | 8 | 0 | 8 | 2 | 0 | 22 |
| Portugal | 8 | 9 | 3 | 1 | 3 | 0 | 24 |
| Romania | 5 | 5 | 1 | 7 | 0 | 0 | 18 |
| Slovakia | 2 | 3 | 0 | 1 | 0 | 0 | 6 |
| Slovenia | 3 | 2 | 0 | 3 | 1 | 0 | 9 |
| Spain | 34 | 37 | 18 | 3 | 1 | 1 | 94 |
| Sweden | 2 | 3 | 1 | 0 | 2 | 0 | 8 |
| Switzerland | 8 | 9 | 9 | 3 | 3 | 0 | 32 |
| Ukraine | 6 | 0 | 0 | 3 | 0 | 0 | 9 |
| United Kingdom | 36 | 56 | 4 | 14 | 10 | 0 | 120 |
| Other* | 21 | 2 | 2 | 20 | 2 | 0 | 47 |
| Total | 311 | 338 | 101 | 134 | 139 | 6 | 1029 |

*former CSFR, USSR, Yugoslavia

Table 2. Assumed maximum values of inbreeding in 50 years of conservation, F-50 (%), and resulting range of effective population size N_e per class of endangerment.

| Species | Class of endangerment | | | | |
|--------------|-----------------------------|--|--|---------------------------|---------------------------------------|
| | 1) Not endang. (≤10%) | 2) Potentially endang. (11-20%) | 3) Minimally endang. (21-30%) | 4) Endang. (31-40%) | 5) Critically endang. (>40%) |
| Pigs | ≥ 157 | 156- 74 | 73- 47 | 46- 33 | < 33 |
| Sheep+goats | ≥ 95 | 94- 45 | 44- 28 | 27- 20 | < 20 |
| Cattle | ≥ 67 | 66- 32 | 31- 20 | 19- 14 | < 14 |
| Horses/asses | ≥ 52 | 51- 25 | 24- 16 | 15- 11 | < 11 |

- low 'effective population' N_e size as a condition which affects the increase of inbreeding as well as random drift of the population's gene frequencies.

A three-step procedure is applied.

Firstly, definition of species' specific minimum values of effective population size N_e for five classes of endangerment, depending on the maximum values of acceptable inbreeding, F-50, after 50 years of conservation. For the five classes of endangerment the following maximum values of F-50 were assumed:

- 1) not endangered ≤10%,
- 2) potentially endangered 10-20%,
- 3) minimally endangered 21-30%,
- 4) endangered 31-40%,
- 5) 5 critically endangered >40%.

The corresponding maximum increase of inbreeding per generation ΔF is deduced from F-50 by solving formula (1)

$$F_g = 1 - (1 - \Delta F)^g \quad (\text{Falconer, 1989}) \quad (1)$$

for $\Delta F = 1 - (1 - F_g)^{1/g} \quad (2)$, where $F_g = F-50$.

The generation interval y (in years) with the resulting number of generations g in 50 years are assumed for the six species as follows (y/g): pigs 1.5/33, sheep and goats 2.5/20, cattle 3.5/14, horses and asses 4.5/11.

The required minimum effective population size per species and class of endangerment is deduced from ΔF per generation by formula (3)

$$N_e = 1/(2\Delta F) \quad (\text{Falconer, 1989}) \quad (3)$$

The resulting values of effective population size N_e per species and class of endangerment are listed in table 2.

Secondly, for the individual breed computation of the effective population size N_e' by means of the formula (4)

$$N_e' = 4mf/(m + f) \quad (\text{Falconer, 1989}) \quad (4)$$

with m and f the number of male and female breeding animals, respectively, which are used for the reproduction of the breed. We defined f as the number of females which are registered in a herdbook (since these allow pedigree-information in planning of matings for reproduction to be observed), and which are used in the order of 100 per cent for purebreeding. In case these requirements are not met f is estimated as $1/4$ of the number of unregistered females U (or of the total population size T), times the percentage of purebreeding p :

$$f = pU/4 \quad \text{or} \quad f = pT/4.$$



Figure 1. Sow of breed German Sattel Back. For seven similar breeds conservation programmes are under way in six European countries. (Photo: Anonymous)

Table 3. Number of breeds classified for endangerment.

| Species | Total | Class of endangerment | | | | | "At risk" (2-5) |
|---------|-------|-----------------------|-----------------------------|---------------------------|---------|----------------------------|--------------------|
| | | Not endang. | Poten- tially endang. | Mini- mally endang. | Endang. | Criti- cally endang. | |
| | | (1) | (2) | (3) | (4) | (5) | |
| Cattle | 305 | 165 | 87 | 24 | 9 | 20 | 140 |
| Sheep | 322 | 210 | 58 | 21 | 7 | 26 | 112 |
| Goats | 88 | 64 | 8 | 7 | 4 | 5 | 24 |
| Pigs | 126 | 61 | 23 | 10 | 3 | 29 | 65 |
| Horses | 135 | 59 | 51 | 12 | 5 | 8 | 76 |
| Asses | 6 | 3 | 1 | 0 | 0 | 2 | 3 |
| Total | 982* | 562 | 228 | 74 | 28 | 90 | 420 |

*47 additional breeds - due to missing information - could not be classified.

For a known number of registered female breeding animals that are used for purebreeding only with a percentage of p f is reduced accordingly by multiplication with p .

If m , the number of male breeding animals used for reproduction of the breed is not known m is estimated from f as $m = f/30$, i.e. by assuming a mating ratio of $m:f = 1:30$.

If m is less than 40 and the number of males MC is known of which cryoconserved semen is available, the number of males m is increased by addition of the value of $MC/3$, i.e. it is assumed that $1/3$ of the number of males with cryoconserved semen can be regarded as additional males which are available for reproduction of the breed.

By comparing N_e of the breed with the minimum values of N_e for the relevant species in table 2 the breed is allocated to one of the five classes of endangerment.

Thirdly, downgrading the breed into one class of higher endangerment for each one of the following additional conditions:

- The number of breeding herds is less than 10 and the number of female breeding animals is below 500,
- the number of female breeding animals is decreasing and already below 1000,
- the percentage of matings for reproduction of the breed with animals of other breed(s) is equal or higher than 20 percent.

Compared with the system of Simon and Buchenauer (1993) the starting points for acceptable F_{50} values of the classes 1 to 4 were raised and the maximum value of incrossing was increased from 10 to 20 percent. This resulted in fewer downgradings and a higher percentage of breeds classified as being not endangered.

Following this system 420 or 42.8 % of a total of 982 breeds with sufficient information were allocated to classes 2 to 5 and as such were classified as breeds 'at risk' (table 3).

The species with the highest percentage of breeds classified as being 'at risk' was horses (56.3%), followed by pigs (51.6), asses (50.0%) and cattle (45.9%). Sheep and goat breeds appear to be least endangered with 34.8% and 27.3% 'at risk', respectively. It is interesting to note that in all species the number of breeds

classified as being 'critically endangered' is remarkably higher than the number of breeds classified as being 'endangered'.

Conservation Activities

Apart from breeding companies for poultry and pigs, nothing is known about activities of commercial breed - societies on conservation of rare breeds. However, in Germany some breed - societies are committed to the herdbook operation for a rare breed of their region. Actual conservation is mainly performed by farmers, hobby-farmers, research institutions and state-farms.

Live-animal conservation programmes

Live-animal conservation in reproducing herds is the most frequently adopted conservation method. It is an attractive method, allowing adaptation of the breed to changing production and environmental conditions and an immediate use of animals for evaluation, research and commercial breeding. Information on the number of live-animal conservation programmes was obtained from three sources: The EAAP-Animal Genetic Data Bank (EA), the newly nominated National Focal Points in Europe (NFP) and from the European Commission (D. Dessylas, Brussels, 1997, personal communication) (EU).

The information from the EU is related to endangered breeds that are supported according to EEC Regulation 2078/92. So far this support has been restricted to local breeds in danger of extinction of the cattle, sheep, goat and *equidae* species in the 15 EU-Member States.

A total of 365 programmes is registered by EA for the major farm animal species, 285 of these or nearly 78 per cent are underway in EU-Member States (table 4). The total sum reported by NFP is in the same order (334); however, within countries and within species the numbers of the reported conservation programmes can be quite different (e.g. in France, Italy and Portugal). The NFP also

Table 4. Live animal conservation programmes in countries of Europe, as registered by EAAP Animal Genetic Data Bank EA, National Focal Points NF and European Union EU, respectively; NR = No response to request

| Country | Cattle | | | Sheep | | | Goats | | | Horses | | | Pigs | | Total per country | | |
|--------------------------------------|--------|----|----|-------|-----|----|-------|----|----|--------|----|----|------|----|-------------------|-----|-----|
| | EA | NF | EU | EA | NF | EU | EA | NF | EU | EA | NF | EU | EA | NF | EA | NF | EU |
| <u>a) 15 EU-member countries</u> | | | | | | | | | | | | | | | | | |
| Austria | 6 | NR | 10 | 1 | NR | 4 | | NR | 4 | 2 | NR | 6 | | NR | 9 | NR | 26 |
| Belgium | 2 | 3 | 2 | | 8 | | | | | | 2 | 2 | | | 2 | 13 | 8 |
| Denmark | | 4 | | 1 | 1 | | 1 | 1 | | 1 | 3 | | 2 | 2 | 5 | 11 | 0 |
| Finland | 3 | 3 | 3 | 1 | 2 | 1 | | | 1 | 1 | 1 | 1 | | | 5 | 6 | 6 |
| France | 40 | 18 | 14 | 43 | 22 | 9 | 1 | 5 | 2 | 2 | | 15 | 5 | 6 | 91 | 51 | 40 |
| Germany | 14 | 11 | 9 | 12 | 14 | 12 | 2 | 3 | 3 | 7 | 12 | 12 | 6 | 4 | 41 | 44 | 36 |
| Greece | | 2 | | | 8 | | | 2 | | 1 | 2 | | | | 1 | 14 | 0 |
| Ireland | | 3 | 3 | 1 | 2 | 7 | | | 1 | | | 1 | | 2 | 1 | 7 | 5 |
| Italy | 25 | 10 | 16 | 1 | 7 | 19 | | 11 | 7 | 7 | 3 | 18 | 4 | 4 | 37 | 35 | 60 |
| Luxembourg | 2 | 2 | | | | | | | | 1 | 1 | 1 | | | 3 | 3 | 1 |
| Netherlands | 3 | 5 | | 4 | 4 | | 2 | 1 | | 1 | 2 | | | | 10 | 12 | 0 |
| Portugal | 6 | 2 | 4 | 6 | 1 | 3 | 1 | | 2 | 3 | | | | | 16 | 3 | 9 |
| Spain | 12 | NR | 25 | 4 | NR | 13 | 2 | NR | 4 | | NR | 12 | 2 | NR | 20 | NR | 54 |
| Sweden | 2 | 3 | 3 | 2 | 3 | 3 | | 2 | 2 | | 2 | | | 1 | 4 | 11 | 8 |
| United Kingdom | 12 | 9 | | 16 | 23 | | 2 | 7 | | 4 | 6 | | 6 | 2 | 40 | 47 | 0 |
| EU-Countries | 127 | 75 | 89 | 92 | 95 | 70 | 11 | 32 | 26 | 30 | 34 | 68 | 25 | 21 | 285 | 257 | 253 |
| <u>b) 18 Non EU-Member countries</u> | | | | | | | | | | | | | | | | | |
| | 16 | 21 | | 22 | 21 | | 12 | 3 | | 19 | 24 | | 11 | 8 | 80 | 77 | |
| Total | 143 | 96 | 89 | 114 | 116 | 70 | 23 | 35 | 26 | 49 | 58 | 68 | 36 | 29 | 365 | 334 | 253 |

Table 5. Ex-situ keeping of farm animal breeds in zoos and farm parks on 124 locations in Germany. Raw data from Falge (1996); N_e = effective population size.

| Species | Number of | | Animals per "herd" | Average number of | | N_e |
|----------|-----------|---------|-----------------------|----------------------|--------------------|-------|
| | Breeds | "Herds" | | Animals per breed | Males per breed | |
| Cattle | 28 | 136 | 4.9 | 20.3 | 6.3 | 20.7 |
| Sheep | 30 | 253 | 10.3 | 86.9 | 15.9 | 54.1 |
| Goats | 24 | 172 | 8.6 | 61.3 | 15.1 | 48.2 |
| Pigs | 12 | 131 | 5.4 | 59.3 | 19.5 | 59.7 |
| Horses | 23 | 120 | 5.0 | 26.1 | 7.0 | 22.1 |
| Asses | 6 | 80 | 3.7 | 48.8 | 14.8 | 45.9 |
| Chickens | 42 | 112 | 7.7 | 20.6 | 5.6 | 18.7 |
| Geese | 10 | 48 | 3.3 | 16.0 | 6.3 | 19.5 |
| Ducks | 12 | 39 | 5.3 | 17.3 | 7.5 | 21.8 |
| Total | 187 | 1091 | 6.0 | 39.6 | 10.9 | 31.1 |

reported live-animal conservation programmes for additional species, i.e. for asses, dogs, rabbits, chickens, ducks, geese and even for bees, fishes and silkworms. It can be noted that the EU-Member States Denmark, Greece, Netherlands and the United Kingdom have not participated in the EEC-project 2078/92 so far.

Contributions of zoos and farm parks

As already mentioned, conservation of endangered breeds of farm animal species is generally performed on farms. However, such animals are also kept 'ex situ' in zoos, farm parks and in so-called Ark-farm projects (Seibold, 1996). Falge (1996) reported animal numbers in 124 institutions of this kind in Germany (table 5):

Animals of 187 breeds of 9 farm animal species are kept in these institutions. The average number of males and of total animals per breed, spread over several locations, is quite low (10.9 and 39.6, respectively). Computation of the effective population size

N_e according to formula (4), chapter 3, resulted in values from $N_e = 19$ for geese and ducks to $N_e = 60$ for pigs. These values appear rather low if long-term conservation should be achieved.

Thirty-five farm-parks with *ex-situ* conservation of several rare breeds are reported from seven European countries, with 25 farm-parks alone in the United Kingdom (J. Guenterschulze, Warder, 1997, personal communication). In the UK, farm-parks are visited on average approximately by 100 000 people per year (L. Alderson, Warwickshire, 1997, personal communication). The Rare Breeds Survival Trust in the UK and the Gesellschaft zur Erhaltung alter und gefährdeter Haustierrassen, GEH, in Germany developed approval schemes for farm-parks in order to ensure given standards (Chiperzak *et al.*, 1995). No information is available on the number of animals and on the farm animal breeds and species which are kept in zoos and farm-parks in the whole region of Europe. The specific value of zoos and farm-parks is that they offer visible

Table 6. Programmes for cryoconservation of semen and embryos. T=total, IS=programmes with information on number of sires involved. Data from EAAP-AGDB.

| Species | Semen | | Embryos | |
|---------|-------|-----|---------|----|
| | T | IS | T | IS |
| Cattle | 194 | 173 | 74 | 49 |
| Sheep | 46 | 39 | 6 | 3 |
| Goats | 18 | 15 | 1 | 1 |
| Pigs | 30 | 29 | 0 | 0 |
| Horses | 26 | 25 | 4 | 3 |
| Asses | 0 | 0 | 0 | 0 |
| Total | 314 | 281 | 85 | 56 |

evidence of endangered breeds to the public, hence contributing to an increased awareness of the need of conservation.

Cryoconservation of semen and embryos

Cryoconservation of gametes and embryos is a quick and rather inexpensive way to prevent the loss of the genetic potential of a breed (Brem *et al.*, 1984). Properly structured stores of frozen semen and embryos can also be used to support live animal conservation programmes with a minimum increase of inbreeding (Simon, 1993). Table 6 summarises the information on cryoconservation as reported by the EAAP-AGDB.

The largest number of projects of semen and embryo storage is reported for breeds of cattle. Here the number of programmes for semen (194) is even higher than the number of live-animal conservation programmes (139, table 4). Possibly, these numbers include projects of cryostored semen for commercial use. Information on the number of males or sires represented in stored semen and embryos is important to judge their usefulness for the re-activation of an extinct

breed. As can be seen in table 6 this information is available in most cases, though completion appears necessary.

Activities on a supra-national level in Europe

Several organisations and institutions are active in projects of conservation which involve more than one country in Europe. In chronological order – according to the start of their activities – the following have to be mentioned:

1. EAAP, the European Association for Animal Production, regularly offers a platform for the presentation of papers on topics of conservation of FAGR at its annual meetings (see

'milestone' 1969, chapter 1). Its Commission on Animal Genetics established the EAAP Working Group on Animal Genetic Resources in 1980, whose main objectives are the study, documentation and cataloguing of conservation and development of animal genetic resources in Europe. In 1987 the working group suggested setting up a data bank, which, with the support of the Deutsche Forschungsgemeinschaft DFG, was founded as the EAAP- Animal Genetic Data Bank at the School of Veterinary Medicine, Hanover, in the same year. Since then the volume and quantity of European breed resources could steadily be increased. During the years 1989-1992 the Hanover data bank accepted responsibilities as 'EAAP-FAO-Global Animal Genetic Data Bank'; since then the responsibility of a Global Animal Genetic Data Bank has been transferred to the newly established FAO Domestic Animal Information System DAD-IS in Rome. Since 1997 EAAP has been acting as co-ordinator for the EU-concerted action 'A Permanent Inventory of European farm animal genetic resources'. This project is



Figure 2. Heifers of breed Tyrol Grey. For five similar breeds conservation programmes are under way in five European countries. (Photo: Averdunk).

supported by EC-Regulation 1467/94 and delegates additional responsibilities to EAAP.

2. NAGB, the Nordic Working Party on Animal Gene Banks, was established in 1979 by the five Scandinavian countries Denmark, Finland, Iceland, Norway and Sweden. It has mapped out conservation needs and activities and built a Nordic Information Centre with a data bank. The Nordic Council of Ministers, NCM, is funding the secretary of the working party, the operation of the Nordic Data Bank and specific research (Maijala *et al.*, 1984). Breed information of Scandinavian countries is transferred via the Nordic Data Bank to the EAAP-AGDB Hanover.
3. DAGENE, the Danubian Countries Alliance for Conservation of Genes in Animal Species, was founded in 1989 as a group of individual experts, NGOs and governmental institutions of 10 countries of the Danubian region which are interested in conservation of FAGR in this area. Main objectives are the exchange of information and co-operation in similar projects and the organisation of meetings on regional aspects (I. Bodo, Budapest, 1997, personal communication).
4. Commission of the European Communities. The European Union has become an important factor in improving co-ordination and actual support for conservation of FAGR in the 15 Member-States: Council Regulation No. 2078/92 'On agricultural production methods compatible with the requirements of the protection of the environment and the maintenance of the country side'. The scheme allows support of farmers who 'rear animals of local breeds in danger of extinction'. An EU-specific system for the classification of endangerment is used, which presently allows the promotion of 253 breeds with up to 100 ECU per livestock unit. - Council Regulation 1467/94 'On the

conservation, characterisation, collection and utilisation of genetic resources in agriculture'. The main objective is to co-ordinate and promote existing work on plant and animal genetic resources in the Member States. Until now 12 plant and 3 animal projects have been supported, the animal side having been severely underrepresented so far. One of the projects deals with the initiation of a 'Permanent Inventory of European farm animal genetic resources and of current work on conservation, characterisation, collection and utilisation of those resources'; it is hoped that this can be developed into an efficient instrument. - AIRE 2066, the concerted action project 'Analysis of genetic diversity to preserve future breeding option'. The main objective is to co-ordinate the work of 27 participating laboratories in 14 countries, in particular to use the same set of DNA-markers for assessing genetic diversity within and between cattle breeds. In addition, a Cattle Diversity Data Base, CaDBase, was set up in Edinburgh with a link to the EAAP-AGDB, Hanover.

5. SAVE, Safeguard for Agricultural Varieties in Europe, was founded in 1995 as an umbrella-organisation for NGOs in Europe. Main objectives are: co-ordination of similar activities in different countries, development of awareness of conservation, exchange of know-how and actual support for specific conservation projects. SAVE became particularly active in countries of Eastern Europe (Gruenenfelder, 1995; W. Kugler, St.Gallen, 1997, personal communication).

Number of Registered Conservation Programmes and Its Relation to the Degree of Endangerment

Table 4 shows that the number of registered live-animal conservation programmes is quite high. The differences in numbers reported by

the three sources, EAAP, National Focal Points and European Union, can be explained in part by different countries involved in the respective survey, different ways of assessing endangerment, different requirements for support, (e.g. number of female breeding animals, accepted herdbook) and possibly by different interpretation of what is meant by conservation.

It is of special interest to look at the relation between the percentage of conservation programmes and the class of endangerment of breeds. The figures are presented in table 7, including the coefficient of Spearman's rank correlation r_s . The average of the five species shows no tendency of an increased proportion of conservation programmes in classes of higher endangerment. Spearman's rank correlation $r_s=0.12$ is low and not significant. None of the five species has the highest proportion of conservation programmes in class five of the breeds with the highest risk status. For sheep, goat and horse breeds the rank correlation even turns out to be negative (-0.44, -0.62 and -0.32, respectively). In class 1, with breeds classified as 'not endangered', conservation programmes are reported for 32% of the breeds. This is remarkable since the system of assessment of endangerment applied here appears to be rather severe, compared with the system used by FAO (1995) (see chapter "Assessment of breeds for endangerment").

It is difficult to explain these results. However, it seems to be meaningful in countries of Europe to examine more closely the objective of conservation, the system of assessing the status of endangerment of breeds and the question whether all breeds which have been classified as being endangered equally deserve to be conserved.

Table 7. Percentage of breeds with live animal conservation programmes by class of endangerment. Rank correlation r_s between class of endangerment and percentage of conservation programmes.

| Species | Class of endangerment | | | | | R_s |
|---------|-----------------------|-------------------|-------------------|----------------------|--------------------|------------|
| | 1 Not end. | 2 Pot. end. | 3 Min. end. | 4 Endan- gered | 5 Crit. end. | |
| Cattle | 38.8 | 59.8 | 41.7 | 66.7 | 50.0 | 0.60 n.s.* |
| Sheep | 30.0 | 48.3 | 66.7 | 28.6 | 19.2 | -0.44 n.s. |
| Goats | 29.7 | 12.5 | 14.3 | 25.0 | 0 | -0.62 n.s. |
| Pigs | 13.1 | 34.8 | 60.0 | 33.3 | 37.9 | 0.62 n.s. |
| Horses | 42.4 | 25.5 | 41.7 | 40.0 | 37.5 | -0.32 n.s. |
| Total | 32.0 | 44.9 | 48.6 | 42.9 | 33.3 | 0.12 n.s. |

* $p < 0,05$

Table 8. Live animal conservation programmes LAC for 'similar' breeds SB, total and in classes of endangerment

| Subgroup of similar breeds, formed by EAAP-AGDB | Total number of | | | Number of SB and LAC | | | |
|--|---------------------------|----|-----|-----------------------------|-----|----------------------------|-----|
| | Coun- tries involv. | SB | LAC | In class 1 (not endang.) | | In class 2-5 (at risk') | |
| | | | | SB | LAC | SB | LAC |
| <u>Cattle</u> | | | | | | | |
| 1.2 Origin Black Pied | 6 | 8 | 6 | 6 | 4 | 2 | 2 |
| 3.7 White Lineback | 4 | 5 | 5 | 4 | 4 | 1 | 1 |
| 5.2 Alpine Brown | 4 | 6 | 4 | 3 | 2 | 3 | 2 |
| 5.4 Iberian Brown | 2 | 11 | 9 | 7 | 6 | 4 | 3 |
| 6.2 Grey Mountain | 5 | 7 | 5 | 4 | 3 | 3 | 2 |
| <u>Sheep</u> | | | | | | | |
| 5.2 S.Europ.Milk Sheep | 6 | 15 | 7 | 11 | 4 | 4 | 3 |
| 8.4 Churra Type | 2 | 8 | 4 | 7 | 3 | 1 | 1 |
| <u>Pigs</u> | | | | | | | |
| 3.1 Saddle Backs | 6 | 8 | 7 | 1 | 0 | 7 | 7 |
| <u>Horses</u> | | | | | | | |
| 5.10 South Europ. Ponies | 4 | 6 | 5 | 3 | 3 | 3 | 2 |
| Total | | 74 | 52 | 46 | 29 | 28 | 23 |



Figure 3. An example of original Black Pied cattle.

Conservation Programmes within Groups of Similar Breeds

For breeds registered in the EAAP-AGDB, an attempt was made to form groups of similar breeds by use of information of breed history, geographic origin, phenotypic appearance, type of use, known genetic background, etc. (Simon and Buchenauer, 1993). For this report an additional analysis was run in order to find out to what extent conservation programmes are underway for similar breeds in different countries. An extract of the results is presented in table 8.

In nine subgroups of similar breeds a total number of 74 breeds is listed; 28 of them were classified as being 'at risk'. The breeds within subgroups are located in 2 to 6 countries, the number of conservation programmes for breeds at risk is $n = 23$. Obviously the decisions to conserve endangered breeds are made without taking into account the existence of conservation programmes for similar breeds in other countries. The situation appears even worse if we realise that

for 46 breeds of the same subgroups, which were classified as being not endangered, 29 additional conservation programmes are reported. This draws attention to the need of clarifying the genetic relationship among breeds and to the need of co-operation across national borders.

Discussion

In the previous chapters it could be shown for Europe that:

- the quantity of farm animal genetic diversity – if expressed in the number of breeds – is still quite large,
- on average some 43 per cent of these breeds have to be regarded as more or less endangered,
- and that an impressive number of conservation programmes (~ 360) is underway.

However, it has become also apparent that in many cases decisions to conserve breeds seem to be not only independent of their status of endangerment but also of the existence of conservation programmes for similar breeds in other countries. This calls for

a closer look at the essential elements of present approaches to conservation in Europe.

Breeds as appropriate units of genetic diversity and of conservation

All groups of interbreeding animals which according to the practice in the reporting countries should be regarded as breeds are registered as such by the EAAP-AGDB. This number probably includes several breeds with the same or similar background. The total number of registered breeds of a species therefore has to be regarded as an overestimate of the available genetic diversity.

From genetic theory we know that genetic diversity of populations is a function of the frequencies of genes and of gene combinations. As a consequence, the objective of conservation in principle should be genes and gene combinations. It is interesting to note that the UN-Agenda 21 (1992), in chapter 15 'Conservation of biological diversity', calls 'to conserve and maintain genes, species and ecosystems', not of breeds.

However, our ability to identify genes of farm animal species and their interaction has been very limited, so far. In addition, for several reasons, farm animal species are subdivided into breeds as operating units within which the decisions and actions for breeding are performed. Therefore, for the time being, it is realistic to use breeds as indicators of available genetic diversity of a species. However, if it comes to conservation of genetic diversity, preference should be given to those breeds which can be assumed to be 'containers' of a unique genetic potential, i.e. of genes or gene combinations which are not available in other breeds.

Assessment of breeds for endangerment

By use of the system explained in chapter 3 a higher percentage of breeds was classified as being 'at risk' in Europe (42.8 %) than by the FAO-system of WWL-2 (1995) (32.8 %). The two systems differ essentially in three criteria:

- The way of considering a minimum population size, below which a breed should be regarded as being 'at risk',
- the way of dealing with incrossing or migration, i.e. the use of animals of other breed(s) for reproduction,
- and taking account of the number of breeding herds in which the breed is kept

For the minimum population size the FAO-system asks for absolute numbers of breeding animals, i.e. that the total number of breeding females and males are greater than 1000 and 20, respectively; same numbers for breeds of all species.

In contrast, the system we used for our analysis asks for a minimum effective population size N_e – a term from population genetics – of the breed in question, where the minimum value of N_e is deduced from the maximum increase of inbreeding, which appears acceptable in a time period of 50 years of conservation (F-50). Taking account of different generation intervals of different species, species-specific values of N_e are obtained (see table 2).

If the minimum numbers of the FAO-system are transformed into the corresponding effective population size the resulting value of $N_e = 82$ is – compared with the respective values of table 2 – above the value for breeds of horses and asses (52), but below the values for sheep and goats (95) and for pigs (157). In other words breeds of the latter three species will be declared earlier to be more or less endangered by the system used here than by the FAO-system. In addition, in our system only females that are registered in a herdbook – or an estimate of these – are used in estimation N_e of a particular breed. This again increases the probability that the breed will be classified as

Accepted herdbook for six generations, less than 20 per cent contribution by other breeds and thirdly parent breeds used in the formation of the breed are no longer available. In addition: fewer than the following number of breeding females: pigs 500, goats 500, cattle 750, horses 1 000 and sheep 1 500. Last but not least, the factors minimum number of distinct male lines, decreasing population size and low number of breeding units are considered.

This acceptance procedure appears to be a well-founded system with many valuable criteria (although the critical animal numbers seem to be defined independently of the generation intervals of species). However, it will probably be difficult, in general, to supply both the quantity and quality of the required information on those breeds which are approaching endangerment.

Compared with the various systems of risk assessment mentioned above we feel that the system we used for the breeds of the EAAP-AGDB is an acceptable compromise between considering population genetic theory, short term risk factors and obtainable information. Table 9 presents information on the relative importance of the four factors used in our system for the downgrading of a breed from the status of 'not endangered' into classes of endangerment. Summarising all species, 57.9 per cent of the downgradings are due to a low effective population size, 18.7 % are caused by incrossing and 15.1 % by a decreasing number of female breeding animals. A low number of herds affects endangerment only in the order of 8.3 %. However, this may be caused in part by missing information of the number of herds. For breeds of cattle and horses the use of breeding animals of other breeds for reproduction is obviously not uncommon, because 29.8% and 29.9% of the downgradings, respectively, are due to these factors. In pigs incrossing (6.2%) is of least significance to endangerment, whereas the effective population size N_e is of relatively highest importance (71.8%).

Of course, the results of the applied classification system depend on the observed criteria and on the way of combining these into a system of risk assessment. For example it may be questioned whether the assumed values of maximum coefficients of inbreeding, length of generation intervals of species, maximum percentage of incrossing, etc., are the best possible ones and whether formula (4) for estimation of the effective population size is adequate for small populations with decreasing numbers of breeding animals and with generally increased variance of family size. Nevertheless, for a rational approach to conservation it appears necessary to take a position on these criteria.

Different objectives of conservation call for different approaches

One of the main differences among the various systems of risk assessment can be seen in the fact, whether migration, i.e. use of breeding animals of other breeds, is considered as a risk factor or not. The question whether this is meaningful depends on the primary objective of conservation (Simon,1999). From the many statements on conservation objectives of FAGR, e.g. by Bowman (1974), DGfZ (1979), Maijala *et al.* (1984), Simon (1984), UNEP (1992), Blair (1995), Hammond (1995), Cunningham (1996), British Society of Animal Science (1997), three main objectives have become apparent:

1. Subunits of species, such as populations, breeds, lines or strains, which under predominantly favourable production conditions are no longer competitive, may possess – unknown so far – a genetic potential which may become useful for future breeding options. The resulting argument, which we may call 'Conservation for potential use, later' (Simon,1999), is mainly expressed in developed countries of Europe and North America.
2. In regions with predominantly unfavourable production conditions, indigenous populations or breeds – in spite of their usually limited production

potential – generally form the basis of food security for an increasing human population because of their generally good adaptation to harsh production conditions. The argument, which we may call ‘Conservation for sustainable use, now’, is expressed mainly in and for developing countries of the world.

3. Rare breeds can be regarded as part of our living heritage and as such deserve to be preserved for historical, ethical or local reasons. This argument, which we may call ‘Conservation for cultural reasons’, seems to be expressed mainly in developed countries of Europe and North America.

Since objective number 1, ‘conservation for potential use, later’, aims to preserve an unknown genetic potential for requirements that are as yet unknown in far distant unknown future, it is essential to avoid all influences that can change the genetic makeup of the population, or – in terms of population genetics – that can change the Hardy-Weinberg-equilibrium of the population (Simon, 1995; 1999).

This requires avoidance of migration or the use of animals of other populations for reproduction, avoidance of artificial selection of mates in pursuit of defined breeding goals ‘for improvement’ and it requires the minimisation of random drift of gene frequencies and inbreeding by providing a sufficient effective population size N_e (Falconer, 1989). Therefore, it appears necessary to observe migration or incrossing already in the risk assessment, if conservation for potential later use is the primary objective of conservation, which at least in Europe is of high relevance (Simon, 1995).

Objective number 2, ‘conservation for sustainable use, now’, asks for a completely other strategy of conservation. Here, immediate use of endangered breeds is required in order to serve the immediate needs of the human population for food security. Genetic changes of breeds for improvements both by artificial selection within the breeds and by planned incrossing of animals of other, highly-productive breeds

is an essential tool of sustainable use and conservation (see, for example, Rege and Bester, 1998; Mariante and Fernandez-Baca, 1998). For this reason the situation of incrossing need not be considered in risk assessment, if ‘conservation for sustainable use, now’ is the main objective.

Finally, if objective number 3, ‘conservation for cultural reasons’ is the main objective of conservation, the situation of incrossing or use of animals of other breeds can be dealt with in either way. Purists may demand strict purebreeding and may reject any minor ‘contamination’ by ‘foreign blood’. Others may tolerate the introduction of animals of similar breeds as long as the outside appearance of the breed is not severely changed. In other words, the consideration of incrossing is not a major issue in risk assessment, if conservation is pursued mainly for cultural reasons. A similar position may be taken on the question of selection of mates for ‘genetic improvement’ or for adaptation during conservation.

Summing up this section, we can see that the primary objectives of conservation in Europe are different from the ones in developing countries, say in Africa, South America or in Asia. It also follows that different procedures of risk assessment and of practical conservation should be applied (Simon, 1999).

The problem of choice of endangered breeds for conservation

An adequate approach to dealing with this problem is again affected by the primary objective of conservation. Within the context of conservation for cultural reasons, preference for specific breeds is usually expressed by the people or institutions which actually work with the breed. In this situation it is probably not adequate to impose criteria from outside as long as support from outside is not requested. Nevertheless, for example the NGO Rare Breeds Survival Trust requires in its acceptance procedure ‘a distinct

characteristic not found elsewhere', if other requirements are not met completely (Alderson,1995).

For the primary objective 'conservation for sustainable use, now' candidate breeds for the combined goal conservation and improvement should be the most promising adapted local breeds, preferably evaluated on the basis of reliable data of their adaptive value and on their combining ability with highly productive exotic breeds, as explained by Rege and Bester (1998).

For the primary objective 'conservation for potential use, later' priority should be given to those endangered breeds which – unknown so far - could possess a genetic potential which could become valuable in the future and which cannot be expected in other breeds. The main criterion for selection of a breed, therefore, should be the degree of genetic uniqueness or the degree of genetic distance in comparison to other breeds, i.e. both to the more popular breeds and to other endangered breeds, as explained for example by DGfZ (1979), Camussi *et al.*(1985), Weitzman (1993), Barker (1994) and Ollivier,1996).

Bearing in mind the relatively high number of breeds 'at risk' in Europe (Table 3), the availability of similar breeds in different countries and the generally high costs of conservation (Brem *et al.*, 1984; Smith, 1984; Lömker and Simon, 1994), clarification of genetic uniqueness of breeds appears to be one of the most urgent tasks in conservation of FAGR in Europe. This can only be achieved in a satisfying way on a supra-national level, for which effective co-operation among all acting institutions across national borders is required.

References

Alderson, L. 1995. The value of global information for better estimating the vulnerable status of endangered breeds. In R.D. Crawford, E.E. Lister & J.T. Buckley (Eds.) Proceedings of the Third Global

Conference on Conservation of Domestic Animal Genetic Resources, Kingston, Canada, August 1994, 13-23.

Barker, J.S.F. 1994. A global protocol for determining genetic distances among domestic livestock breeds. Proceedings of the 5th World Congress on Genetics Applied to Livestock Production, Guelph, Canada, August 1994, 21, 501-508.

Blair, H.T. 1995. Domestic livestock conservation – is it achieving its goal? In R.D. Crawford, E.E. Lister & J.T. Buckley (Eds.) Proceedings of the Third Global Conference on Conservation of Domestic Animal Genetic Resources, Kingston, Canada, August 1994, 3-11.

Bodo, I. 1995. Minimum number of individuals in preserved domestic animal populations. In R.D. Crawford, E.E. Lister & J.T. Buckley (Eds.) Proceedings of the Third Global Conference on Conservation of Domestic Animal Genetic Resources, Kingston, Canada, August 1994, 57-65.

Bowman, J.C. 1974. Conservation of rare livestock breeds in the United Kingdom. Proceedings of the 1st World Congress on Genetics Applied to Livestock Production, Madrid, Spain, 2, 23-29.

Brem, G., Graf, F. & Kräusslich, H. 1984. Genetic and economic differences among methods of gene conservation in farm animals. *Livestock Production Science*, 11, 65-68.

Camussi, A., Ottaviano, E., Calinski T. & Kaczmarek, Z. 1985. Genetic distances based on quantitative traits. *Genetics*, 111, 945-962.

Chiperzak, J., Guenterschulze, J. & Alderson, L. 1995. The role of rare breeds parks in genetic conservation programs in Canada, Germany and Britain. In R.D. Crawford, E.E. Lister & J.T. Buckley (Eds.) Proceedings of the Third Global Conference

on Conservation of Domestic Animal Genetic Resources, Kingston, Canada, August 1994, 223-229.

Cunningham, P. 1996. Genetic diversity in domestic animals: Strategies for conservation and development. In R.H. Miller, V.G. Pursal & H.D. Norman (Eds.) Beltsville Symposia in Agricultural Research XX, Biotechnology's Role in the Genetic Improvement of Farm Animals, ASA, USA, 13-23

Falconer, D.S. 1989. Introduction to quantitative genetics, 3rd ed., Longman Scientific & Technical, pp. 438.

Falge, R. 1996. Maintenance and conservation of domestic animal resources *ex-situ* in zoological gardens and domestic animal parks. In F. Begemann, C. Ehling & R. Falge (Eds.) Schriften zu genetischen Ressourcen, IGR, Bonn, 5, 60-77.

Gruenenfelder, H.P. 1995. Monitoring programmes in Central and Eastern Europe. In R.D. Crawford, E.E. Lister & R.T. Buckley (Eds.) Proceedings of the Third Global Conference on Conservation of Domestic Animal Genetic Resources, Kingston, Canada, August 1994, 234-239.

Hammond, K. 1995. A global program for conservation of domestic animal diversity. In R.D. Crawford, E.E. Lister & R.T. Buckley (Eds.) Proceedings of the Third Global Conference on Conservation of Domestic Animal Genetic Resources, Kingston, Canada, August 1994, 27-39.

Kidd, K.K. & Pirchner, F. 1971. Genetic relationships of Austrian cattle breeds. Anim. Blood Grps. Biochem. Genet., 2, 145-158.

Lömker, R. & Simon, D.L. 1994. Costs of and inbreeding in conservation strategies for endangered breeds of cattle. Proceedings of the 5th World Congress of Genetics Applied to Livestock Production, Guelph, Canada, August 1994, 21, 393-396.

Maijala, K. 1970. Need and method of gene conservation in animal breeding. Ann. Génét. Sél. anim., 2, 403-415.

Maijala, K., Cherekaev, A.V., Devillard, J.M., Reklewski, Z., Rognoni, G., Simon D.L. & Steane, D. 1984. Conservation of animal genetic resources in Europe. Final report of EAAP Working Party. Livestock Production Science, 11, 3-22.

Maijala, K., Adelsteinsson, S., Danell, B., Gjelstad, B., Vangen, O., & Neimann-Soerensen, A. 1992. Conservation of animal genetic resources in Scandinavia. In L. Alderson & I. Bodo (Eds.), Genetic Conservation of Livestock. CAB International, 2, 30-46.

Mariante, A. da S. & Fernandez-Baca, S. 1998. Animal genetic resources and sustainable development in the Americas. Proceedings of the 6th World Congress of Genetics Applied to Livestock Production, Armidale, Australia, 28, 27-34.

Ollivier, L. 1996. The role of domestic animal diversity in the improvement of animal production. AAA Biotec, Ferrara, 1-11.

Ollivier, L., Bodo I. & Simon, D.L. 1994. Current developments in the conservation of domestic animal diversity in Europe. Proceedings of the 5th World Congress of Genetics Applied to Livestock Production, Guelph, Canada, August 1994, 455-461.

Rege, J.E.O. & Bester, J. 1998. Livestock resources and sustainable development in Africa. Proceedings of the 6th World Congress on Genetics Applied to Livestock Production, Armidale, Australia, 28, 19-26.

Seibold, R. 1996. Das Arche-Hof Projekt der Gesellschaft zur Erhaltung alter und gefährdeter Haustierrassen, e.V. In F. Begemann, C. Ehling & R. Falge (Eds.) Schriften zu genetischen Ressourcen, IGR; Bonn, 5, 78-87.

Simon, D.L. 1984. Conservation of animal genetic resources – a review. *Livestock Production Science*, 11, 23 – 36.

Simon, D.L. 1990. The global animal genetic data bank. In G. Wiener (Ed.) *Animal Genetic Resources; A global programme for sustainable development*. Proceedings of an FAO Expert Consultation, Rome, Sept. 1989, FAO Animal Prod. and Health Paper 80, 153-166.

Simon, D.L. 1993. Kryokonservierung zur Unterstützung der Lebenderhaltung gefährdeter Rinderrassen. *Züchtungskunde*, 65, 91-101.

Simon, D.L. 1995. Factors for defining the status of endangeredness of a breed. In R.D. Crawford, E.E. Lister & R.T. Buckley (Eds.) *Proceedings of the Third Global Conference on Conservation of Domestic Animal Genetic Resources*, Kingston, Canada, August, 1994, 384-390.

Simon, D.L. 1999. Genetic resources and conservation. In R. Fries & A. Rudinsky (Eds.) *The Genetics of Cattle*. CAB International, Wallingford, Oxon, UK, 475-495 (in press).

Simon, D.L. & Buchenauer, D. 1993. Genetic diversity of European livestock breeds. *EAAP Publication No. 66*, Wageningen Pers, pp. 580.

Smith, C. 1984. Estimated costs of conservation of farm animals. In *Animal Genetic Resources, Conservation by Management, Data Banks and Training*, Animal Prod. and Health Paper, FAO, Rome, 44, 21-30.

Weitzman, M.L. 1993. What to conserve? An application of diversity theory to crane conservation. *The Quarterly Journal of Economics*, 157-183.

Wright, S. 1922. Coefficients of inbreeding and relationships. *Amer. Nat.*, 56, 330-338.

AIRE 2066 CEC Concerted Action. 1998. 'Analysis of genetic diversity to preserve future breeding options', Final Report,.

British Society of Animal Production. 1997. Meeting & Workshop Publication: The potential role of rare livestock breeds in UK farming systems. Appleby, United Kingdom.

Deutsche Gesellschaft für Züchtungskunde. 1979. Ausschuss genetisch-statistische Methoden in der Tierzucht, Vors. F. Fewson, Stellungnahme zur Bildung von Genreserven in der Tierzucht. *Züchtungskunde*, 51, 329-331.

European Commission, Council Regulation (EEC). 1992. No 2078/92 of June 30 1992, Official Journal of the European Communities No L 215, 85-90; with Comité des structures agricoles et du développement rural (STAR)- Working Document VI/5104/92 and VI/3879/94.

European Commission, Council Regulation (EC). 1994. No 1467/94 of June 20 1994, Official Journal of the European Communities No L 159, 1-10.

Earth Summit, Agenda 21. 1992. The United Nations Programme of Action from Rio, United Nations Conference on Environment and Development (UNCED), 3-14 June 1992, Rio de Janeiro, Brazil.

FAO -World Watch List for Domestic Animal Diversity. 1995. B.D. Scherf (Ed.); FAO-Rome, Italy, pp. 769.

Inbreeding and loss of founder alleles in four variations of a conservation programme using circular mating, for Danish Shorthorn Cattle

M. Trinderup¹, J.N. Jørgensen¹ & M. Hansen²

¹Department of Animal Science and Animal Health, the Royal Veterinary and Agricultural University, 13 Bülowssvej, 1870 Frederiksberg C, Denmark

²The Danish Agricultural Advisory Centre, The National Committee of Danish Cattle Husbandry, 15 Udkjærsvøj, Skejby, 8200 Århus N, Denmark

Summary

The beef cattle breed Danish Shorthorn is used as a model for simulation of four variations of the circular mating scheme running for 50 years. Schemes 1 and 2 used a fixed exchange of bulls between female groups while schemes 3 and 4 used a random exchange of bulls between female groups. The number of bulls used in schemes 1 and 4 was 16 while the number of bulls used in schemes 2 and 3 was 8. Inbreeding levels were computed and gene dropping was applied to estimate the risk of founder allele loss. In the population of year 50 the inbreeding levels for the four mating schemes are not statistically different. The male founder alleles contribute strongly to the population of year 50. The female founder alleles are in greater risk of being lost than the male founder alleles.

Résumé

La race bovine à viande Danish Shorthorn est utilisée comme modèle pour la simulation de quatre variations du schéma circulaire d'accouplement au long de 50 ans. Les schémas 1 et 2 utilisent un échange fixe de taureaux dans des groupes de femelles. Le nombre de taureaux utilisés dans les schémas 1 et 4 a été de 16, tandis que le nombre de taureaux utilisés dans les schémas 2 et 3 était de 8. On a établi les niveaux de consanguinité et appliqué le comptage des gènes pour estimer le risque de perte d'allèles de base.

Dans la population de la 50ème année les niveaux de consanguinité pour les quatre schémas d'accouplement ne sont pas statistiquement différents. Les allèles mâles de base contribuent fortement à la population de la 50ème année. Les allèles femelles de base se trouvent en plus grand péril de perte que ceux des mâles.

Key words: Gene dropping, Diversity, Beef cattle.

Introduction

Several methods for the conservation of animal genetic diversity have been developed. Cryogenic storage is a conservation method free from human interference which might bring about genetic change (Bodó, 1990). Though storage of frozen semen and embryos has been known and used for years, the most important and practical method of conserving genetic diversity now and in near future is to manage genetic variability in some small living populations (Smith, 1984; Wang *et al.*, 1994). For most livestock breeds the conservation of a living population involves private breeders. Therefore it is necessary to have the support of the breeders if a conservation programme is to be a success (Rognoni & Finzi, 1984). This calls for breeding plans that are fairly easy for the breeders to implement in the population, e.g. breeding plans that fit the demographic structure of the population.

Many studies have been carried out on the topic of conserving small populations with a minimum of inbreeding (Kimura & Crow, 1963; Smith, 1984; Chevalet & Rochambeau, 1985; Bodó, 1990; Wang *et al.*, 1994). The majority of the authors suggested dividing the population into breeding groups of females and a rotation of males among the groups in a circular mating scheme. Chevalet & Rochambeau (1985) compared three turnover rates of the males in a circular mating scheme implemented in a small dairy cattle breed by simulations. In the first and second systems the males were used for two years in each group of females; totally the males were used for 16 years. In the first system the bulls were replaced by a son. In the second system the programme was initiated by selection of eight bulls and one male offspring of each bull as a replacement male. When a bull was culled the replacement male became the new bull and one of the culled bulls sons was selected as a new replacement male. Thus, the second system prolonged the generation interval of the males as compared to the first. In the third system the males were only used for two years and only in one group of females. Chevalet & Rochambeau (1985) found that prolonging the generation interval of the males delayed the inbreeding, but the rapid turnover (system three) gave the lowest inbreeding level.

Not only is the inbreeding of concern in conservation genetics, attention has also been given to the founder representation in the population under study and to preventing loss of founder alleles in the future (Chevalet & Rochambeau, 1985; MacCluer *et al.*, 1986; Lacy, 1989; Boichard *et al.*, 1997). With less breeding males than females, as is the case in cattle populations, there is a tendency of loss of female founder alleles, but a rapid turnover of the males enables the population to keep alleles from the female founders (Chevalet & Rochambeau, 1985).

In the circular mating schemes outlined by other authors (Kimura & Crow, 1963; Chevalet & Rochambeau, 1985) a fixed scheme is used for the exchange of males

between the groups of females. In practice breeders might find it difficult to follow such a strict plan for the use of the males.

In this paper the objectives are:

1. to test the hypothesis that a random exchange of males between the groups of females increases the inbreeding level compared to an exchange of males that follows a fixed scheme;
2. to test the hypothesis that increasing the number of males will result in a decrease in female founder alleles.

Materials and Methods

Data of the beef cattle breed Danish Shorthorn (figure 3), provided by The National Committee of Danish Cattle Husbandry, is used as a model for simulations of four variations of the circular mating scheme. The first generation in the simulation of the mating schemes is founded by the Danish Shorthorn population of 1997 which consisted of 96 females and 8 males (Trinderup *et al.*, 1998). This year is referred to as year zero in the simulations.

Initially the females are divided into eight breeding groups, each of 12, referring to the original herds found in the data material. The number of females is fixed through the time period simulated to exclude the effect of fluctuations in population size. Based on the age distribution of the females found in the data the replacement rates over age classes were computed (table 1). These replacement rates were used as culling probabilities for the cows in the simulations. As can be seen in table 1, the maximum age of females was set to 12 years in the simulations. The females were at least two years before they were mated for the first time. Once a cow was mated it was assumed that she gave birth to one calf of random sex each year until she was culled. To prevent loss of female founder alleles and to reduce variation in female family size the first choice of a replacement heifer was a daughter of the culled cow. A cow's offspring were kept in the population until she was replaced in order to maximize the possibility of having a daughter for

replacement. Otherwise a heifer from the same breeding group or herd was randomly selected.

The difference between the four mating schemes was the way the breeding bulls were used in the population as illustrated in figure 1 and outlined in the following.

Scheme 1

Each of the eight bulls were assigned to one of the eight herds in year 0. Each bull was mated to all the females of his herd in year 0 and one son was randomly selected. Each of the eight selected young bulls were sent from the j 'th herd to the $j+1$ 'th herd, except for the 8'th herd where the young bull was sent to the first herd. After year 0 the 'old' bulls were mated to the cows and the 'young' bulls were mated to the heifers. The following year, after a new rotation of young bulls, the 'old' bulls were culled and replaced by the 'young' bulls, who now were mated to the cows, and the new 'young' bulls were mated to the heifers. Thus, each breeding bull was used in two years. The rotation of bulls follows a fixed system which ensures that the male descendant of a certain bull returns to the herd with the female descendants after eight rotations. This scheme doubles the number of males from the original eight to sixteen.

Scheme 2

This scheme was similar to scheme one, except that here was only one bull per herd. This bull was mated to both heifers and cows. The bulls were only used for one year before they were replaced by one of their sons and the number of bulls was kept at the original eight.

Scheme 3

As scheme 2, but here the bulls were randomly assigned to a herd. This means that a bull was allowed to breed in the herd that he was born in.

Scheme 4

As scheme 1, except that the 'young' bulls were randomly selected among all the bull calves born and all living, non-breeding bulls up to 10 years of age. As in scheme 1 the breeding bulls were culled after two years. The selected bulls were not assigned to any herd, but could be mated to all cows and heifers in the population just like bulls from artificial insemination centres (A.I.-bulls).

PASCAL programmes were written to simulate the circular mating schemes. Each simulation of a mating scheme had a time span of 50 years or approximately 12 generations and were repeated 200 times.

The inbreeding coefficient for each animal was computed as proposed by Quaas (1976). The base animals for the inbreeding coefficient computation were the animals with unknown parents found in the data of Danish Shorthorn (Trinderup *et al.*, 1998). The founders of the circular mating schemes, the Danish Shorthorn population of 1997, were therefore not unrelated animals as defined by other authors (e.g. Lacy, 1989), but the animals entering the circular mating schemes (Foose, 1986; Falconer & Mackay, 1996). The founders of the circular mating schemes were the reference generation of a gene dropping (MacCluer *et al.*, 1986) which was conducted in order to estimate the founder representation after 50 years of breeding.

Results

Figure 2 shows the change in mean inbreeding coefficient over time in the four circular mating schemes. The inbreeding coefficients were highest in the schemes with random exchange of bulls in the first seven years of the simulations. But for animals born after year eight the inbreeding coefficients were almost the same in all four mating schemes.

Table 2 shows the inbreeding levels and trends with the standard deviation for the animals in the population of year 50. To reduce the confounding of the year of birth and the number of ancestral generations the

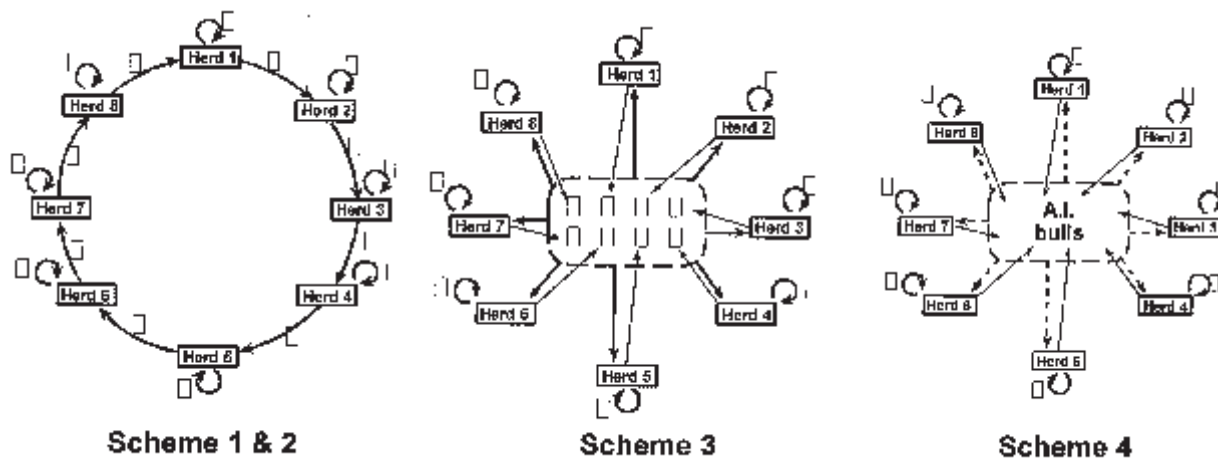


Figure 1. Illustration of the differences between the four mating schemes.

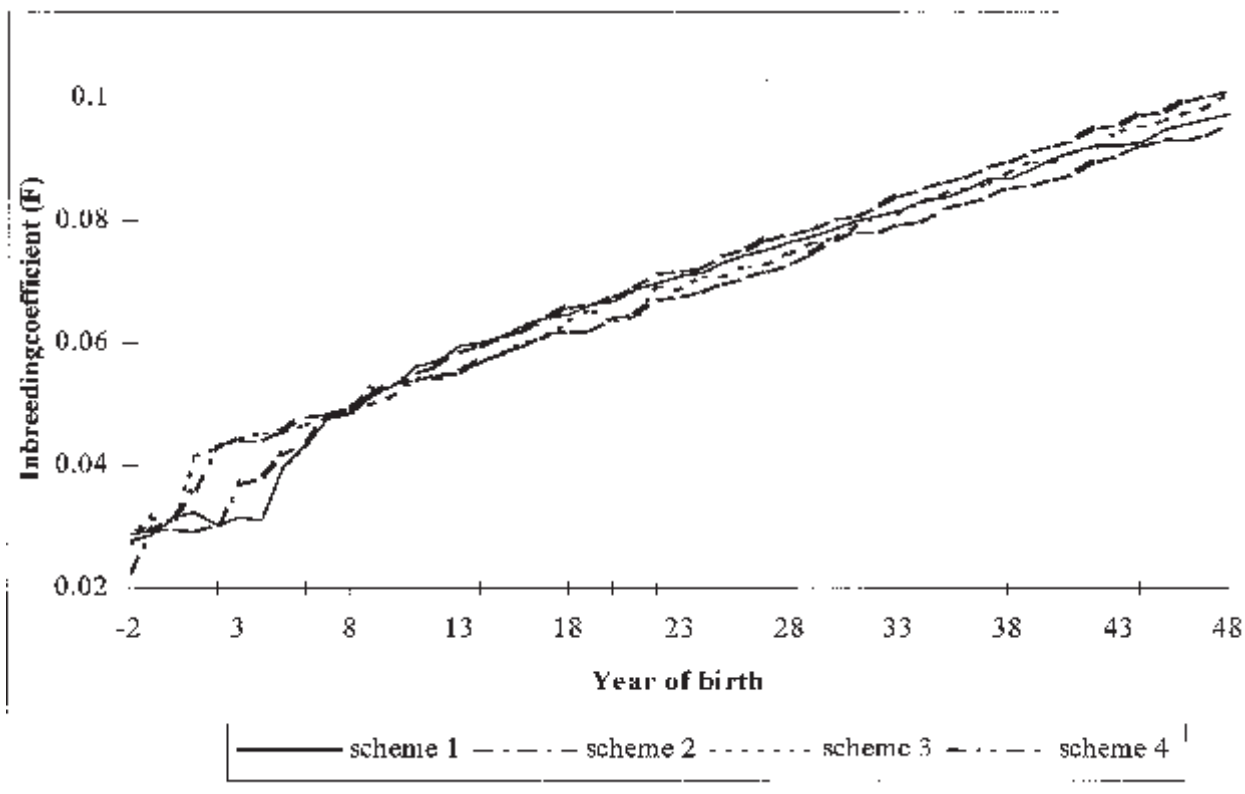


Figure 2. Average inbreeding levels per year of birth.

inbreeding trends should be tabulated for animals with more than seven ancestral generations (Te Braake *et al.*, 1994). Thus, the inbreeding trends were tabulated from the two last generations in the simulations. The schemes with random exchange of bulls had the largest inbreeding trend and thus the smallest effective population size. Though scheme 4 (16 bulls) had the second highest inbreeding trend, this scheme resulted in the lowest mean inbreeding trend in the population of year 50. Scheme 1 (16 bulls) had the lowest inbreeding trend and the second lowest mean inbreeding levels at the end of the simulations. However, the standard deviations indicate that there is no statistical difference between the four mating schemes in rate of inbreeding and mean inbreeding level in the population of year 50.

Table 3 shows the result of the gene dropping. The criteria used here were the average contribution of alleles from each male and female founder, the total male and female founder allele contribution and the minimum and maximum percentage of the 200 replicates in which the alleles of a male or female founder was lost. The mating schemes using 16 bulls (i.e. schemes 1 and 2) resulted in the largest individual and total contribution of male founder alleles and the smallest minimum values of percent replicates with male founder allele loss. In all four mating schemes the maximum and minimum percent replicates with male founder allele loss were lower than the percent replicates with female allele loss. The percentages of replicates where the alleles of a female founder were lost were very much the same for all four mating schemes, due to the fact that the replacement strategy was identical in all four mating schemes.

Discussion

It was expected that if there were any differences in inbreeding trends among the four mating schemes they would be small, because the same replacement strategy for the females were used in all the mating schemes, the difference in population size was very

small and there were only small variations in the generation length for the males. In the simulations there was no significant difference in inbreeding trend or in inbreeding level after 50 years. In the first few years there seems to be a difference between the schemes using random exchange of males and the schemes using a fixed plan for exchange of males. This confirms the hypothesis that random use of males results in an instantly higher inbreeding level, but the difference is not permanent. When the first rotation of males is completed, the inbreeding increases in the schemes with fixed exchange of males to the level of the schemes with random exchange of males after which the rate of inbreeding is fairly similar in all four mating schemes.

Concerning the inbreeding level and trend it can be concluded that it is not important that the breeders follow a strict plan for the exchange of males as long as they follow the guidelines used in these simulations: minimising the variation of family size within sires and dams and avoid fluctuations of population size. The size of inbreeding trend found in these simulations of about 0.5% is acceptable in any breeding plan, because with such a low increase in inbreeding it is possible to select the animals showing the least inbreeding depression (Adalsteinsson *et al.*, 1994).

The distribution of male and female alleles in the founder population (year 0) is 7.7% male founder (8 bulls among 104 animals) alleles versus 92.3% female founder alleles (96 cows among 104 animals). The total contribution to the population of year 50 found in schemes 2 and 3, using eight bulls, was 20% male founder alleles and 80% female founder alleles (table 3), as found by Chevalet & Rochambeau (1985) in a simulation study running over 40 years. The change in the contribution of male and female founder alleles over years is due to the more intense use of bulls. Hence the male founder alleles are over-contributing to the population of year 50, whereas the schemes fail to some extent to keep the female founder alleles segregating.

Table 1. Replacement probabilities for female age classes.

| Age, years | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 | 11 | 12 |
|-------------|-------|-------|-------|-------|-------|-------|-------|-------|-------|-----|
| Probability | 0.178 | 0.159 | 0.209 | 0.137 | 0.267 | 0.189 | 0.267 | 0.318 | 0.333 | 1.0 |

Table 2. Mean inbreeding and inbreeding trend with standard deviation in the population of year 50.

| Mating scheme | Inbreeding trend ^{a)} | | N _e ^{b)} | Mean inbreeding ^{c)} | |
|---------------|--------------------------------|------|------------------------------|-------------------------------|------|
| | % | S.D. | | % | S.D. |
| 1 | 0.47 | 0.22 | 106 | 9.48 | 0.29 |
| 2 | 0.52 | 0.19 | 97 | 9.82 | 0.37 |
| 3 | 0.62 | 0.37 | 81 | 9.71 | 0.53 |
| 4 | 0.55 | 0.32 | 91 | 9.28 | 0.46 |

a) Computed from the last two generations and averaged over the 200 replicates.

b) Calculated by the equation: $N_e = 1 / (2 * \Delta F)$.

c) Mean inbreeding coefficient of the last generation averaged over the 200 replicates.

Table 3. Founder contribution to the population of year 50 and risk of allele loss.

| Mating scheme | Individual contribution | | Total contribution | | Replicates with founder allele loss, % | | | |
|---------------|-------------------------|---------|---------------------|---------|--|------|---------|------|
| | of the two sexes, % | | of the two sexes, % | | males | | females | |
| | males | females | males | females | min. | max. | min. | max. |
| 1 | 3.07 | 0.79 | 24.52 | 75.48 | 4.5 | 12.0 | 30.5 | 48.0 |
| 2 | 2.44 | 0.84 | 19.54 | 80.46 | 10.5 | 16.5 | 30.0 | 48.0 |
| 3 | 2.54 | 0.83 | 20.31 | 79.69 | 11.5 | 20.0 | 33.0 | 48.0 |
| 4 | 3.04 | 0.79 | 24.34 | 75.66 | 4.0 | 22.0 | 32.0 | 47.5 |

Our simulations indicate that in a conservation programme using a circular mating scheme it can be up to the individual breeders to manage the exchange of bulls among the groups of females. The over contribution of founder sire alleles found in this investigation indicates that the initiating sires of a conservation programme should be selected carefully. A method to ensure that

the founder sires of a conservation programme are of the type of interest is the gene dropping method as shown by Trinderup *et al.* (1998). The observed small differences between the four mating schemes indicate that other factors, such as economy, should be considered in the choice of conservation programme.



Figure 3. Danish Shorthorn cattle.

References

- Adalsteinsson S., Ó.R. Dýrmondsson, S. Bjarnadóttir & E. Eythórsdóttir.** 1994. The Icelandic Goat - a case study of a small old population. Proceedings of the 5th World Congress on Genetics Applied to Livestock Production. Guelph, Canada 4: 517-519.
- Bodó I.** 1990. Methods and experiences with in situ preservation of farm animals. In: FAO Animal Production and Health Paper. Ed. by Gerald Wiener, FAO Rome. 80: 85-102.
- Boichard D., L. Maignel & E. Verrier.** 1997. The value of using probabilities of gene origin to measure genetic variability in a population. Genetic Selection Evolution. 29: 5-23.
- Chevalet C. & H. De Rochambeau.** 1985. Predicting the Genetic Drift in small Populations. Livestock Production Science. 13: 207-218.
- Falconer D.S. & T.F.C. Mackay.** 1996. Introduction to quantitative genetics. Fourth edition. Longman Scientific & Technical, pp. 464.
- Foose T.J.** 1986. Genetics and demography of small populations. In: The Przewalski Horse and restoration to its natural habitat in Mongolia. FAO Animal Production and Health Paper, FAO, Rome, 61:9 3-103.
- Kimura M. & J.F. Crow.** 1963. On the Maximum Avoidance of Inbreeding. Genetical Research, 4: 399-415.

Lacy R.C. 1989. Analysis of founder representation in pedigrees: Founder equivalents and founder genome equivalents. *Zoo Biology*, 8: 111-123.

MacCluer J.W., J.L. VandeBerg, B. Read & O.A. Ryder. 1986. Pedigree analysis by computer simulation. *Zoo Biology*, 5: 147-160.

Quaas R.L. 1976. Computing the diagonal elements and inverse of a large numerator relationship matrix. *Biometrics*, 32: 949-953.

Rognoni G. & A. Finzi. 1984. Aspects of conservation of animal genetic resources - Italian experiences. *Livestock Production Science*, 11: 61-64.

Smith C. 1984. Genetic aspects of conservation in farm livestock. *Livestock Production Science*, 11: 37-48.

Te Braake M.F.H., A.F. Groen & A.W. Van der Lugt. 1994. Trends in inbreeding in Dutch Black and White dairy cattle. *Journal of Animal Breeding and Genetics*, 111: 356-366.

Wang J.L., J.C. Xu, J.Z. Song & W.S. Tang. 1994. Study on mating systems for controlling inbreeding levels in domestic animal populations with overlapping generations. *Proceedings of the 5th World Congress on Genetics Applied to Livestock Production*. Guelph, 21: 536-539.

Advances in the Brazilian animal genetic resources conservation programme

A. da S. Mariante¹, M. do S. M. Albuquerque¹, A. A. do Egito¹ & C. McManus²

¹National Centre for Genetic Resources, EMBRAPA-CENARGEN, Sain Parque Rural, P.O.Box 02372, 70849-970 **Brasilia**, Brazil

²Universidade de Brasília, **Brasília**, DF, Brazil

Summary

Brazil has various species of domestic animals which developed from breeds brought by the Portuguese settlers soon after the discovery. Over the last five centuries, these breeds have been submitted to natural selection in particular environments and therefore today, they present characteristics adapted to the specific environmental conditions. From the beginning of this century, some exotic breeds, selected in temperate regions, have begun to be imported. Although more productive, these breeds lack adaptation traits, such as resistance to disease and parasites found in breeds considered to be "native", but even so, little by little, they have substituted the native breeds to such an extent that the latter are, today, in danger of extinction. To avoid the loss of this important genetic material, Brazil created an Animal Genetics Resource Conservation Programme, coordinated by the National Research Centre for Genetic Resources and Biotechnology (Cenargen) of the Brazilian Agricultural Research Corporation (EMPRAPA). The conservation has been carried out by various Research Centres of EMPRAPA, Universities, State Research Corporations, as well as by private farmers, with a single coordinator at national level, Cenargen. The conservation is being carried out through Conservation Nuclei, situated in the habitats where the animals have been subjected to natural selection (*in situ*), and by the storage of semen and embryos (*ex situ*). The recently created Animal Genetics Laboratory of Cenargen allowed genetic characterisation studies on

cattle and horse breeds to begin, and, in the near future, work with asses, buffalo and sheep will be conducted.. From the results of this research it will be possible to compare the native breeds and estimate genetic distances between them. The harmonisation of chosen micro-satellites with those which have been used in other Latin America and Iberian Peninsula countries will be extremely useful for comparative studies and will allow future exchange of germplasm between countries.

Resumen

Los colonizadores portugueses, cuando vinieron al Brasil, trajeron consigo, animales domésticos. Estos se multiplicaron, y han sido sometidos a un amplio proceso de selección natural, adquiriendo características adaptativas y/o de producción para las diversas situaciones ecológicas del país y se han transformado en lo que conocemos como razas "locales" o "criollas". Cerca de tres siglos después del descubrimiento, muchos criadores comenzaron a importar animales de razas nuevas, buscando mayor productividad. El establecimiento de políticas que promovieron la dilución de germoplasma autóctono a través de programas extensivos de cruzamiento con esas razas importadas resultó en rápida substitución de las razas locales. Aunque éstas presenten productividad más baja que las exóticas, están extremadamente bien adaptadas a las condiciones ambientales, a los que estuvieron sometidos bajo selección natural. En este trabajo discutimos la situación actual de la

conservación de las razas todavía existentes en Brasil. La investigación sobre evaluación, conservación y utilización de reservas genéticas de razas con características únicas, debe proveer la fundación para la utilización efectiva del germoplasma a nivel global. Complementariamente al uso tradicional de recursos genéticos animales, están los avances significativos en genética animal alcanzados en las dos últimas décadas, usando técnicas de biología molecular, tales como el mapeamiento y la identificación de genes. Toda esta preocupación con la diversidad de los animales domésticos ha llamado la atención de los investigadores a la rápida desaparición de las razas locales, y, consecuentemente con la dilución del germoplasma autóctono a través del uso extensivo de programas de hibridación. La conservación de recursos genéticos animales en Brasil viene realizada en diversos Centros de Investigación de EMPRAPA (Empresa Brasileira de Pesquisa Agropecuária), Universidades, Empresas de Investigación de distintos Estados, bien como por productores privados, involucrando todo el país, bajo la coordinación del Cenargen (Centro Nacional de Investigación de Recursos Genéticos y Biotecnología). La conservación se realiza por medio de núcleos de conservación, mantenidos en el habitat donde los animales están adaptados (*in situ*) y por el almacenamiento de semen y embriones (*ex situ*), incluyendo 7 especies animales: bovinos, bubalinos, cerdos, ovejas, cabras, caballos y asnos. Una importante tarea del Programa es aumentar la conscientización sobre la importancia de la conservación de recursos genéticos animales.

Introduction

Brazil has various species of domestic animals (Figure 1 to 4), which developed from breeds brought by the Portuguese settlers soon after the discovery. For almost five centuries, these breeds have been submitted to natural selection in determined environments so that, today, they present characteristics adapted to the specific environmental conditions.

From the beginning of this century, some exotic breeds, selected in temperate regions, have begun to be imported. Although more productive, these breeds lack adaptation traits, such as resistance to disease and parasites found in breeds considered to be "native", but even so, little by little, they have substituted the native breeds, to such an extent that the latter are, today, in danger of extinction. To avoid the loss of this important genetic material, Brazil created an Animal Genetic Resources Conservation Programme, coordinated by the National Research Centre for Genetic Resources and Biotechnology (Cenargen) of the Brazilian Agricultural Research Corporation (EMPRAPA). The conservation has been carried out by various Research Centres of EMPRAPA, Universities, State Research Companies, as well as by private farmers, with a single coordinator at national level, Cenargen. This programme includes the following stages:

1. identification of populations in an advanced stage of genetic dilution;
2. phenotypic and genetic characterisation of germplasm; and
3. evaluation of productive potential. The conservation is being carried out by Conservation Nuclei, situated in the habitats where the animals have been subjected to natural selection (*in situ*), and by the storage of semen and embryos (*ex situ*). An important challenge for this programme is to increase awareness among the different segments of society for the importance of the conservation of animal genetic resources.

Objectives of The Brazilian Conservation Programme

The objectives of the Brazilian Animal Genetic Resource Conservation Programme are:

1. Identifying and characterising phenotypically conservation nuclei, establishing centres of origin and assessing genetic diversity and variability for the groups of animals in danger of extinction;



Figure 1. Lavradeiro horse (Northern region).

2. Monitoring existing animal conservation nuclei;
3. Starting new conservation nuclei of breeds, which are identified as being in danger of extinction;
4. Conserving *ex situ* genetic material by cryopreservation of semen and embryos;
5. Genetically characterising the breeds involved in the Programme; and
6. Increasing the awareness of the diverse segments of society about the importance of the conservation of animal genetic resources.

Brazilian Animal Genetic Resources

Native breeds

The first cattle arrived in the American continent with the settlers in 1493, when some animals were left on the *Hispaniola* Island, which today is the Dominican Republic and

Haiti. Cattle arrived in Brazil for the first time in 1532, introduced by the Portuguese. New introductions were made from the Archipelago of Cape Verde, where the Portuguese had been since 1460.

With its continental dimensions, Brazil has a huge variety of ecosystems where the different species of domestic animals brought by the first settlers began to establish themselves. Through centuries of natural selection, these animals attained special adaptation features specific to the ecological niche where they developed. Little by little, these animals dispersed over the whole Brazilian territory, and adapted to the very diverse environmental conditions with special characteristics such as in the Mato Grosso Pantanal, the North Eastern Agreste, the southern Brazilian Plateau and the Pampas of Rio Grande do Sul.

Due to the increasing demand for food of animal origin, farmers from many developing countries followed a course which, inevitably, led to the dilution of the "local" germplasm

by the use of intensive crossbreeding with animals of exotic breeds. Many of these programmes failed, since the introduced animals had lower productive indices than the "local" breeds. This meant that a considerable number of farmers, in establishing their production systems, started to give a deserved importance to the "local" breeds, because of their adaptation to the environment which is usually hostile especially in the tropical region.

Zebu breeds

At the end of the last century, the introduction of animals, which until then had been considered extremely exotic – the zebu, began. Today these breeds are responsible for almost the total meat production in the country, as purebred or crossbred animals, from the latitude of São Paulo State northwards.

The zebu was first introduced into Brazil at the end of the 19th century. These animals, which originated in India, were originally crossed with the local breeds. Little by little the local breeds were absorbed. As a result, Brazil has today the largest zebu population in the world, while in their country of origin, where they are considered sacred, they are raised freely and protected by the community until they die a natural death.

Before the first importation of zebu cattle to the Southeast region of Brazil, the Southern region, of temperate climate, had already imported British cattle breeds which were highly productive. The latter did not have the same adaptation problems as animals of the same breeds in the tropical regions of the country. As in central Brazil, the British breeds introduced in the south of the country were used in crossbreeding schemes, leading again to a drastic reduction in the effective population size of the local breeds.

The expansion of the zebu breeds in Brazil is an undisputed reality. Today about 80% of the Brazilian cattle population is made up of zebu cattle or their crosses with Creole and European cattle. Although, up to this time, the conservation Programme is only involved

with the native breeds, as they are threatened with extinction, there is already a demand from the zebu breeders that these breeds also be included. This is due to the almost total domination of the Nelore breed in Central Brazil. Of the six principal zebu breeds that exist in Brazil, approximately 85% of the total number of animals registered are Nelore.

Another aspect, which should be highlighted, is the fact that the use of new technologies and more efficient methods in genetic improvement programmes have led to profound changes in the procedures used by the breeders in the selection and reproductive management of their herds. Since 1984, when the first Bull Summary was published (Mariano *et al.*, 1984), the breeders began to have more precise information about the genetic merit of each individual selection candidate. This, together with techniques which allowed the increase in reproductive capacity of the genotypes of interest (artificial insemination, embryo transfer and in vitro fertilization), have made the decision taking process more objective, especially when referring to the choice of animals for reproduction and mating schemes.

From the beginning of the publication of the Bull Summaries a large number of breeders started to put greater emphasis on the productive traits (taken from the Classification Table of the Summary), leaving behind the qualitative traits, such as breed characteristics and type. Bulls which up to then may have been left aside because of one or another external trait, which did not please the more conservative breeders, sired offspring with exceptional weight gains, proving that they imprinted production traits which were more important at slaughter. These bulls then became highly demanded by the Artificial Insemination Centres and their semen was sold at very high prices.

This situation resulted in the use of a small number of bulls, especially those classified as Elite, which, certainly led to the reduction in genetic variability. Ironically, this bull classification due to merit, together with more modern practices in animal reproduction



Figure 2. Pantaneiro horse (Northern region).

already mentioned, are partially responsible for the reduction in genetic variation (Mariane, 1990).

These preoccupations of breeders led to the demand for the creation of a Germplasm Bank where semen and oocytes of zebu breeds would be stored, and would be monitored by EMPRAPA-Cenargen, although it would be situated at the headquarters of the Brazilian Association of Zebu Breeders in Uberaba, Minas Gerais State. The choice of animals which would be selected to donate semen and oocytes could be based on the Family Catalogue, which to date has been published for the Nellore breed (Magnabosco *et al.*, 1997). The catalogues for the other zebu breeds should be published in the near future.

Information is presented about the zebu breeds originally introduced and selected in Brazil (Gir, Guzerat, Nellore and Sindi), and those formed there (Indubrasil and Tabapuã). Some animals of the Kangaian breed were also imported, but today their effective population size is greatly reduced.

Gir

The first animals of the Gir breed were probably imported around 1906. The greater portion of the importation, which resulted in creation of nuclei which exist today, dates from 1920.

Guzerat

The main importations of the Guzerat breed were to the Curvelo region, Minas Gerais State. Later, farmers in the Uberaba district, of the same state, became interested in raising this breed. The breed continued to expand to the north of São Paulo State and from there to other regions of the country. In the region known as the Minas Triangle, Minas Gerais State, it was used in crossing programmes with the Gir and Nellore breeds, creating the Indubrasil breed.

Nellore

The Nellore breed belongs to the second Indian group (cattle with white or grey hair and short horns), by the classification of Joshi and Phillips, referred to by SANTIAGO (1987). The Brazilian Nellore population is originated from importations made in 1930 and notably those in 1960 and 1962. At the present time the Brazilian Nellore is tending to the type of the Indian Ongole. Of all the zebu cattle found in Brazil the Nellore stands out because of its production qualities and it is becoming more prized by the breeders. Although for some time it had been left aside because of its short ears, similar to those of European cattle, today it is in first place in terms of Genealogical Registration, as well as its overwhelming use as bulls in herds of other breeds. The animals are highly fertile, resistant to parasites and to tropical disorders, precocious and have extraordinary maternal ability. In Brazil, the Nellore is essentially a

meat producer, which has been subjected to highly intensive selection to obtain males for slaughter.

Sindi

The Sindi breed is very similar to the Gir from the west of India, to Sahiwal, from Punjab and the red cattle of Afghanistan. Due to the movement of nomad tribes in its home track, it had been crossed with Gir in some regions. The Sindi, which arrived in Brazil in about 1906 and 1930, had the Baixada Fluminense region, in the state of Rio de Janeiro, as their destination as well as the Novo Horizonte and Jardinópolis townships, in the São Paulo State. In general, these animals were small, good looking and useful in areas where there was a lack of forage, where it would be more difficult to maintain larger animals.

Table 1. Species and breeds included in research projects in the Brazilian Programme of Animal Genetic Resources Conservation - 1999.

| Species | Breed | Region of the country |
|-----------------|---|--------------------------|
| Cattle | Mocho Nacional (National Polled) | Southeast |
| | Pantaneiro | Centrewest (Pantanal) |
| | Curraleiro or Pé-duro | North east |
| Buffaloes | Criollo Lageano | South |
| | Baio | North |
| Asses | Carabao | North |
| | Jumento Nordestino (North-eastern Ass) or Jegue | Northeast |
| Horses | Jumento Brasileiro (Brazilian Ass) | Southeast |
| | Pantaneiro | Centrewest (Pantanal) |
| Goats | Lavradeiro | North |
| | Canindé, Gurguéia, Moxotó, Marota, Repartida | Northeast |
| Sheep | Criollo Lanado | Northeast |
| Diverse species | Animal Gene Bank (AGB) | Centrewest |

Table 2. Number of semen doses and embryos stored in the Animal Germplasm Bank (AGB) of Cenargen, in 1999.

| Species/Breed | No. of semen doses | No. of embryos |
|-------------------|--------------------|----------------|
| Cattle | | |
| Criollo Lageano | 6 159 | 9 |
| Mocho Nacional | 6 533 | 54 |
| Caracu | 3 950 | 47 |
| Curraleiro | 5 300 | 17 |
| Criollo Argentino | 288 | - |
| Pantaneiro | 4 277 | 20 |
| Junqueira | 2 143 | 4 |
| Patuá | 250 | - |
| Goats | | |
| Moxotó | 546 | - |
| Canindé | 109 | - |
| Sheep | | |
| Criolo Lanado | 500 | 56 |
| Horses | | |
| Pantaneiro | - | 1 |
| Asses | | |
| J. Nordeste | 150 | - |
| Total | 30 205 | 208 |

Indubrasil

The Indubrasil, according to Santiago (1984), was the first zebu breed formed by Brazilian breeders, based on cattle imported from India. Initially, its pioneers in the Minas Triangle gave it the name of Induberaba, but only in 1936 was it officially recognised as the Indubrasil, a name that it keeps today. Its origin, although basically founded from the Gir, Nellore and Guzerat breeds, is obscure as to the exact genetic contribution of each of the breeds or zebu types used.

Tabapuã

According to Santiago (1984), the first polled zebu, object of selection in Brazil, was the Tabapuã, named after the township where it was formed in São Paulo State.

Phenotypically, these cattle resemble the American Zebu (Brahman), which means that several Brahman cows, when dehorned, are confused with Tabapuã animals. Despite this, its racial composition is mainly Nellore with some Guzerat and Gir.

Formation of Conservation Nuclei

The Conservation Nuclei, organised in the form of research projects, can be found all over the country. The elaboration of research projects, usually based in research centres near the habitats where the animals were naturally selected over the last few centuries, was the solution adopted to try and rescue the small effective populations of the breeds in danger of extinction.

The articulation of Cenargen with these Conservation Nuclei (Germplasm Banks) is made by Germplasm Curator (based in Cenargen), in collaboration with the Curators of the Germplasm Banks (normally the research project leaders). In the present collaborative programme, there are 3 Germplasm Curators for animals at Cenargen: one for large species (cattle, buffaloes, horses and asses), one for small species (sheep, goats and pigs) and one for wildlife.

The Animal Germplasm Curators are researchers of Cenargen, attributed with giving advice to the Technical Head in relation to germplasm considered relevant to national agriculture and animal production. They work at national and international level, with subjects related to the enrichment of knowledge and conservation of product germplasm, being supported by technical areas for these activities. Among other functions, the animal Germplasm Curator has to promote, start and follow activities related to conservation, multiplication and/or regeneration of germplasm of products under their responsibility. The Curators of the Germplasm Banks have the responsibility to

Table 3. Recent Scientific Events which included Conservation of Animal Genetic Resources in their programmes.

| | Date | Event |
|----------------------------|----------------|---|
| Tampico, Mexico | November, 1998 | 4 th Ibero American Congress for Criollo Breeds |
| Kathmandu, Nepal | August 1998 | 4 th World Conference on Conservation – RBI |
| Colina, SP, Brazil | July 1998 | National Equine Forum |
| Soeul, Korea | July 1998 | 8 th World Congress of Animal Production |
| Valdivia, Chile | May 1998 | X Chilean Veterinary Medicine Congress |
| Armidale, Australia | January 1998 | 5 th World Congress on Genetics Applied An. Production |
| Niterói, RJ, Brazil | December 1997 | Workshop of the Rio de Janeiro Research Corporation |
| Amsterdam, Holland | November, 1997 | CEDLA Workshop |
| Brasília, DF, Brazil | July 1997 | Workshop on Conservation of Animal Genetic Recourses |
| B.Horizonte, MG, Brazil | May 1997 | Brazilian Animal Production Congress – Zootec'97 |
| Ottawa, Canada | February 1997 | Symposium organised by Ag-Canada |
| Guadeloupe, F.W.I. | December 1996 | Caribbean Meeting, INRA |
| Bogota, Colombia | November 1996 | 3 rd Ibero American Congress for Creole Breeds |
| C.Grande, MS, Brazil | October 1996 | XV Pan American Veterinary Congress (PANVET) |
| R. Preto, SP, Brazil | May 1996 | 33 rd Brazilian Animal Breeding Society Congress |
| Brasília, DF, Brazil | November 1995 | Workshop for curators of Conservation Nuclei |
| Orlando, USA | July 1995 | 87 th Annual Meeting American Society of An. Science |
| San José, Costa Rica | July 1995 | Towards an Inter American System for An. Genet. Res. |
| Circello, Italy | June 1995 | Expert Consultation of FAO |
| Guelph, Canada | August 1994 | 4 th World Congress on Genetics Applied An. Production |
| Kingston, Canada | August 1994 | 3 rd World Conference on Conservation – RBI |
| Buenos Aires, Argentina | July 1994 | Argentinean Association of Animal Production Congress |
| Rio de Janeiro, RJ, Brazil | July 1993 | Annual Meeting of the Brazilian Na. Production Society |
| Santiago, Chile | July 1993 | Conservation Symposium (ALPA Meeting) |
| Zafra, Spain | September 1992 | 1 st Ibero American Congress for Creole Breeds |
| Córdoba, Spain | September 1992 | World Meeting on Domestic Animal Breeds |
| Turrialba, Costa Rica | July 1992 | Conservation & Development of An. Genet. Res. in L.A. |
| Rio de Janeiro, RJ, Brazil | June 1992 | Science Forum (Earth Summit) |
| Nanjing, China | January 1992 | FAO International Course on Regional Gene Banks |

maintain the Conservation Nuclei as well as to multiply, regenerate and distribute the germplasm.

At the present time the Programme of Conservation of Animal Genetic Resources has 13 ongoing research projects (Table 1).

As already mentioned, the Conservation Nuclei are being kept where the animals were naturally selected over centuries. In a huge country such as Brazil, with several different climates, there is no reason to conserve animals in environments different from those to which they are adapted. Trips for the identification of new nuclei, in order to maintain endangered breeds considered to be native, means that new populations are continually being identified. Some are very similar to breeds officially included in the *in situ* conservation programme and others have totally distinct characters. In general, these populations, which have been identified in the last few years, have extremely reduced effective population sizes. The strategy used

has been to include them initially in the cryopreservation programme, so as to assure the storage of genetic material in the BGA in Cenargen. At the same time, their blood is collected to be used for genetic characterisation.

The results of genetic characterisation could reduce existing doubts about breed groupings. As an example, we can cite the case of the Criollo Lageano, Franqueiro and Junqueira cattle. All have huge horns and some breeders insist that they are the same breed, while others treat them as separate breeds. With genetic characterisation, strategies could be developed based on facts and not on suppositions.

Cryopreservation

To avoid the disappearance of local breeds, FAO began contacts in 1987 to install regional Animal Gene Banks (RAGBs) for developing



Figure 3. Criollo Lageano cow (Southern region).

Table 4. Number of animals of various breeds threatened with extinction on Cenargen's Experimental Farm in 1999.

| Species/Breed | Number of males | Number of females |
|-----------------|-----------------|-------------------|
| Cattle | | |
| Caracu | 1 | 1 |
| Criollo Lageano | 1 | 5 |
| Curraleiro | 7 | 10 |
| Junqueira | 2 | 3 |
| Mocho Nacional | 1 | 1 |
| Pantaneiro | 6 | 18 |
| Patuá | 1 | 2 |
| Puganu | 1 | 1 |
| Goats | | |
| Azul | 1 | 4 |
| Canindé | 2 | 3 |
| Marota | 3 | 6 |
| Moxotó | 3 | 8 |
| Nambi | 2 | 2 |
| Repartida | 1 | 4 |
| Horses | | |
| Campeiro | 2 | 6 |
| Pantaneiro | 4 | 5 |
| Asses | | |
| Casco de Mula | 1 | 1 |
| Inhata | 1 | 2 |
| Monteiro | 1 | 2 |
| Moura | 1 | 1 |
| Nilo | 1 | 2 |
| Piau | 1 | 1 |
| Rabo de Peixe | 1 | 2 |
| Total | 51 | 104 |

countries. At that time Cenargen/EMPRAPA had been chosen to host the Bank that would be responsible for the storage of semen and embryos of the breeds of domestic animals in danger of extinction in South America. A copy of the material would be stored in the Instituto Nacional de Tecnologia Agropecuaria, Argentina, for security reasons. Because of health legislation differences, regulating the entry and exit of genetic material between countries, none of these RAGBs was established. There is a need

to reach a common denominator, so that the future exchange of germplasm is facilitated, as the RAGBs have no commercial purpose, just storage for future use. RAGBs were also proposed for Asia and Africa. Seeing that, at that time, this goal would be difficult to achieve, FAO decided to stimulate these countries to create their own Animal Germplasm Banks or to strengthen those that already existed. At the same time it gave priority to the Domestic Animal Diversity - Information System - DAD-IS (FAO, 1998a). The erosion of domestic animal diversity is very clear as is evident from the number of breeds threatened with extinction world-wide (Hammond, 1993). With the help of many countries and organisations, particularly the European Association for Animal Production (EAAP), in 1991 FAO started a world-wide programme, with the collection of population data for seven main species of domestic animals. To date, two editions of the *World Watch List for Domestic Animal Diversity* have been published (FAO, 1993 and FAO, 1995).

In Brazil, the conservation of animal genetic resources was not jeopardised, since when FAO proposed the creation of the RAGBs, Brazil had already created its own Animal Germplasm Bank (AGB). The Brazilian AGB is kept at Cenargen, which is responsible for the storage of semen and embryos of various breeds of domestic animals threatened with extinction in Brazil. It has existed since 1983 and it is because of its existence that Cenargen was chosen to host the South American RAGB, as mentioned above. Table 2, shows the present state of the Brazilian AGB.

Conservation Nuclei must exist for semen, embryos and oocytes of the species/breeds in danger of extinction to be collected. For some breeds included in the Brazilian programme, there are less donor bulls than the number recommended by Smith (1984), which are 25 donors per breed with 100 doses/bull. Unfortunately, when the *ex situ* conservation began, some of the breeds did not have this number of males. Therefore, although more than the recommended 100 doses/bull has been collected, the number of bulls was reduced. In other cases, as the Argentine

Creole, as it is not a Brazilian Breed and its use in Brazil is not common (although some bulls were used on the Criollo Lageano of the Fazenda Canoas, Santa Catarina State) it was decided to stop the collection, and the semen already collected was kept by the AGB.

As the Conservation Nuclei are increased in size and number, or new herds are identified, the intention is to collect genetic material from the greatest possible number of animals, thereby increasing the genetic representation in the AGB.

Another National research programme of EMPRAPA, is that of Biotechnology Applied to Agriculture and Animal Production. This contemplates a project which intends to develop animal reproduction techniques that may be used in Conservation work. Among the techniques being studied are embryo bisection, *in vitro* fertilisation, cloning and the formation of transgenic animals.

Genetic Characterisation

For a long time the characterisation of different breeds of domestic animals in Brazil was based, almost exclusively, on phenotypic data (morphology and production), which sometimes is insufficient to distinguish between pure breeds and those heavily influenced by environmental factors (Panepucci, 1986). With reference to genetic characterisation, the few papers published on Brazilian native breeds include only cytogenetic studies, blood groups and protein polymorphisms.

In the bovine species, structural differences of the Y chromosome observed in different karyotypes of animals of European origin (*Bos taurus*) show that this chromosome is submetacentric, while Afro-Asian breeds (*B. indicus*) are acrocentric. While studying the Curraleiro breed, Brito (1995) verified the occurrence of polymorphism of this chromosome at a cytogenetic level. About 68% of the animals had an acrocentric Y chromosome, which indicates that, at some time during the formation of this breed, zebu type animals were introduced. This dimorphism of the Y chromosome had been

observed by Tambasco (1985) in four native bovine breeds (Caracu, Mocho Nacional, Curraleiro and Criollo Lageano), which indicates that both bovine subspecies were involved in the formation of native breeds. Although the Criollo Lageano has a lower acrocentric Y chromosome frequency, this could be due to the geographical location of this breed (found in the South of the country where the zebu influence is much smaller). Crossing the two species may have been favourable for these breeds, because it associated certain qualities of the taurines, such as precocity and productivity, with hardiness and disease resistance of the zebus.

Another study with native cattle breeds was carried out by Dr. Mário Poli (personal communication), of INTA, Argentina, consultant to Cenargen, in 1985. Poli cites that from phylogenetic trees, based on data obtained from the study of thirty blood factors and genetic frequencies, only three systems were determined (F, J and L), since the lack of genealogical registers made it impossible to estimate the frequency of more complete phenogroups.

In the Caracu breed, Bicalho (1985) carried out a population study of different breeding nuclei based on blood groups and protein polymorphisms. It was concluded that this breed originated from the Portuguese breeds Alentejana and Mertolenga, although there are no registers of the latter breed entering Brazil. The author concluded that the Caracu has low genetic variability and was subdivided into four genetically distinct subpopulations. From the analyses, it was suggested that animals should be exchanged between nuclei to prevent greater loss of genetic variability.

The genetic characterisation of several cattle breeds was the subject of a doctorate thesis using protein polymorphisms. The study included three native Brazilian breeds (Caracu, Pantaneiro and Mantiqueira), one native Argentinean (Argentine Creole) and two zebu breeds (Nellore and Gir). The genetic distances between pairs of populations were calculated using NEI method (1972 and 1978), from gene

frequencies of eight protein systems, using the DISPAN programme. The highest values were obtained between pairs where one of the breeds was of *Bos taurus* origin and the other of *Bos indicus*. The average distance between the native Brazilian and the Nellore and Gir breeds was 0.1083 and 0.0964, respectively (LARA, 1998). The small values obtained for genetic distances between the Caracu, Mantiqueira and Pantaneiro with the Argentine Creole, suggest a great similarity between them and support the hypothesis that these breeds were founded by Iberian cattle, probably sharing the same ancestry.

The small number of projects in this area, up to the present time, helped establish the Animal Genetics Laboratory of Cenargen as a priority. The laboratory started to function in February 1998 and will work primarily with DNA polymorphisms for the genetic characterisation of the populations of animals of the conservation programme. At the same time, a DNA Bank is being set up, which already has samples from several species of animals (60 heads of cattle, 18 horses, 113 asses, 154 goats and 89 buffaloes). The intention is that, in the short term, cells and tissues of several species, including wild animals, should be stored.

For the first stage, the intention is to verify the degree of diversity within the breeds/native populations, to gain a global idea of genetic distances using similarity indices. Following an FAO recommendation, 50 animals of each breed will be used. Since the sampling process is vital for the success of the proposed plan, an investigation will be made into the Conservation Nuclei and/or Breed Societies, when they exist, of the geographical distribution of each breed, so that the samples are representative of the whole population known to exist. The DNA may be collected from blood or semen.

For comparative studies, two methods for the evaluation of DNA polymorphisms will be used. Since there is not sufficient data in the literature to evaluate, from molecular information, what constitutes a distinct breed, it is necessary to compare different markers (Grattapaglia, personal communication).

A priori, the RAPD will be used as it is relatively cheap and fast, as well as relevant to the study (Egito, 1995). Micro-satellites will also be used, as they are well developed in cattle studies. This technique differentiates the heterozygotes which are co-dominant thus generating more information to discriminate the variability within populations, helping in the choice of individuals for conservation. In the Animal Genetic Laboratory, work has already begun on cattle and horses and will soon include asses, buffaloes, goats and sheep species.

The primers for the micro-satellites on cattle were selected from thirty identified by the International Society of Animal Genetics (ISAG), after a meeting in 1996. These primers are the same recommended by MoDAD-FAO (FAO, 1998b) and are being used for genetic characterisation of the Iberian breeds in a project being carried out by the University of Porto, Portugal, and financed by the European Community. This fact will make the comparison of the breeds studied in both projects easier and facilitate the exchange of germplasm between the countries.

Public Awareness of the Importance of Conservation of Animal Genetic Resources

Another objective of the programme is to make the various sectors of society aware of the importance of conservation of animal genetic resources. Presenting the programme on various opportunities has helped significantly. The creation of a *Farm Park*, as has occurred in several European countries will be another strategy to bring this theme to the public attention.

Presentation of the Brazilian Animal Genetic Resources Conservation Programme

The presentation of the Brazilian Programme has stimulated the creation of Conservation Nuclei in Brazil, as well as of other National

Animal Germplasm Banks in other Latin American countries. The creation of these other banks may allow, in the medium term, the collection of all the material in one place - the RAGB (with a duplicate in a second country) as proposed by FAO in 1987. In this way, the Programme has been presented in different events in Brazil and abroad. In Brazil the intention is to bring the philosophy of the work with animal conservation to society, as well as the actual state of development of the programme (often stimulating the creation of new Conservation Nuclei); abroad the intention is to

- 1) show the state of animal genetics resources conservation in Brazil, stimulating the creation of new National Banks and
- 2) present the actual conservation situation on the continent as a whole. It is hoped that the awareness level of society on the

importance of conservation of animal genetic resources is being raised at both the national and international level.

Since the conservation of animal genetic resources is a relatively new topic, it has only recently been included in the programmes of congresses and symposia. Until recently, the researchers who dared to breach this topic were labelled as philosophers. It was said that the so-called "native" breeds should be conserved in Zoological Gardens. Fortunately, this point of view is changing rapidly, and the most important congresses in the animal area are including sessions or symposia on this topic. At last, traits such as adaptation, hardiness and disease and parasite resistance, which many of these local breeds have, are being recognised and valued. Table 3 presents some of the places and sessions where the Brazilian Animal Genetic



Figure 4. Tatu pig (South-Eastern region).

Resources Conservation Programme has been presented over the last seven years, so that the increasing status of conservation can be evaluated.

Brazilian farm park

In 1993, the Animal Germplasm Bank (AGB) of Cenargen was recognised by the Brazilian Ministry of Agriculture, who donated an area of 900 ha for the installation of a *Farm Park*, which will be called Farm Park for the Animal Diversity in Brazil. The main objective of this park will be to bring together, in Brasilia, living examples of domestic animal breeds of different species (cattle, buffaloes, horses, asses, sheep, goats, pigs and poultry) which are in danger of extinction. This Farm Park will be open to the public and is already being built on Cenargen's Experimental Farm. Up to the present time, a large part of the animals that will be shown on the Farm Park have already been brought to Brasilia, and are being used for semen and embryo collection stored in the AGB. Table 4 shows the number of animals of the various species/breeds that can be found in Brasilia and which will be part of the Farm Park.

References

- Bicalho, H.M.S.** 1985. Grupos sangüíneos e polimorfismos de proteínas do sangue da raça Caracu (*Bos taurus taurus*). Análise populacional. Belo Horizonte: UFMG. Tese de Mestrado, pp. 114.
- Brito, C.M.C.** 1995. Polimorfismo do cromossomo Y no plantel de gado Pé-duro da EMPRAPA/PI. Campinas: UNICAMP, Tese de Doutorado, pp. 86.
- Egito, A.A. do** 1995. Uso de marcadores RAPD na identificação e caracterização de raças bovinas existentes no Brasil. Ribeirão Preto: USP, Dissertação de Mestrado, pp. 98.
- FAO.** 1998a. DAD-IS 2.0 User's Manual for National Coordinators: for the Management of Farm Animal Genetic Resources, Rome, Italy
- FAO.** 1998b. Secondary Guidelines for Development of National Farm Animal Genetic Resources Management Plans. Measurement of Domestic Animal Diversity (MoDAD): Original Working Group Report, pp. 55.
- FAO.** 1993. World Watch List for Domestic Animal Diversity. 1st Edition. Rome, Italy, 376 pp.
- FAO.** 1995. World Watch List for Domestic Animal Diversity. 2nd Edition. Rome, Italy, pp. 769.
- Hammond, K.** 1993. The status and Conservation of Animal Genetic Resources. In: Reunião Anual da Sociedade Brasileira de Zootecnia, 30, Rio de Janeiro, RJ, Brasil.
- Lara, M.A.C.** 1998. Variabilidade genética em bovinos e bubalinos através de polimorfismos protéicos: análise populacional e suas implicações no melhoramento. Ribeirão Preto: USP, Tese de Doutorado, pp. 215.
- Magnabosco, C. de U., Cordeiro, C.M.T., Trovo, J.B. de F., Mariante, A. da S., Lobo, R.B., Josahkian, L.A.** 1997. Catálogo de linhagens do germoplasma zebuino: raça Nelore. Brasília: EMPRAPA-Cenargen, Documentos 23, pp. 52.
- Mariante, A. da S.** 1990. O futuro de uma raça especializada de gado de corte: seleção versus variabilidade genética. In: Sociedade Brasileira de Zootecnia, Campina, SP. Bovinocultura de corte. Piracicaba, FEALQ, 109-112.

.....

Mariante, A. da S., Nobre, P.R.C., Silva, L.O.C. da, Rosa, A. do N., Figueiredo, G.R. de. 1984. Resultados do controle de desenvolvimento ponderal. I. Nelore. Campo Grande, EMPRAPA-CNPGC, Documentos, 18, pp. 76.

Nei, M. 1972. Genetic distance between populations. *The American Naturalist*, 106: 283-292.

Nei, M. 1978. Estimation of average heterozygosity and genetic distance from a small number of individuals. *Genetics*, 89: 583-590,

Panepucci, L. 1986. Importância dos marcadores genético-bioquímicos e sua aplicação ao melhoramento animal e à pesquisa em geral. EMPRAPA-UEPAE São Carlos, Documento 6. DDT. Brasília - D.F., pp. 26.

Pereira, J.C.C. 1996. Melhoramento genético aplicado à produção animal/ Jonas Carlos Pereira - Belo Horizonte: J.C.C. Pereira, pp. 416.

Santiago, A.A. 1984. Os cruzamentos na pecuária bovina. Campinas: Instituto Campineiro de Ensino Agrícola, pp. 549.

Santiago, A.A. 1987. A raça Nelore. In: Gado Nelore: 100 anos de seleção. São Paulo: Ed. dos Criadores, 153-154.

Smith, C. 1984. Genetic aspects of conservation in farm livestock. *Livestock Production Science*, vol. 11, n. 1, 23-36.

Tambasco, A.J., Trovo, J.B. de F. & Barbosa, P.F. 1985. Estudo cromossômico em racas naturalizadas de bovinos. In: Reuniao Anual da SBZ, Camboriu, SC. Anais, 154.

Indexes

AGRI 1 to AGRI 25

| | |
|------------------------------|-----|
| Contents | 126 |
| Authors' | 134 |
| Species' | 137 |
| Breeds/types | 138 |
| Main geographical entities | 142 |
| Selected secondary citations | 143 |

Contents AGRI 1-25¹**AGRI 1/1983**

- Follow-up action by FAO/UNEP to the recommendations of the technical consultation on animal genetic resources, Rome, June 1980: 1-3
- Sheep germplasm in Ethiopia. E.S.E. Galal: 5-12
- A model programme for the preservation and genetic improvement of the Sahiwal breed in India. R. Nagarcenkar: 13-16
- Cattle breed evaluation studies by the International Livestock Centre for Africa (ILCA). J.C.M. Trail: 17-20
- European Friesians - the Canadian and American invasion. E.P. Cunningham: 21-24
- Les parcs naturels de France et la conservation génétique animale. A. Audiot: 25-26
- Cataloguing of poultry stocks in North America. R.D. Crawford: 27-28
- Model progeny testing programme for draught in the Harijana breed. R. Nagarcenkar: 29-30

AGRI 2/1984

- Criollo cattle of the Americas. J.V. Wilkins: 1-19
- Scandinavian activities on the conservation of animal genetic resources. K. Majjala: 20-26
- The native sheep of Sri Lanka. L.A. Goonewardene, V. Ravindran, K. Nadarajah & R. Rajamahendran: 27-30

AGRI 3/1984

- The indigenous goats of Nigeria. L.O. Ngere, I.F. Adu & I.O. Okubanjo: 1-9
- Economic benefits of conserving animal genetic resources. C. Smith: 11-16
- Evaluation and conservation of native strains of chicken in the SABRAO region. B.L. Sheldon: 17-20
- The Criollo cattle project of Santa Cruz, Bolivia. J.V. Wilkins, F. Rojas & L. Martinez: 21-32
- Organizational activities for the conservation of farm animals in the United States. C. Brooke: 33-34
- Preservation of the Spanish horse in North America. D.P. Sponenberg: 35-38

AGRI 4/1985

- Ruminant livestock genetic resources in Cyprus. A. Constantinou: 1-9
- The Native pig of Sri Lanka. R. Rajamahendran, V. Ravindran, L.A. Goonewardene, P. Sahaayaruban & A.S.B. Rajaguru: 9-13
- The Barroso cattle of Guatemala. R. A. Melgar, R. Solano & J. De Alba: 15-17
- Hungarian activities on the conservation of domestic animal genetic resources. I. Bodó: 19-25
- Preservation and improvement of Ongole cattle. M.N. Nath: 27-29
- The conservation of animal genetic resources in Great Britain. L. Alderson: 31-36

¹Title. author(s): pages

AGRI 5/1986

- Mobilization of the forces of society for the conservation of animal genetic resources. L. Alderson: 1-5
- A role for feral mammals in conserving the genetic diversity of livestock. M.R. Rudge: 9-25

AGRI 6/1987

- The feral cattle of Swona, Orkney Islands. S.J. Hall: 1-9
- Standard de la beni ahsen race ovine marocaine en peril. M. Bourfia: 11-13
- Conservation of animal genetic resources in the USSR. N.G. Dmitriev: 15-18
- Ovino Criollo negro en Brasil. A.H.A. Camargo: 19-23
- Mpwapwa cattle of Tanzania. P.M. Katyega: 25-28
- History of horse domestication. S. Bokonyi: 29-34

AGRI 7/1990

- The Criollo Cattle Project of the British Tropical Agricultural Mission and el Centro de Investigacion Agricola Tropical as a model of investigation and development. I.V. Wilkins: 7-10
- Ganado Criollo del Brasil: origen y características zootecnicas. A.H.A. Camargo: 11-18
- Creation of conservation areas for Hu Sheep in Jangsu province. C. Ruihe: 19-22
- Interspecific hybridization of the Cyprus mouflon (Agrinon) with domestic sheep. A.P. Mavrogenis & A. Herzogi: 23-27
- Evolution of Friesian cattle populations in Spain. V. Calcedo Ordoñez: 29-33
- Conservation des ressources génétiques bovines. L. Avon: 35-43
- Conservation of gene resources of farm animals in the Nordic countries. K. Maijala, A. Neimann-Sorensen, S. Adalsteinsson, N. Kolstad, B. Danell & B. Gjelstad: 45-53
- Research on genetic markers in the Surqo zebu of Somalia. I.A. Abdulcadir, L. Di Stasio, R. Rasero, & G. Sartore: 55-62
- Indigenous pig of Nigeria. N. Pathiraja & E.O. Oyedipe: 63-70
- Polish Red cattle: a scheme for their conservation. K. Zukowski: 71-76
- Istrian cattle. P. Caput & N. Rimanic: 77-82
- The Naked Neck fowl. I. Bodò, G. Kovics, & F. Ludrovsky: 83-88

AGRI 8/1991

- Production potential and breeding schemes of some mediterranean dairy sheep breeds. N.P. Zervas, J. Hatziminaoglou, J. Boyazoglu & A. Georgoudis: 13-26
- La raza ovina latxa: características morfológicas y productivas. Programa de mejora genética. D. Gabiña, E. Urarte, J. Arranz, F. Arrese, I. Beltrán de Heredia: 27-33
- Goats in Yemen. H.U. Hasnain, A.A. Hokhie & A.R.F. Iryani: 35-41
- Apport de l'endocrinologie en selection laitiere bovine. R. Renaville, S. Massart, D. Shojae, M. Sneyers, A. Goffinet, A. Burny & D. Portetelle: 43-47
- Dairy cattle breeding programme and genetic progress in Kenya. R.M. Mzee: 49-54
- La race bovine marocaine blonde d'oulmès-zaër. A. Asri, A. Aittaleb & J.M. Duplan: 55-57
- The maremmana cattle. G. Emiliani: 59-68
- The maremmano horse. M. Silvestrelli: 69-77
- Indigenous chicken genotypes of Ethiopia. M.M. Shanawany & A.K. Banerjee: 79-82

AGRI 9/1992

- Note on the FAO expert consultation on management of global animal genetic resources, Rome, 7-10 April 1992. D.E. Steane: 3-6
- Protection of genetic resources in Central and Eastern Europe. H.-P Grunenfelder: 7-11
- La race Kouri : une population en danger d'absorption. D. Bourzat, A. Idriss & V. Zeuh: 13-21
- Bovino Criollo argentino "Patagonico". A. Rodriguez & R. Martinez: 23-26
- El ganado Criollo colombiano blanco orejinegro (BON). G. Martinez Correal: 27-35
- Ganado Polled Criollo Pereira Camargo, A.H. Almeida Camargo: 37-43
- Wenling Humped and Grassland Red cattle of China. F. Weiqi & Z.W Zhang: 45-47
- Achievements of chinese sheep and goat raising industries over the last forty years. C. Guangren & M. Naixiang: 49-60
- Le mouton Breton. B. Denis & X. Malher: 61-67
- The Skopelos goat breed of Greece. B.G. Pappas, J. Boyazoglu, & Ch. Vasiloudis: 69-76
- El cerdo Zungo. R. Sabogal Ospina & A.A. Owen: 77-83
- The equine breeds of the Murge region of Italy. A. Basile: 85-89

AGRI 10/1992

- Animal genetic resources conservation programme in Brazil. A. da Silva Mariante & A.R. de Bem: 7-26
- Les races animales anciennes: un atout economique pour le sud-ouest de la France. A. Audiot et J.C. Flamant: 27-33
- Study of the Andalusian minor breeds: evaluation of the priorities of conservation. E. Rodero, M.E. Camacho, J.V Delgado & A. Rodero: 35-45
- The camel breeds of India in social and historical perspective. I. Kohler-Rollefson: 47-56
- La raza ovina Rasa aragonesa: caracteres morfológicos y productivos. I. Sierra Alfranca: 57-65
- The Lipizzaner in Italy. A. Borghese: 67-73

AGRI 11/1993

- Documenting the world's domestic animal resources. J. Ruane: 13-20
- Note on the local animal genetic resources and their conservation in Vietnam. L. Viet Ly: 21-24
- Le mérinos de Rambouillet. L. Fiat: 25-33
- Autochthonous sheep breeds in Hungary. I. Bodò, G. Koppany, I. Gera & F. Manczur: 35-40
- Goat breeds in Italy. R. Rubino: 41-55
- Punganur - the miniature Bos indicus cattle. M. Narendra Nath: 57-60
- Mortality patterns in Gir crosses. K.M. Kale & S.D. Mandakmale: 61-67
- La race bovine Asturienne des vallees. S. Dunner, J. Canon, J.P. Gutierrez, M. Vallejo, L. Alonso & F. Goyache: 69-76
- La race bovine Asturienne de montagne. S. Dunner, J. Canon, J.P. Gutierrez, M. Vallejo, L. Alonso & F. Goyache: 77-82

AGRI 12/1993

- Factors of keeping and managing genetic resources of farm animal in 1992. J.J. Lauvergne & P. Souvenir Zafindrajaona: 5-23
- Conservation of animal germplasm at risk of extinction in Italy: the centre for the defense of animal genetic resources of Circello. D. Matassino, A. Cappuccio, T. Grasso & M. Palazzo: 25-43
- Yak (*Poephagus grunzens* L) of India. R.N. Pal: 59-67
- The Garole microsheep of Bengal. India, P. M. Ghalsasi & B. V. Nimbkar: 69-75
- The goats and sheep of the Deccan plateau in the Maharashtra State of India. C. Nimbkar: 77-87
- El cerdo Ibérico y su sistema de producción. J. Rodríguez, L. Silió & S. Martín Rillo: 89-95

AGRI 13/1994

- Introducing the first world watch list for domestic animal diversity. R. Loftus: 3-10
- Japanese native livestock breeds. T. Obata, H. Takeda & T. Oishi: 11-22
- Pourquoi et comment préserver les races bovines Namchi et Kapsiki au Cameroun. B. Sauveroché & E. Thys: 23-37
- Une race trypanotolérante méconnue: la Borgou. J.P. Dehoux & A. Verhulst: 39-45
- Egyptian sheep resources. R.A. Guirgis: 47-58
- Sheep and cattle in Yemen. H.U. Hasnain, A. A. Nokhie & A.R.F. Iryani: 59-63
- Le cheval de Merens.- CNCE & UNIC: 65-73
- The Hungarian Racka. I. Bodó: 75-82

AGRI 14/1994

- Le rôle des biotechnologies de la reproduction pour la conservation des ressources génétiques animales. D. Chupin: 13-25
- Saving the Turopolje pig in Croatia. H.P. Grunenfelder, G. Gugic & F. Punz: 27-32
- Improvement and adaptation of the Fayoumi chicken. M.A. Hossaryl & E.S.E. Galal: 33-39
- Preservation of livestock genetic resources in Bulgaria. T. Dimitrov and L. Dimitrova: 41-59
- The Korean Hanwoo beef cattle. Chan -Won Song: 61-71
- La raza Rubia Gallega ecotipo de montaña. M. Legide & A. Ceular: 73-78
- Population characteristics of water buffaloes in Greek wetlands. A. Georgoudis, Ch. Ligda & J. Boyazoglu: 79-90
- Genetic erosion on camelidae. S. Fernandez-Baca: 91-98
- The German coach horse Saxony Warm Blood. K.U. Sprenger: 99-105

AGRI 15/1995

- Les races d'animaux élevés en Mauritanie. K. Mamoudou: 3-22
- The indigenous domestic animal genetic resources of Uganda. F.M.B. Mbuza: 23-42
- La Chevre Noire marocaine capacités d'adaptation aux conditions arides. J. Hossaini-Hilali & S. Benlamlih: 43-78
- Avances en la conservación y estudio del bovino criollo argentino patagónico. R.D. Martínez & C. A. Rodríguez: 49-57
- Influencia del toro y del sexo sobre el peso al nacer de terneros de raza Criolla de Santa Catarina - Brasil. A.H. Almeida Camargo & C.T. dos Santos Dias: 59-62
- Chinese meat goat breeds and their crosses water buffalo and yak production in China. J. Ying: 63-73
- Water buffalo and Yak production in China. Q. Huai & L. Jun: 75-91

AGRI 16/1995

- History of the Aurochs (*Bos taurus primigenius*) in Poland. Mieczyslaw Rokosz: 5-12
- Genetic improvement of dual purpose cattle in Latin America. L. Vaccaro & D. López: 13-27
- Four interesting endangered breeds of animals in China. You-Chun Chen: 29-35
- Livestock production and animal genetic resources in Croatia. R. T. Wilson: 37-45
- Ressources génétiques animales du Cameroun. O. Messine, V.N. Tanya, D.A. Mbah & C.L. Tawah: 47-63
- Native cattle and horse breeds in Estonia. R. Teinberg, K. Kalamees & A. Kallaste: 65-70
- The Pineia horse. C. Alexandridis: 71-74
- Characterization and conservation of the Malpura sheep breed. S.C. Mehta, P.K. Vij, B.K. Joshi, R. Sahai & A.E. Nivsarkar: 75-82
- Conservation of the Sonadi breed of sheep in India. S.C. Mehta, P.K. Vij, B.K. Joshi, R. Sahai & A.E. Nivsarkar: 83-89
- Description de certaines populations traditionnelles de caprins d'albanie: performances laitières et croissance dans les noyaux de race en milieux difficiles. K. Kume & Z. Bajrami: 91-103

AGRI 17/1996

- Genetic improvement and germplasm conservation in support of a quality products policy. D. Matassino & B.M. Moioli: 5-10
- Le point sur les ressources génétiques en matière d'élevage au Burkina Faso. A.J. Nianogo, R. Sanfo, S.D. Kondombo & S.B. Neya: 11-28
- Note on the founding of the association for the conservation of the early domesticated animals (ACEDA SA) of Southern Africa. J. Bester: 29-33
- Les populations de Bétail présentes au Togo. Y.N. Hadzi: 35-79
- Animal genetic resources in Botswana. E.K. Senyatso & B.S. Masilo: 51-60
- Etudes des ressources génétiques caprines de l'Algérie du nord à l'aide des indices de primarité. E. Khemici, M. Mamou, A. Lounis, D. Bounihi, D. Ouachem, T. Merad & K. Boukhetala: 61-71
- The state of poultry genetic resources in Russia. I.G. Moiseyeva: 73-86
- Situation of inbreeding in a Retinto population. I. Serrano, M. Mayer, A. Rodero & J.M. Jimenez : 87-93
- Fréquence des allèles de la caséine alfa-S1 en race Poitevine. G. Ricordeau, M.F. Mahe, Y. Amigues, F. Grosclaude & E. Manfredi: 95-99
- Bhadawari buffaloes in India. R.K. Pundir, P.K. Vij, Ran Vir Singh and A.E. Nivsarkar: 101-113
- The Criollo sheep in Peru. P.J. Burfening & C. Juan Chavez: 115-125
- White Fulani cattle of West and Central Africa. C.L. Tawah & J.E.O. Rege : 127-145
- Gudali cattle of West and Central Africa. C.L. Tawah & J.E.O. Rege: 147-164

AGRI 18/1996

- Repoblamiento y bioecología de la Vicuña Silvestre en la provincia de Jujuy, Argentina. A.A. Canedi & P.S. Pasini: 7-21
- Mini-horses in China. You-Chun Chen and Tiequan Wang: 23-27
- The Malvi camel: a newly discovered breed from India. I.K. Rollefson & H.S. Rathore: 29-39
- Mithun - an important bovine species of Indian origin. N. Gupta, S.C. Gupta, N.D. Verma, R.K. Pundir, B.I.K. Joshi, A.E. Nivsarkar & R. Sahai: 41-48
- Hariana - an Indian cattle breed in its native ecology. B.K. Joshi, M.S. Tantia, N. Gupta, S.C. Gupta & R. Sahai: 49-57
- The Vechur cattle of Kerala. S. Type: 59-63
- Conservation and utilization of beef cattle genetic resources in Japan. T. Obata, M. Satoh & H. Takeda: 65-78
- Conservation of genetic stock of cattle breeds in the Ukraine. J.D. Ruban: 79-91

AGRI 19/1996

- Growing interest in the Water Buffalo: a short bibliographic update. J. Boyazoglu: 7-15
- El caballo Losino. J.M. Martinez Saiz, M. Valera Cordoba & A. Molina Alcalá: 17-26
- Livestock wealth of the Ladakh: a cold arid region in India. S.C. Gupta, T. Tundup, Neelam Gupta, Pushpendra Kumar, K.N. Yadav, A.E. Nivsarkar, B.K. Joshi & R. Sahai: 27-36
- Siri: the cattle of Eastern Himalayas. M.S. Tantia, P.K. Viji, R.K. Viji, P. Kumar, B.K. Joshi, A.E. Nivsarkar and R. Sahai: 37-43
- Caballo Gallego de Monte (Poney Gallego). I. Sánchez García, A. Iglesias, A. Fernández & J.L. Viana: 45-56
- Goat genetic resources in India and their improvement for increasing productivity. S.C. Chopra: 57-68
- Genetic and phenotypic profiles of endangered Andalusian sheep and goat breeds. E. Rodero, M.R. de la Haba, A. Rodero & M. Herrera: 69-89
- Revue de la situation des races d'animaux domestiques de Cote d'Ivoire. C.V. Yapi-Gnaore, B.A. Oya & Ouattara Zana: 91-108

AGRI 20/1996

- Diversity on animal genetic resources and sustainable development of animal production in China. Shen Changjiang: 3-8
- Studies on feed utilization growth pattern and milk composition in Mithun (*Bos frontalis*). B.P.S. Yadav & N.D. Verma: 9-15
- Conservation of the Tswana cattle breed in Botswana. N. Mpofu: 17-26
- Characterisation of the Siri breed and the Mithun cross Siri in Bhutan. Phanchung & J.A. Roden : 27-34
- Le cheval Mbayar dans la partie centrale du bassin Arachidier sénégalais. J.-P. Dehoux, A. Dieng & A. Buldgen: 35-54
- Paramètres de reproduction et de production des petits ruminants locaux au Burundi. J. Mbayahaga, C. Baudoux, S.N.M. Mandiki, J.L. Bister, R. Branckaert & R. Paquay: 55-69
- Image analysis in morphological animal evaluation: a group for the development of new techniques in zoometry. G. Zehender, L.P. Cordella, A. Chianese, L. Ferrara, A. del Pozzo, S. Barbera, A. Bosticco, P. Negretti, G. Bianconi, G. Filippi Balestra & R. Tonielli: 71-79

AGRI 21/1997

- Conservation de la diversité des ressources génétiques animales dans les pays d'Afrique francophone sub-saharienne. D. Planchenault & J.P. Boutonnet : 1-22
- Morena Gallega cattle breeds with limited numbers: origin, productive characteristics and conservation programmes. . Sánchez García, A. Iglesias, A. Fernández, J.L. Viana & M. Vallejo: 23-33
- Phenotypic characterization of the Saudi Arabian Hassawi cattle breed. T.A. Mohammed: 35-42
- Establishment of a breeding season and evaluation of the reproductive performance of the Pantaneira horse in Mato Grosso do Sul, Brazil. J.R.B. Sereno, S.A. Santos, C.E.S.N. Zúccari & M.C.M. Mazza: 43-48
- The Hungarian Grey cattle: a traditional European breed. L. Bartosiewicz: 49-60
- Zootechnical description of the creole goat of the Oaxaca region, Mexico. A. Sierra, A. Molina, J. Delgado, J. Hernández & M. Rivera: 61-70
- Characterisation of Indian Kathiawari horses. R.K. Pundir, R.K. Vijn, R.N. Shukla, A.S. Vyas, B.K. Bhavsar & A.E. Nivsarkar: 71-80
- Short note: laying hens may have lost important genes. P. Sørensen & L.G. Christensen: 81

AGRI 22/1997

- The Betizu Cattle of the Basque country. M. Gómez, J.M. Plazaola & J.P. Seiliez: 1-5
- The Florina (Pellagonia) sheep breed. D. Triantafillidis, C. Ligda, A. Georgoudis & J. Boyazoglu : 7-13
- Characteristics of Bonpala sheep. P.K. Vij, M.S. Tania & A.E. Nivsarkar: 15-18
- The Chokla Sheep in India. B.P. Kushwaha, S. Kumar, R. Kumar & B.S Mehta: 19-27
- Conservation of an equine feral breed: the Asturcón Pony. J.L. Vega, A. Molina, M. Valera & P.P. Rodriguez-Gallardo: 29-42
- The Criollo horse in Uruguay. F. Vila, M. Valera & A. Molina : 43-52
- Le zébu Gobra: caractères ethniques et performances zootechniques. A. Missohou, A.A. Bankole, A. T. Niang, G. Ragounandea, E. Talaki & I. Bitar: 53-60
- Characteristics of indigenous chickens of Malawi. A.C.L. Safalaoh: 61-69
- The population of laying hens loses important genes: a case history. P. Sørensen: 71-78
- A note on Indian farm animal genetic resources. S.C. Chopra: 79-82

AGRI 23/1998

- Genetic of disease resistance in *Bos taurus* cattle. C.A. Morris: 1-11
- Indigenous cattle of Zanzibar: the need for conservation. K.O. Ali: 13-20
- The use of DNA markers in deciding conservation priorities in sheep and other livestock. A.M. Crawford & R.P. Littlejohn: 21-26
- The development and maintenance of animal recording systems in Greece: a case study. A. Georgoudis & A. Baltas: 27-39
- Caballo Chilote. A. Escobar, J. Oltra, M. Ortiz & J. Voeltz: 41-47
- Review of global rabbit genetic resources: special emphasis on breeding programs and practices in the lesser developed countries. S. D. Lukefahr: 49-67
- Indigenous domestic turkeys of Oaxaca and Quintana Roo, Mexico. J.G. Mallia: 69-78

AGRI 24/1998

- El ganado criollo Romosinuano (Romo). G. Martínez Correal: 1-11
- Genetic resistance to endoparasites in sheep and goats. A review of genetic resistance to gastrointestinal nematode parasites in sheep and goats in the tropics and evidence for resistance in some sheep and goat breeds in sub-humid coastal Kenya. R.L. Baker: 13-30
- The Ponies of the Giara highland. M. Dattilo: 31-39
- The Black Maltese: a Mediterranean, light breed of poultry. J.G. Mallia: 41-48
- Amélioration des performances pondérales du Pigeon au Maroc par croisement de deux races locales avec une race d'origine européenne. B. Benazzouz, A. Soulaymani & A. Mokhtari: 49-61
- Caractéristiques morphobiométriques de la poule du Sénégal. A. Missohou, R.S. Sow & C. Ngwe-Assoumou : 63-69

AGRI 25/1999

- The state of African cattle genetic resources. I. Classification framework and identification of threatened and extinct breeds. J.E.O. Rege: 1-26
 - Characteristics of Munjal sheep. B.P. Kushwaha, Riyazuddin, R. N. Singh & S. Parthasarathy: 27-31
 - Conservation of livestock breed diversity. J.S.F. Barker: 33-43
 - The development of a system of linear measurements to provide an assessment of type and function of beef cattle. G.L.H. Alderson: 45-55
 - A programme for the inventory, characterisation, evaluation, conservation and utilisation of European rabbit (*Oryctolagus cuniculus*) genetic resources. G. Bolet, M. Monnerot, C. Arnal, J. Arnold, D. Bell, G. Bergoglio, U. Besenfelder, S. Bosze, S. Boucher, J.M. Brun, N. Chanteloup, M.C. Ducourouble, M. Durand-Tardif, P.J. Esteves, N. Ferrand, G. Hewitt, T. Joly, P.F. Koehl, M. Laube, S. Lechevestrier, M. Lopez, G. Masoero, R. Piccinin, G. Queney, G. Saleil, A. SurrIDGE, W. van der Loo, J. Vanhommerig, J.S. Vicente, G. Virag & J.M. Zimmermann: 57-71
 - The Peel-Neck chicken of Belize and Guatemala, Central America. J.G. Mallia: 73-78
 - European approaches to conservation of farm animal genetic resources. D.L. Simon: 79-99
 - Inbreeding and loss of founder alleles in four variations of a conservation programme using circular mating, for Danish Shorthorn Cattle. M. Trinderup, J.N. Jørgensen & M. Hansen: 101-108
 - Advances in the Brazilian animal genetic resources conservation programme. A. da S. Mariante, M. do S. M. Albuquerque, A. A. do Egit & C. McManus: 109-123
-

Authors' Index ^{1,2}

- Abdulcadir I.A.: **7:** 55-62
 Adalsteinsson S.: **7:** 45-53
 Adu I.F.: **3:** 1-9
 Aittalebz A.: **8:** 55-57
 Albuquerque M. do S. M.:
25: 111-127
 Al Iryani A.R.F.: **13:** 59-63
 Alderson L.: **4:** 31-36; **5:** 1-5;
25: 45-55
 Alexandridis C.: **16:** 71-74
 Ali K.O.: **23:** 13-20
 Almeida Camargo A.H.: **9:** 37-43;
15: 59-62
 Alonso L.: **11:** 69-76; **11:** 77-82
 Amigues Y.: **17:** 95-99
 Arnal C.: **25:** 57-71
 Arnold J.: **25:** 57-71
 Arranz J.: **8:** 27-33
 Arrese F.: **8:** 27-33
 Asri A.: **8:** 55-57
 Audiot A.: **1:** 25-26; **10:** 27-33
 Avon A.: **7:** 35-43
 Bajrami Z.: **16:** 91-103
 Baker R.L.: **24:** 13-30
 Baltas A.: **23:** 27-39
 Banerjee A.K.: **8:** 79-82
 Bankole A.A.: **22:** 53-60
 Barbera S.: **20:** 71-79
 Barker J.S.F.: **25:** 33-43
 Bartosiewicz L.: **21:** 49-60
 Basile A.: **9:** 85-89
 Baudoux C.: **20:** 55-69
 Bell D.: **25:** 57-71
 Beltrán de Heredia I.: **8:** 27-33
 Benazzouz A.: **24:** 49-61
 Benlamlil S.: **15:** 43-78
 Bergoglio G.: **25:** 57-71
 Besenfelder U.: **25:** 57-71
 Bester J.: **17:** 29-33
 Bhavasar B.K.: **21:** 71-80
 Bianconi G.: **20:** 71-79
 Bister J.L.: **20:** 55-69
 Bitar I.: **22:** 53-60
 Bodó I.: **4:** 19-25; **7:** 83-88;
13: 75-82; **11:** 35-40
 Bökönyi S.: **6:** 29-34
 Bolet G.: **25:** 57-71
 Bonnemaire J.: **12:** 45-57
 Borghese A.: **10:** 67-73
 Bosticco A.: **20:** 71-79
 Bosze S.: **25:** 57-71
 Boucher S.: **25:** 57-71
 Boukhetala K.: **17:** 61-71
 Bounihi D.: **17:** 61-71
 Bourfia M.: **6:** 11-13
 Bourzat D.: **9:** 13-21
 Boutonnet J.P.: **21:** 1-22
 Boyazoglu J.: **8:** 13-26; **9:** 69-76;
14: 79-90; **19:** 7-15; **22:** 7-13
 Branckaert R.: **20:** 55-69
 Brooke C.: **3:** 33-34
 Brun J.M.: **25:** 57-71
 Buldgen A.: **20:** 35-54
 Burfening P.J.: **17:** 115-125
 Burny A.: **8:** 43-47
 Camacho M.E.: **10:** 35-45
 Camargo A.H.A.: **6:** 19-23; **7:** 11-18
 Canedi A.A.: **18:** 7-21
 Canon J.: **11:** 69-76; **11:** 77-82
 Capuccio A.: **12:** 25-43
 Caput P.: **7:** 77-82
 Ceular A.: **14:** 73-78
 Changjiang Shen: **20:** 3-8
 Chanteloup N.: **25:** 57-71
 Chavez J. C.: **17:** 115-125
 Chianese A.: **20:** 71-79
 Chopra S.C.: **19:** 57-68; **22:** 79-82
 Christensen L. G.: **21:** 81
 Chupin D.: **14:** 13-25
 CNCE : **13:** 65-73
 Constantinou A.: **4:** 1-9
 Cordella L.P.: **20:** 71-79
 Crawford A.M.: **23:** 21-26
 Crawford R.D.: **1:** 27-28
 Cunningham E.P.: **1:** 21-24
 da Silva Mariante A.: **10:** 7-26
 Danell B.: **7:** 45-53
 Dattilo M.: **24:** 31-39
 De Alba J.: **4:** 15-17
 de Bem A.R.: **10:** 7-26
 Ducourouble M.C.: **25:** 57-71
 de la Haba M.R.: **19:** 69-89
 Dehoux J.P.: **13:** 39-45; **20:** 35-54
 del Pozzo A. A.: **20:** 71-79
 Delgado J.V.: **10:** 35-45; **21:** 61-70
 Denis B.: **9:** 61-67
 Di Stasio L.: **7:** 55-62
 Dieng A.: **20:** 35-54
 Dimitrov Ts.: **14:** 41-59
 Dimitrova I.: **14:** 41-59
 Dmitriev N.G.: **6:** 15-18
 dos Santos Dias C.T.: **15:** 59-62
 Dunner S.: **11:** 69-76; **11:** 77-82
 Duplan J.M.: **8:** 55-57
 Durand-Tardif M.: **25:** 57-71
 Egit A. A. do: **25:** 111-127
 Emiliani G.: **8:** 59-68
 Escobar A.: **23:** 41-47
 Esteves P.J.: **25:** 57-71
 FAO : **1:** 1-3
 Femandez-Baca S.: **14:** 91-98
 Fernández A.: **19:** 45-56; **21:** 23-33
 Ferrand N.: **25:** 57-71
 Ferrara L.: **20:** 71-79
 Fiat L.: **11:** 25-33
 Filippi Balestra G.: **20:** 71-79
 Flamant J.C.: **10:** 27-33
 Gabiña D.: **8:** 27-33
 Galal E.S.E.: **1:** 5-12; **14:** 33-39
 Georgoudis A.: **8:** 13-26; **14:** 79-90;
22: 7-13; **23:** 27-39
 Gera I.: **11:** 35-40
 Ghalsasi P.M.: **12:** 69-75
 Gjelstad B.: **7:** 45-53
 Goffinet A.: **8:** 43-47
 Gómez M.: **22:** 1-5
 Goonewardene L.A.: **2:** 27-30;
4: 9-13
 Goyache T.: **11:** 69-76; **11:** 77-82
 Grasso F.: **12:** 25-43
 Grosclaude F.: **17:** 95-99
 Grunenfelder H.P.: **9:** 7-11;
14: 27-32
 Guangren C.: **9:** 49-60
 Gugic G.: **14:** 27-32
 Guirgis R.A.: **13:** 47-58
 Gupta Neelam: **18:** 41-48; **18:** 49-57;
19: 27-36
 Gupta S.C.: **18:** 41-48; **18:** 49-57;
19: 27-36
 Gutierrez J.P.: **11:** 69-76; **11:** 77-82
 Hadzi Y.N.: **17:** 35-79
 Hall S.J.: **6:** 1-9
 Hansen M. : **25:** 101-109
 Hasnain H.U.: **8:** 35-41; **13:** 59-63
 Hatziminaoglou J.: **8:** 13-26
 Hernández J.: **21:** 61-70
 Herrera M.: **19:** 69-89
 Herzogi A.: **7:** 23-27
 Hewitt G.: **25:** 57-71
 Hokhie A.A.: **8:** 35-41
 Hossaini-Hilaii J.: **15:** 43-78
 Hossary M.A.: **14:** 33-39
 Huai Qiu: **15:** 75-91
 Idriss A.: **9:** 13-21
 Iglesias A.: **19:** 45-56; **21:** 23-33
 Iryani A.R.F.: **8:** 35-41
 Jest C.: **12:** 45-57
 Jimenez J.M.: **17:** 87-93

¹Author, initials: **issue number**: pages

²Underlined italic indicates main author

- Joly T.: **25**: 57-71
 Jørgensen J.N. : **25**: 101-109
 Joshi B.K.: **16**: 75-82; **16**: 83-89
 Joshi B.: **18**: 41-48; **18**: 49-57;
19: 27-36; **19**: 37-43
 Jun Leo: **15**: 75-91
 Kalamees K.: **16**: 65-70
 Kale K.M.: **11**: 61-67
 Kallaste A.: **16**: 65-70
 Katyega P.M.: **6**: 25-28
 Khemici E.: **17**: 61-71
 Koehl P.F.: **25**: 57-71
 Köhler Rollefson I.: **18**: 29-39;
10: 47-56
 Kolstad B.: **7**: 45-53
 Kondombo S.D.: **17**: 11-28
 Koppány G.: **11**: 35-40
 Kovács G.: **7**: 83-88
 Kumar Pushpendra: **19**: 27-36
 Kumar P.: **19**: 37-43
 Kumar R.: **22**: 19-27
 Kumar S.: **22**: 19-27
 Kume K.: **16**: 91-103
 Kushwaha B.P.: **22**: 19-27; **25**: 27-31
 Laube M.: **25**: 57-71
 Lauvergne J.J.: **12**: 5-23
 Lechevestrier S.: **25**: 57-71
 Legide M.: **14**: 73-78
 Ligda Ch.: **14**: 79-90; **22**: 7-13
 Littlejohn R.P.: **23**: 21-26
 Loftus R.: **13**: 3-10
 López D.: **16**: 13-27
 Lopez M.: **25**: 57-71
 Lounis A.: **17**: 61-71
 Ludrovsky F.: **7**: 83-88
 Lukefahr S.D.: **23**: 49-67
 Mahe M.F.: **17**: 95-99
 Majjala K.: **2**: 20-26; **7**: 45-53
 Malher X.: **9**: 61-67
 Mallia J.G.: **23**: 69-78; **24**: 41-48; **25**:
73-78
 Mamou M.: **17**: 61-71
 Mamoudou K.: **15**: 3-22
 Manczur F.: **11**: 35-40
 Mandakmale S.D.: **11**: 61-67
 Mandiki S.N.M.: **20**: 55-69
 Manfredi E.: **17**: 95-99
 Mariante A. da S.: **25**: 111-127
 Martin Rillo S.: **12**: 89-95
 Martinez L.: **3**: 21-32
 Martinez R.: **9**: 23-26; **15**: 49-57
 Martinez Correal G.: **9**: 27-35;
24: 1-11
 Martinez Saiz J.M.: **19**: 17-26
 Masilo B.S.: **17**: 51-60
 Masoero G.: **25**: 57-71
 Massart S.: **8**: 43-47
 Matassino D.: **12**: 25-43; **17**: 5-10
 Mavrogenis A.P.: **7**: 23-27
 Mayer M.: **17**: 87-93
 Mazza M.C.M.: **21**: 43-48
 Mbah D.A.: **16**: 47-63
 Mbayahaga J.: **20**: 55-69
 Mbuza F.M.B.: **15**: 23-42
 McManus C.: **25**: 111-127
 Mehta B.S.: **22**: 19-27
 Mehta S.C.: **16**: 75-82; **16**: 83-89
 Melgar R.A.: **4**: 15-17
 Merad T.: **17**: 61-71
 Messine O.: **16**: 47-63
 Missohou A.: **22**: 53-60; **24**: 63-69
 Mohammed T.A.: **21**: 35-42
 Moioli B.: **17**: 5-10
 Moiseyeva I.G.: **17**: 73-86
 Mokhtari A.: **24**: 49-61
 Molina A.: **21**: 61-70; **22**: 29-42;
22: 43-52
 Molina Alcala A.: **19**: 17-26
 Monnerot M.: **25**: 57-71
 Moore C.F.: **6**: 1-9
 Morris C.A.: **23**: 1-11
 Mporofu N.: **20**: 17-26
 Mzee R.M.: **8**: 49-54
 Nadarajah K.: **2**: 27-30
 Nagarcanar R.: **1**: 13-16; **1**: 29-30
 Naixiang M.: **9**: 49-60
 Narendra Nath M.: **11**: 57-60
 Nath Mullanpudi N.: **4**: 27-29
 Negretti P.: **20**: 71-79
 Neimam-Sorensen A.: **7**: 45-53
 Neya S.B.: **17**: 11-28
 Ngere L.O.: **3**: 1-9
 Ngwe-Assoumou C.: **24**: 63-69
 Niang A. T.: **22**: 53-60
 Nianogo A.J.: **17**: 11-28
 Nimbkar C.: **12**: 77-87
 Nimbkar B.V.: **12**: 69-75
 Nivsarkar A.E.: **16**: 75-82;
16: 83-89; **17**: 101-113;
18: 41-48; **19**: 27-36; **19**: 37-43;
21: 71-80; **22**: 15-18
 Nokhie A.A.: **13**: 59-63
 Obata T.: **13**: 11-22; **18**: 65-78
 Oishi T.: **13**: 11-22
 Okubanjo I.O.: **3**: 1-9
 Oltra J.: **23**: 41-47
 Ordoñez C.: **7**: 29-33
 Ortiz M.: **23**: 41-47
 Ouachem D.: **17**: 61-71
 Ouattara Z.: **19**: 91-108
 Owen A.A.: **9**: 77-83
 Oya B.A.: **19**: 91-108
 Oyedipe E.O.: **7**: 63-70
 Pal R.N.: **12**: 59-67
 Palazzo M.: **12**: 25-43
 Pappas B.G.: **9**: 69-76
 Paquay R.: **20**: 55-69
 Parthasarathy S.: **25**: 27-31
 Pasini P.S.: **18**: 7-21
 Pathiraja N.: **7**: 63-70
 Phangchung : **20**: 27-34
 Piccinin R.: **25**: 57-71
 Planchenault D.: **21**: 1-22
 Plazaola J.M.: **22**: 1-5
 Portetelvez D.: **8**: 43-47
 Pundir R.K.: **17**: 101-113; **18**: 41-48;
21: 71-80
 Punz E.: **14**: 27-32
 Queney G.: **25**: 57-71
 Ragounandea G.: **22**: 53-60
 Rajaguru A.S.B.: **4**: 9-13
 Rajamahendran R.: **2**: 27-30;
4: 9-13
 Rasero R.: **7**: 55-62
 Rathore H.S.: **18**: 29-39
 Ravindran V.: **2**: 27-30; **4**: 9-13
 Rege J.E.O.: **17**: 127-145; 147-164;
25: 1-26
 Renaville R.: **8**: 43-47
 Ricordeau G.: **17**: 95-99
 Rimanic N.: **7**: 77-82
 Rivera M.: **21**: 61-70
 Riyazuddin.: **25**: 27-31
 Roden J.A.: **20**: 27-34
 Rodero A.: **10**: 35-45; **17**: 87-93;
19: 69-89
 Rodero E.: **10**: 35-45; **19**: 69-89
 Rodríguez J.: **12**: 89-95
 Rodríguez A.: **9**: 23-26
 Rodríguez C. A.: **15**: 49-57
 Rodríguez-Gallardo P.P.: **22**: 29-42
 Rojas F.: **3**: 21-32
 Rokosz M.: **16**: 5-12
 Ruane J.: **11**: 13-20
 Ruban J.D.: **18**: 79-91
 Rubino R.: **11**: 41-55
 Rudge M.R.: **5**: 9-25
 Ruihe C.: **7**: 19-22
 Sabogal Ospina R.: **9**: 77-83
 Safalaoh A.C.L.: **22**: 61-69
 Sahaayaruban P.: **4**: 9-13
 Sahai R.: **16**: 75-82; **16**: 83-89;
18: 41-48; **18**: 49-57; **19**: 27-36
 Sahai R.: **19**: 37-43
 Saleil G.: **25**: 57-71
 Sánchez García I.: **19**: 45-56;
21: 23-33
 Sanfo R.: **17**: 11-28
 Santos S.A.: **21**: 43-48
 Sartore G.: **7**: 55-62
 Satoh Masahiro: **18**: 65-78
 Sauveroché B.: **13**: 23-37

- Seiliez J.P.: **22:** 1-5
 Senyatso E.K.: **17:** 51-60
 Sereno J.R.B.: **21:** 43-48
 Serrano I.: **17:** 87-93
 Shanawany M.M.: **8:** 79-82
 Sheldon B.L.: **3:** 17-20
 Shojae D.: **8:** 43-47
 Shukla R.N.: **21:** 71-80
 Sierra A.: **21:** 61-70
 Sierra Alfranca I.: **10:** 57-65
 Silió L.: **12:** 89-95
 Silvestrelli M.: **8:** 69-77
 Simon D.L.: **25:** 79-99
 Singh. R. N.: **25:** 27-31
 Smith C.: **3:** 11-15; **3:** 11-16
 Sneyers M.: **8:** 43-47
 Solano R.: **4:** 15-17
 Sørensen P.: **21:** 81; **22:** 71-78
 Soulaymani A.: **24:** 49-61
 Souvenir Zafindrajoana P.:
12: 5-23
 Sow R.S.: **24:** 63-69
 Sponenberg D.P.: **3:** 35-38
 Sprenger K.U.: **14:** 99-105
 Steane D.E.: **9:** 3-6
 Surridge A.: **25:** 57-71
 Takeda H.: **13:** 11-22; **18:** 65-78
 Talaki E.: **22:** 53-60
 Tantia M.S.: **18:** 49-57; **19:** 37-43;
22: 15-18
 Tanya V.N.: **16:** 47-63
 Tawah C.L.: **16:** 47-63; **17:** 127-145;
17: 147-164
 Teinberg R.: **16:** 65-70
 Thys E.: **13:** 23-37
 Tiequan Wang: **18:** 23-27
 Tonielli. R.: **20:** 71-79
 Trail J.C.M.: **1:** 17-20
 Triantafillidis D.: **22:** 7-13
 Trinderup M.: **25:** 101-109
 Tundup T.: **19:** 27-36
 Type S.: **18:** 59-63
 UNIC : **13:** 65-73
 Urarte E.: **8:** 27-33
 Vaccaro L.: **16:** 13-27
 Valera M.: **22:** 29-42; **22:** 43-52
 Valera Cordoba M.: **19:** 17-26
 Vallejo M.: **11:** 69-76; **11:** 77-82;
21: 23-33
 van der Loo W. : **25:** 57-71
 Vanhommerig J.: **25:** 57-71
 Vasiloudis Ch.: **9:** 69-76
 Vega J.L.: **22:** 29-42
 Verhulst A.: **13:** 39-45
 Verma N.D.: **18:** 41-48; **20:** 9-15
 Viana J.L.: **19:** 45-56; **21:** 23-33
 Vicente J.S.: **25:** 57-71
 Viet Ly Le: **11:** 21-24
 Vij P.K.: **16:** 75-82; **16:** 83-89;
17: 101-113; **19:** 37-43;
22: 15-18
 Vijn RK.: **19:** 37-43; **21:** 71-80
 Vila F.: **22:** 43-52
 Virag G.: **25:** 57-71
 Vir Singh Ran: **17:** 101-113
 Voeltz J.: **23:** 41-47
 Vyas A.S.: **21:** 71-80
 Weiqi F.: **9:** 45-47
 Wilkins J.V.: **2:** 1-19; **3:** 21-32;
7: 7-10
 Wilson R. T.: **16:** 37-45
 Won Song Chan: **14:** 61-71
 Yadav B.P.S.: **20:** 9-15
 Yadav K.N.: **19:** 27-36
 Yapi-Gnaore C.V.: **19:** 91-108
 Ying Jian: **15:** 63-73
 You-Chun Chen: **16:** 29-35;
18: 23-27
 Zehender G.: **20:** 71-79
 Zervas N.P.: **8:** 13-26
 Zeuh V.: **9:** 13-21
 Zhang Z.W.: **9:** 45-47
 Zimmermann J.M.: **25:** 57-71
 Zúccari C.E.S.N.: **21:** 43-48
 Zukowski K.: **7:** 71-76

Species Index¹

- Buffalo: **4:** 19-25; **10:** 7-26; **11:** 21-24;
14: 41-59; **15:** 75-91; **17:** 101-113;
19: 7-15
- Camelidae: **14:** 91-98; **18:** 7-21
- Camels: **10:** 47-56; **15:** 3-22;
18: 29-39; **19:** 27-36
- Cattle: **1:** 13-16; **1:** 17-20; **1:** 21-24;
1: 25-26; **1:** 29-30; **2:** 1-19;
3: 21-32; **4:** 1-9; **4:** 15-17; **4:** 19-25;
4: 27-29; **6:** 1-09; **6:** 25-28; **7:** 7-10;
7: 11-18; **7:** 29-33; **7:** 35-43;
7: 55-62; **7:** 71-76; **7:** 77-82;
8: 49-54; **8:** 55-57; **8:** 59-68;
9: 13-21; **9:** 23-26; **9:** 27-35;
9: 45-47; **10:** 7-26; **10:** 27-33;
10: 35-45; **11:** 21-24; **11:** 57-60;
11: 61-67; **11:** 69-76; **11:** 77-82;
13: 03-10; **13:** 11-22; **13:** 23-37;
13: 39-45; **14:** 41-59; **14:** 61-71;
14: 73-78; **15:** 03-22; **15:** 23-42;
15: 49-57; **15:** 59-62; **16:** 05-12;
16: 13-27; **16:** 29-35; **16:** 37-45;
16: 47-63; **16:** 65-70; **17:** 11-28;
17: 127-145; **17:** 147-164;
17: 29-33; **17:** 35-79; **17:** 51-60;
17: 87-93; **18:** 49-57; **18:** 59-63;
18: 65-78; **18:** 79-91; **19:** 37-43;
19: 91-108; **20:** 9-15; **20:** 17-26;
20: 27-34; **21:** 23-33; **21:** 35-42;
21: 49-60; **22:** 1-05; **22:** 53-60;
23: 1-11; **23:** 13-20; **24:** 1-11;
25: 45-55; **25:** 101-108;
25: 109-123
- Chicken: **1:** 27-28; **3:** 17-20; **4:** 19-25;
7: 83-88; **8:** 79-82; **11:** 21-24;
13: 11-22; **14:** 33-39; **16:** 37-45;
17: 51-60; **17:** 73-86; **19:** 27-36;
22: 61-69; **22:** 71-78; **24:** 41-48;
24: 63-69; **25:** 73-78
- Dog: **16:** 37-45
- Donkey: **1:** 25-26; **9:** 85-89; **10:** 07-26;
19: 27-36
- Gaur: **13:** 3-10
- General: **1:** 1-3; **2:** 20-26; **7:** 45-53;
8: 43-47; **9:** 7-11; **10:** 35-45;
11: 13-20; **12:** 05-23; **12:** 25-43;
14: 13-25; **17:** 5-10; **20:** 03-8;
20: 71-79; **21:** 1-22; **22:** 79-82;
23: 21-26; **23:** 27-39; **25:** 33-43;
25: 79-99
- Goat: **1:** 25-26; **3:** 1-09; **8:** 35-41;
9: 49-60; **10:** 07-26; **10:** 27-33;
10: 35-45; **11:** 41-55; **12:** 77-87;
13: 11-22; **15:** 03-22; **15:** 23-42;
15: 43-78; **15:** 63-73; **16:** 37-45;
16: 47-63; **16:** 91-103; **17:** 11-28;
17: 29-33; **17:** 35-79; **17:** 51-60;
17: 61-71; **17:** 95-99; **19:** 27-36;
19: 57-68; **19:** 69-89; **19:** 91-108;
20: 55-69; **21:** 61-70; **24:** 13-30
- Horse: **3:** 35-38; **4:** 19-25; **6:** 29-34;
8: 69-77; **9:** 85-89; **10:** 7-26;
10: 27-33; **10:** 67-73; **13:** 11-22;
13: 65-73; **14:** 99-105; **15:** 3-22;
16: 37-45; **16:** 65-70; **16:** 71-74;
17: 11-28; **17:** 29-33; **18:** 23-27;
19: 17-26; **19:** 27-36; **19:** 45-56;
20: 35-54; **21:** 43-48; **21:** 71-80;
22: 29-42; **22:** 43-52; **23:** 41-47;
24: 31-39
- Livestock: **3:** 11-16; **3:** 33-34
- Pig: **4:** 9-13; **4:** 19-25; **7:** 63-70;
9: 77-83; **10:** 7-26; **10:** 27-33;
10: 35-45; **11:** 21-24; **12:** 89-95;
13: 3-10; **14:** 27-32; **14:** 41-59;
15: 23-42; **16:** 29-35; **16:** 37-45;
17: 35-79; **19:** 91-108
- Pigeon: **24:** 49-61
- Rabbit: **23:** 49-67; **25:** 57-71
- Sheep: **1:** 5-12; **1:** 25-26; **2:** 27-30;
4: 01-09; **4:** 19-25; **6:** 11-13;
6: 19-23; **7:** 19-22; **7:** 23-27;
8: 13-26; **8:** 27-33; **9:** 49-60;
9: 61-67; **9:** 69-76; **10:** 7-26;
10: 27-33; **10:** 35-45; **10:** 57-65;
11: 21-24; **11:** 25-33; **11:** 35-40;
12: 69-75; **12:** 77-87; **13:** 3-10;
13: 47-58; **13:** 75-82; **14:** 41-59;
15: 03-22; **15:** 23-42; **16:** 29-35;
16: 37-45; **16:** 47-63; **16:** 75-82;
16: 83-89; **17:** 11-28; **17:** 115-125;
17: 35-79; **17:** 51-60; **19:** 27-36;
19: 69-89; **19:** 91-108; **22:** 7-13;
22: 15-18; **22:** 19-27; **24:** 13-30;
25: 27-31
- Turkey: **23:** 69-78
- Yak: **12:** 45-57; **12:** 59-67; **15:** 75-91;
19: 27-36

¹Species: issue number: pages.

Breed/type Index^{1,2}

- Aansi sheep: **13**: 59-63
 Adal sheep: **1**: 5-12
 Adamawa Gudali cattle:
 17: 147-164
 Afrikaner cattle: **17**: 29-33
 Agrinon sheep: **7**: 23-27
 Americaine cattle: **7**: 35-43
 Amran sheep: **13**: 59-63
 Andalusian Black (Negra Serrana)
 goat: **19**: 69-89
 Andalusian minor breeds: **10**: 35-45
 Andalusian White (Blanca Serrana)
 goat: **19**: 69-89
 Ankole (Songa) cattle: **15**: 23-42
 Arab horse: **15**: 3-22; **17**: 11-28
 Argentata dell'Etna goat: **11**: 41-55
 Arouquesa cattle: **2**: 1-19
 Ashanti Dwarf (Bush) pig: **17**: 35-79
 Asturcón pony: **22**: 29-42
 Asturiana de la Montana cattle:
 11: 77-82
 Asturiana de los Valles cattle:
 11: 69-76
 Attaq goat: **8**: 35-41
 Aurochs: **16**: 5-12
 Auroise cattle: **7**: 35-43
 Australop Black Speckled chicken:
 17: 73-86
 Awassi sheep: **8**: 13-26
 Ayrshire cattle: **8**: 49-54
 Azawak cattle: **17**: 11-28
 Ba Xuyen pig: **11**: 21-24
 Bahawalpur camel: **10**: 47-56
 Bakosi (Kosi) cattle: **13**: 3-10;
 16: 47-63
 Balkan goat: **16**: 37-45
 Baoulé cattle: **17**: 11-28; **19**: 91-108
 Bapedi goat: **17**: 29-33
 Barbe horse: **17**: 11-28; **15**: 3-22
 Barégeoise sheep: **10**: 27-33
 Barki sheep: **13**: 47-58
 Barroso cattle: **2**: 1-19; **4**: 15-17
 Bazadaise cattle: **7**: 35-43
 Be'arnaise cattle: **7**: 35-43
 Beetal goat: **19**: 57-68
 Beldi pigeon: **24**: 49-61
 Beni Ahsen sheep: **6**: 11-13
 Berrenda en Colorado cattle:
 10: 35-45
 Berrenda en Negro cattle: **10**: 35-45
 Betizu cattle: **22**: 1-5
 Bhadawari buffalo: **17**: 101-113;
 19: 7-15
- Bikaneri camel: **10**: 47-56
 Black and White Fresian cattle:
 1: 21-24
 Black Bengal goat: **19**: 57-68
 Black Gascon pig: **10**: 27-33
 Black Maltese chicken: **24**: 41-48
 Black Moroccan (Mambrine
 marocaine) goat: **15**: 43-78
 Black-head sheep: **15**: 23-42
 Blanca Serrana goat: **10**: 35-45
 Blanco Orejinegro Criollo cattle:
 2: 1-19
 Blanco Orejinegro criollo cattle:
 9: 27-35
 Bleue du Nord cattle: **7**: 35-43
 Blonde d'Aquitaine cattle: **10**: 27-33
 Boer goat: **17**: 51-60
 Bonpala sheep: **22**: 15-18
 Bonsmara cattle: **17**: 51-60
 Boran cattle: **1**: 17-20
 Borgou cattle: **17**: 35-79; **13**: 39-45
 Borgou-zébu: **17**: 35-79
 Bornou Foulani sheep: **16**: 47-63
 Bosnian pony horse: **16**: 37-45
 Brahman cattle: **17**: 51-60
 Bretone Pie Noire cattle: **7**: 35-43
 Breznishka sheep: **14**: 41-59
 British rabbit: **25**: 57-71
 Brittany ram: **9**: 61-67
 Brown Carpathian cattle: **18**: 79-91
 Belgian Hare rabbit: **25**: 57-71
 Bulgarian Mediterranean Buffalo:
 14: 41-59
 Burundi local goat: **20**: 55-69
 Cachena cattle: **21**: 23-33
 Caldelana cattle: **21**: 23-33
 Canastra pig: **10**: 7-26
 Caninde goat: **10**: 7-26
 Capore goat: **16**: 91-103
 Carabao buffalo: **10**: 7-26;
 19: 7-15
 Caracu cattle: **2**: 1-19
 Caracu cattle: **10**: 7-26; **13**: 3-10
 Cardena Andaluza cattle: **10**: 35-45
 Caruncho pig: **10**: 7-26
 Casta cattle: **10**: 27-33
 Castillonais horse: **10**: 27-33
 Castillonnaise sheep: **10**: 27-33
 Central American Dairy Criollo
 cattle: **2**: 1-19
 Cerdo Iberico pig: **12**: 89-95
 Chaku alpaca: **14**: 91-98
 Champagne Argente rabbit:
 25: 57-71
 Changthangi goat: **19**: 27-36
 Chegu goat: **19**: 57-68
 Chilote horse: **23**: 41-47
 Chinampo cattle: **2**: 1-19
 Chinchilla rabbit: **25**: 57-71
 Chios sheep: **4**: 1-9; **8**: 13-26
 Chokla sheep: **22**: 19-27
 Churra sheep: **8**: 13-26
 Churro Lebrijano sheep: **10**: 35-45
 Ciggja (Tsigai) sheep: **4**: 19-25
 Cikta sheep: **4**: 19-25; **11**: 35-40
 Comisana sheep: **8**: 13-26
 Copper-Red Shoumen sheep:
 14: 41-59
 Costeño Criollo cattle: **2**: 1-19
 Criollo Argentina cattle: **2**: 1-19
 C. Argentino cattle: **9**: 23-26
 C. Bolivian cattle: **2**: 1-19;
 3: 21-32; **7**: 7-10
 C. Brazilian cattle: **2**: 1-19;
 7: 11-18
 C. Brazilian donkey: **10**: 7-26
 C. Colombian cattle: **2**: 1-19;
 9: 27-35
 C. Costa Rica cattle: **2**: 1-19
 C. Cuban cattle: **2**: 1-19
 C. de Santa Catarina (Brasil)
 cattle: **15**: 59-62
 C. Dominican Republic cattle:
 2: 1-19
 C. Ecuador cattle: **2**: 1-19
 C. Guatemala cattle: **2**: 1-19
 C. Lanado Brazilian sheep:
 10: 7-26
 C. Mexico cattle: **2**: 1-19
 C. Negro sheep: **6**: 19-23
 C. of Lages (Franqueiro) cattle:
 10: 7-26
 C. Patagonic cattle: **15**: 49-57
 C. Peru sheep: **17**: 115-125
 C. Uruguay horse: **22**: 43-52
 C. Venezuela cattle: **2**: 1-19
 Croatian sheepdog: **16**: 37-45
 Curraleiro (Hard Hoof Criollo)
 cattle: **10**: 7-26
 Cyprus Fat Tailed sheep: **4**: 1-9
 Dabenska sheep: **14**: 41-59
 Damascus (Shami) goat: **4**: 1-9
 Danish Shorthorn cattle: **25**: 101-108
 Dapaong pig: **17**: 35-79
 Datum cattle: **20**: 27-34
 De (Small) buffalo: **11**: 21-24

¹As the main concern of the paper.²Breed/type: **issue number**: pages.

- Debao Mini-horse: **18:** 23-27
 Deccani sheep: **12:** 77-87
 Dechang Water buffalo: **15:** 75-91
 Dhamari (Burri or Jahrani) sheep: **13:** 59-63
 Dhamariz sheep: **13:** 47-58
 Dioungy goat: **15:** 3-22; **19:** 91-108
 Djallonké sheep: **16:** 47-63; **17:** 35-79; **19:** 91-108
 Doebum cattle: **20:** 27-34
 Dong Cao chicken: **11:** 21-24
 Dorper sheep: **17:** 51-60; **24:** 13-30
 Dragobia goat: **16:** 91-103
 Dukati goat: **16:** 91-103
 East African goat: **15:** 23-42
 East African long-tail sheep: **15:** 23-42
 East-African Short-Horn (Nkedi) zebu: **15:** 23-42
 Eastern-Balkan (Iztochno-Balkanska) pig: **14:** 41-59
 Egyptian buffalo: **19:** 7-15
 Espiritu Criollo cattle: **2:** 1-19
 Estonian Native cattle: **16:** 65-70
 Estonian Native horse: **16:** 65-70
 Fauve de Bourgogne rabbit: **25:** 57-71
 Fayoumi (Ramadi) chicken: **14:** 33-39
 Ferrandaise cattle: **1:** 25-26; **7:** 35-43
 Flamande cattle: **7:** 35-43
 Flemish Giant rabbit: **25:** 57-71
 Florina (Pellagonia) sheep: **22:** 7-13
 French Lop rabbit: **25:** 57-71
 Friesian cattle: **4:** 1-9; **7:** 29-33; **8:** 49-54
 Frieiresa cattle: **21:** 23-33
 Froment du Leon cattle: **7:** 35-43
 Frontalasca goat: **11:** 41-55
 Gaddi goat: **19:** 57-68
 Galician pony: **19:** 45-56
 Galla goat: **24:** 13-30
 Gangam goat: **19:** 57-68
 Garganica goat: **11:** 41-55
 Garole sheep: **12:** 69-75
 Gasconne cattle: **10:** 27-33
 Gaur of Northern India: **13:** 3-10
 Gebshima chicken: **8:** 79-82
 General: **1:** 1-3; **6:** 29-34; **6:** 15-18; **7:** 45-53; **9:** 3-6
 General (cattle): **8:** 43-47
 Giara pony: **24:** 31-39
 Gidran horse: **4:** 19-25
 Gir cattle: **25:** 109-123
 Girgentana goat: **11:** 41-55
 Gobra zebu: **22:** 53-60
 Gohilwadi goat: **19:** 57-68
 Golurratke kokos poultry: **16:** 37-45
 Goudali cattle: **16:** 47-63
 Grassland Dwarf goat: **16:** 47-63
 Grassland Red cattle: **9:** 45-47
 Grazalema Merino sheep: **19:** 69-89
 Grey Iskar cattle: **14:** 41-59
 Grey Ukrainian cattle: **18:** 79-91
 Gudali cattle: **17:** 147-164
 Guernseys cattle: **8:** 49-54
 Guizhou Mini-horse: **18:** 23-27
 Guzerat cattle: **25:** 109-123
 Hainan cattle: **16:** 29-35
 Haitian pig: **13:** 3-10
 Halfbred horse: **4:** 19-25
 Hanwoo cattle: **14:** 61-71
 Hariana cattle: **1:** 29-30; **18:** 49-57
 Hasi goat: **16:** 91-103
 Hassawi cattle: **21:** 35-42
 Hengduan (High mountain or gorge) type yak: **15:** 75-91
 Hi-line chicken: **17:** 51-60
 Himalayan rabbit: **25:** 57-71
 Hinadori fowl: **13:** 11-22
 Ho chicken: **11:** 21-24
 Hokkaido pony: **13:** 11-22
 Holstein-Friesian cattle: **16:** 37-45
 Horro sheep: **1:** 5-12
 Heavy Warmblood horse: **14:** 99-105
 Hu sheep: **7:** 19-22; **16:** 29-35; **20:** 3-8
 Huai goat: **15:** 63-73
 Humped (bacterin) camel: **19:** 27-36
 Hungarian grey cattle: **4:** 19-25; **21:** 49-60
 Indigenous chicken types of Ethiopia: **8:** 79-82
 Indigenous Malawi chickens: **22:** 61-69
 Indigenous Mexican turkey: **23:** 69-78
 Indigenous Nigerian pig: **7:** 63-70
 Indigenous Uganda pig: **15:** 23-42
 Indubrasil cattle: **25:** 109-123
 Ionica goat: **11:** 41-55
 Isa Brown chicken: **17:** 51-60
 Istrian cattle: **7:** 77-82
 Istrian Podolaz cattle: **16:** 37-45
 Jafarabandi buffalo: **19:** 7-15
 Jaisalmeri camel: **10:** 47-56
 Jamunapari goat: **19:** 57-68
 Japanese Black cattle: **18:** 65-78
 Japanese Brown cattle: **18:** 65-78
 Japanese Poll cattle: **18:** 65-78
 Japanese Shorthorn cattle: **18:** 65-78
 Jatsha cattle: **20:** 27-34
 Jatsham cattle: **20:** 27-34
 Jerseys cattle: **8:** 49-54
 Jhakrana goat: **19:** 57-68
 Jurlov Crower chicken: **17:** 73-86
 Kabylie dwarf goat: **17:** 61-71
 Kano Brown goat: **3:** 1-9
 Kapsiki cattle: **13:** 23-37
 cattle: **16:** 47-63
 Kara alpaca: **14:** 91-98
 Karagouniko sheep: **8:** 13-26
 Karakachanska sheep: **14:** 41-59
 Karnobatska sheep: **14:** 41-59
 Kathiawari horse: **21:** 71-80
 Kei chicken: **8:** 79-82
 Kigezi goat: **15:** 23-42
 Kirdi sheep: **16:** 47-63
 Kisbér Halfbred horse: **4:** 19-25
 Kiso pony: **13:** 11-22
 Koeyoshi fowl: **13:** 11-22
 Korhogo pig: **19:** 91-108
 Kottenska sheep: **14:** 41-59
 Kouri cattle: **9:** 13-21
 Kuchino Jubilee chicken: **17:** 73-86
 Kuchinoshima cattle: **13:** 11-22
 Kurokashiwa fowl: **13:** 11-22
 Kutch camel: **10:** 47-56
 Lacaune cattle: **10:** 27-33
 Lacaune sheep: **8:** 13-26
 Lacha sheep: **8:** 13-26; **8:** 27-33
 Ladakh poultry: **19:** 27-36
 Ladakhi donkey: **19:** 27-36
 Lagunaire cattle: **17:** 35-79
 Lagune (Lagunaire) cattle: **19:** 91-108
 Land Race pig: **17:** 35-79
 Landais sheep: **1:** 25-26
 Landes de Bretagne ram: **9:** 61-67
 Large White pig: **16:** 37-45; **17:** 35-79
 Lavradeiro Criollo (Wild Horse of Roraima) horse: **10:** 7-26
 Lebedinskaya cattle: **18:** 79-91
 Lebrijan Churro sheep: **19:** 69-89
 Leningrad Golden Gray chicken: **17:** 73-86
 Lick Ovca sheep: **16:** 37-45
 Limiana cattle: **21:** 23-33
 Limonero Criollo cattle: **2:** 1-19
 Lipizza horse: **4:** 19-25; **10:** 67-73; **16:** 37-45
 Livestock (general): **3:** 11-16
 Local Brazilian pig: **10:** 7-26
 Longhaired Maure sheep: **15:** 3-22
 Longling goat: **15:** 63-73

- Losino horse: **19**: 17-26
 Lourdaise cattle: **7**: 35-43; **10**: 27-33
 Lourdaise sheep: **10**: 27-33
 Macau pig: **10**: 7-26
 Malpura sheep: **16**: 75-82
 Maltese goat: **11**: 41-55
 Malvi camel: **18**: 29-39
 Manchado de Jabugo pig: **10**: 35-45
 Manchega sheep: **8**: 13-26
 Mangalica (Mangalitsa) pig: **4**: 19-25
 Marab white sheep: **13**: 59-63
 Maracay Criollo cattle: **2**: 1-19
 Maradi goat: **17**: 35-79
 Maremmana cattle: **8**: 59-68
 Maremmano horse: **8**: 69-77
 Marota (Curaca) goat: **10**: 7-26
 Maroua sheep: **16**: 47-63
 Martinafranca donkey: **9**: 85-89
 Marwari camel: **10**: 47-56
 Marwari goat: **19**: 57-68
 Masai sheep: **15**: 23-42
 Massese sheep: **8**: 13-26
 Mati goat: **16**: 91-103
 Matishka sheep: **14**: 41-59
 Matou goat: **15**: 63-73
 Maure zebu: **11**: 3-22
 Mawr goat: **8**: 35-41
 Mbayar horse: **20**: 35-54
 Mbororo (Red Fulani or Red-longhorn) zebu: **17**: 35-79
 Mbororo cattle: **16**: 47-63
 Mediterranean buffalo: **19**: 7-15
 Menz sheep: **1**: 5-12
 Méré cattle: **19**: 91-108
 Merens horse: **13**: 65-73
 Merino de Grazalema sheep: **10**: 35-45
 Merino Espanol sheep: **10**: 35-45
 Merino of Rambouillet sheep: **11**: 25-33
 Mewari camel: **10**: 47-56
 Mezohegyes horse: **4**: 19-25
 Mgandi pigeon: **24**: 49-61
 Mia chicken: **11**: 21-24
 Minhota cattle: **2**: 1-19
 Mini Co pig: **11**: 21-24
 Mini-horse: **18**: 23-27
 Minohiki fowl: **13**: 11-22
 Mirandesa cattle: **2**: 1-19
 Misaki pony: **13**: 11-22
 Mishima cattle: **13**: 11-22
 Mithun cattle: **20**: 9-15; **18**: 41-48
 Miyako pony: **13**: 11-22
 Mocho Nacional cattle: **10**: 7-26
 Mong Cai pig: **11**: 21-24
 Montecristo goat: **11**: 41-55
 Montesina sheep: **10**: 35-45
 Morada Nova (Deslanado do Nordeste) sheep: **10**: 7-26
 Morena Gallega cattle: **21**: 23-33
 Moroccan Blonde d'Oulmès-Zaër cattle: **8**: 55-57
 Moscow chicken: **17**: 73-86
 Mossi sheep: **17**: 11-28
 Moura pig: **10**: 7-26
 Moutourou (Bakweri) cattle: **16**: 47-63
 Moxoto goat: **10**: 7-26
 Mpwapwa cattle: **6**: 25-28
 Mubende goat: **15**: 23-42
 Munjal sheep: **25**: 27-31
 Muong Khuong pig: **11**: 21-24
 Murciana cattle: **10**: 35-45
 Murge horse: **9**: 85-89
 Murrah buffalo: **19**: 7-15;
 Muzhaka goat: **16**: 91-103
 N'Dama cattle: **1**: 17-20; **16**: 47-63;
17: 11-28; **17**: 35-79; **19**: 91-108
 N'Damance cattle: **19**: 91-108
 N'Damaze cattle: **19**: 91-108
 Naked Neck Fowl: **7**: 83-88
 Naked-Neck Black poultry: **4**: 19-25
 Namchi cattle: **13**: 23-37
 Nanjiang yellow goat: **15**: 63-73
 Nantaise cattle: **7**: 35-43
 Native: see also Baladi, Beldi
 Native pig of Sri Lanka: **4**: 9-13
 Native sheep Sri Lanka: **2**: 27-30
 Negra de las Campinas cattle: **10**: 35-45
 Negra Serrana goat: **10**: 35-45
 Nellore cattle: **25**: 109-123
 Netch chicken: **8**: 79-82
 Ngo (Big) buffalo: **11**: 21-24
 Nguni cattle: **17**: 29-33
 Nilo pig: **10**: 7-26
 Ningqiang Mini-horse: **18**: 23-27
 Noma pony: **13**: 11-22
 Nonius horse: **4**: 19-25
 Nooitgedachy pony: **17**: 29-33
 Old-Oldenburger horse: **14**: 99-105
 Olkuz sheep: **13**: 3-10
 Onagadori fowl: **13**: 11-22
 Ongole cattle: **4**: 27-29
 Orloff chicken: **17**: 73-86
 Orobica goat: **11**: 41-55
 Osmanabadi goat: **12**: 77-87;
19: 57-68
 Ossimi sheep: **13**: 47-58
 Ouessant sheep: **9**: 61-67
 Pajuna cattle: **10**: 35-45
 Paka Ovca sheep: **16**: 37-45
 Pantaneiro Criollo (Swamp Criollo) cattle: **10**: 7-26
 Pantaneiro Criollo horse: **10**: 7-26;
21: 43-48
 Parthenaise cattle: **7**: 35-43
 Pashmina type goat: **19**: 27-36
 Pastoreño creole goat: **21**: 61-70
 Payoya goat: **10**: 35-45
 Peel-Neck chicken: **25**: 73-78
 Peuhl sheep: **16**: 47-63
 Peulh (Chemama) zebu: **15**: 3-22;
17: 11-28
 Phan Rang sheep: **11**: 21-24
 Piau pig: **10**: 7-26
 Pineia horse: **16**: 71-74
 Pinzgau cattle: **18**: 79-91
 Pirapetinga pig: **10**: 7-26
 Plateau (Qingzang or Grassland) type yak: **15**: 75-91
 Plymouth Rock Barred Speckled chicken: **17**: 73-86
 Poitevine goat: **17**: 95-99
 Poitou ass: **1**: 25-26
 Polish Red cattle: **7**: 71-76
 Polled Criollo Pereira Camargo cattle: **9**: 37-43
 Posavina horse: **16**: 37-45
 Poultry (general): **1**: 27-28
 Pramenka sheep: **16**: 37-45
 Punganur dwarflike cattle: **11**: 57-60
 Pushkin Barred Speckled chicken: **17**: 73-86
 Pyrénées goat: **10**: 27-33
 Racka sheep: **4**: 19-25; **11**: 35-40;
13: 75-82
 Rahmani sheep: **13**: 47-58
 Raiole sheep: **1**: 25-26
 Rare breeds: **9**: 7-11
 Rasa Aragonesa sheep: **10**: 57-65
 Red and White Holstein cattle: **1**: 21-24
 Red Fulani cattle: **16**: 47-63
 Red Maasai sheep: **24**: 13-30
 Red Polled cattle: **18**: 79-91
 Red Sokoto goat: **3**: 1-9; **17**: 35-79
 Red Steppe cattle: **18**: 79-91
 Regeuibi (Sahel) dromadaire: **15**: 3-22
 Repartida goat: **10**: 7-26
 Replyanska sheep: **14**: 41-59
 Retinto cattle: **17**: 87-93
 Rewari (Rebari) camel: **10**: 47-56
 Rhodope's Short-Horned cattle: **14**: 41-59
 Ri chicken: **11**: 21-24
 River buffalo: **19**: 7-15
 Romosinuano (Romo) Criollo cattle: **2**: 1-19; **24**: 1-11
 Rossa Mediterranea goat: **11**: 41-55
 Rove goat: **1**: 25-26
 Rubia Gallega cattle: **14**: 73-78
 Ruda Dubrovacka sheep: **16**: 37-45
 Russian Black Bearded chicken: **17**: 73-86
 Russian White chicken: **17**: 73-86
 Sahel goat: **3**: 1-9

- Sahel goat: **15:** 3-22; **17:** 35-79;
17: 11-28
- Sahiwal cattle: **1:** 13-16; **1:** 17-20
- Saidi sheep: **13:** 47-58
- San Martinero Criollo cattle: **2:** 1-19
- Sanara white sheep: **13:** 59-63
- Sangamneri goat: **12:** 77-87;
19: 57-68
- Santa Ines sheep: **10:** 7-26
- Sarda goat: **11:** 41-55
- Sarda sheep: **8:** 13-26
- Saxony Warmblood horse:
14: 99-105
- Schweres Warmblut horse:
14: 99-105
- Senegal chicken: **24:** 63-69
- Shagya Arab (Babolna) horse:
4: 19-25
- Shiba goat: **13:** 11-22
- Sindi cattle: **25:** 109-123
- Shorthaired Maure sheep: **15:** 3-22
- Shuwa-aral (Fellatat or Wadara)
zebu: **17:** 35-79
- Shyta goat: **16:** 91-103
- Sichuan Mini-horse: **18:** 23-27
- Simmental cattle: **16:** 37-45
- Siri cattle: **19:** 37-43; **20:** 27-34
- Sirohi goat: **19:** 57-68
- Skalborg Hen: **22:** 71-78
- Skopelos sheep: **9:** 69-76
- Small East African goat: **24:** 13-30
- Small East African zebu: **23:** 13-20
- Sokoto (Sokoto-Gudali or Bokoloji)
zebu: **17:** 35-79
- Sokoto goat: **16:** 47-63
- Sokoto Gudali cattle: **17:** 147-164
- Somali Blackhead sheep: **1:** 5-12
- Somba (Atakora) cattle: **17:** 35-79
- Sonadi sheep: **16:** 83-89
- Sottobanaca pigeon: **24:** 49-61
- Spanish goat: **15:** 3-22
- Spanish (Mustang) horse: **3:** 35-38
- Speckled Hungarian poultry:
4: 19-25
- Srednogorska sheep: **14:** 41-59
- Starozagorvka sheep: **14:** 41-59
- Strandjanska sheep: **14:** 41-59
- Surdudi goat: **8:** 35-41
- Surqo zebu: **7:** 55-62
- Surti goat: **12:** 77-87; **19:** 57-68
- Svishtoiska sheep: **14:** 41-59
- Swamp buffalo: **19:** 7-15
- Swedish Landrace pig: **16:** 37-45
- Swona cattle: **6:** 1-9
- Tabapuã cattle: **25:** 109-123
- Taishu pony: **13:** 11-22
- Taiz Black goat: **8:** 35-41
- Taiz Red goat: **8:** 35-41
- Taiz red sheep: **13:** 59-63
- Tan sheep: **20:** 3-8
- Thamud goat: **8:** 35-41
- Thoroughbred horse: **16:** 37-45
- Thrabum cattle: **20:** 27-34
- Thuoc Nhieu pig: **11:** 21-24
- Thuringer rabbit: **25:** 57-71
- Tibetan sheep: **19:** 27-36
- Tihami sheep: **13:** 59-63
- Tikur chicken: **8:** 79-82
- Tipo Baio (Brown type) buffalo:
10: 7-26
- Tokara goat: **13:** 11-22
- Tokara pony: **13:** 11-22
- Totenko fowl: **13:** 11-22
- Trakehner horse: **16:** 37-45
- Tropical Dairy Criollo cattle:
2: 1-19
- Trotter horse: **16:** 37-45
- Tsigai sheep: **11:** 35-40
- Tswana cattle: **17:** 51-60; **20:** 17-26
- Tswana goat: **17:** 51-60
- Tucura or Cuiabano cattle: **10:** 7-26
- Tuli cattle: **17:** 51-60
- Turopolje pig: **14:** 27-32; **16:** 37-45
- Tuyen Quang buffalo: **11:** 21-24
- Vallese goat: **11:** 41-55
- Variou: **9:** 49-60
- Vechur cattle: **18:** 59-63
- Velipoja goat: **16:** 91-103
- Verzaschese goat: **11:** 41-55
- Vianesa cattle: **21:** 23-33
- Vicugna: **14:** 91-98
- Vienna White rabbit: **25:** 57-71
- Villard de Lans cattle: **7:** 35-43
- Vogan sheep: **17:** 35-79
- Vosgieme cattle: **7:** 35-43
- Wakwa cattle: **16:** 47-63
- Water buffalo: **4:** 19-25; **15:** 75-91;
25: 33-43
- Wenling Humped cattle: **9:** 45-47
- West African Dwarf goat: **3:** 1-9
- White Fulani cattle: **16:** 47-63;
17: 127-145
- White Hungarian poultry: **4:** 19-25
- White Leghorn chicken: **22:** 71-78
- White-Fulani zebu: **17:** 35-79
- White-Headed Ukrainian cattle:
18: 79-91
- White Park cattle: **25:** 45-55
- Wuzhishan pig: **16:** 29-35
- Xilin buffalo: **15:** 75-91
- Xinulung buffalo: **15:** 75-91
- Yacumeño Criollo cattle: **2:** 1-19
- Yak: **19:** 27-36
- Yangka cattle: **20:** 27-34
- Yangkum cattle: **20:** 27-34
- Yellow (Gold) Hungarian poultry:
4: 19-25
- Yellow Vietnam cattle: **11:** 21-24
- Yemen white sheep: **13:** 59-63
- Yemeni goat: **8:** 35-41
- Yemeni Mountain goat: **8:** 35-41
- Yen Bai buffalo: **11:** 21-24
- Yerevan chicken: **17:** 73-86
- Yonaguni pony: **13:** 11-22
- Yunnan Mini-horse: **18:** 23-27
- Zalawadi goat: **19:** 57-68
- Zaniskari horse: **19:** 27-36
- Zanzibar Zebu: **23:** 13-20
- Zaryanka chicken: **17:** 73-86
- Zebu: **16:** 47-63
- Zhoushan cattle: **16:** 29-35
- Zungo pig: **9:** 77-83

**Main geographical entities
of material reported in
AGRI¹**

- Albania: **16:** 91-103
 Algeria: **17:** 61-71
 Argentina: **18:** 07-21; **15:** 49-57;
9: 23-26
 Belize: **25:** 73-78
 Benin: **13:** 39-45
 Bhutan: **20:** 27-34
 Bolivia: **3:** 21-32; **7:** 07-10
 Botswana: **17:** 51-60; **20:** 17-26
 Brazil: **7:** 11-18; **6:** 19-23; **21:** 43-48;
15: 59-62; **9:** 37-43; **10:** 07-26;
25: 109-123
 Bulgaria: **14:** 41-59
 Burkina Faso: **17:** 11-28; **13:** 39-45
 Burundi: **20:** 55-69
 Cameroon: **16:** 47-63; **13:** 23-37
 Canada: **1:** 27-28
 Central and Eastern Europe:
9: 07-11
 Central Asia region: **12:** 45-57
 Chad: **9:** 13-21
 China: **20:** 03-8; **15:** 63-73; **18:** 23-27;
15: 75-91; **16:** 29-35; **9:** 49-60;
9: 45-47; **7:** 19-22
 Chile: **23:** 41-47
 Colombia: **24:** 01-11; **9:** 77-83;
9: 27-35
 Croatia: **16:** 37-45; **14:** 27-32
 Cyprus: **4:** 01-09; **7:** 23-27
 Denmark: **22:** 71-78; **25:** 101-108
 Egypt: **13:** 47-58; **14:** 33-39
 Estonia: **16:** 65-70
 Ethiopia: **1:** 05-12; **8:** 79-82
 Europe: **1:** 21-24; **25:** 57-71; **25:** 79-99
 European Nordic Countries:
7: 45-53
 France: **10:** 27-33; **1:** 25-26;
17: 95-99; **13:** 65-73; **7:** 35-43;
9: 61-67; **11:** 25-33
 General: **3:** 11-16; **17:** 05-10;
8: 43-47; **13:** 03-10; **6:** 29-34;
12: 05-23; **11:** 13-20; **14:** 13-25;
9: 03-6; **2:** 01-19; **23:** 21-26; **19:**
 07-15; **20:** 71-79; **23:** 01-11;
23: 49-67
 Germany: **14:** 99-105
 Greece: **14:** 79-90; **22:** 07-13;
16: 71-74; **9:** 69-76; **23:** 27-39
 Guatemala: **4:** 15-17; **25:** 73-78
 Hungary: **7:** 83-88; **21:** 49-60;
13: 75-82; **11:** 35-40
 India: **12:** 77-87; **12:** 69-75;
11: 57-60; **22:** 19-27; **19:** 27-36;
12: 59-67; **19:** 37-43; **20:** 09-15;
10: 47-56; **11:** 61-67; **18:** 41-48;
1: 29-30; **16:** 83-89; **16:** 75-82;
1: 13-16; **17:** 101-113; **4:** 27-29;
18: 59-63; **18:** 29-39; **21:** 71-80;
18: 49-57; **22:** 79-82; **22:** 15-18;
19: 57-68; **25:** 27-31
 Italy: **9:** 85-89; **24:** 31-39; **8:** 69-77;
8: 59-68; **12:** 25-43; **10:** 67-73;
11: 41-55
 Ivory Coast: **19:** 91-108
 Japan: **18:** 65-78; **13:** 11-22
 Kenya: **24:** 13-30; **1:** 17-20; **8:** 49-54
 Korea: **14:** 61-71
 Latin America: **16:** 13-27
 Malawi: **22:** 61-69
 Malta: **24:** 41-48
 Mauritania: **15:** 03-22
 Mediterranean basin: **8:** 13-26
 Mexico: **21:** 61-70; **23:** 69-78
 Morocco: **8:** 55-57; **15:** 43-78;
6: 11-13; **24:** 49-61
 New Zealand: **5:** 01-05; **5:** 09-25
 Nigeria:
 Nigeria: **3:** 01-09; **7:** 63-70; **13:** 39-45
 North America: **3:** 35-38
 Peru: **17:** 115-125
 Poland: **16:** 05-12; **7:** 71-76
 Russia: **17:** 73-86
 SABRAO Region: **1:** 01-03; **3:** 17-20
 Saudi Arabia: **21:** 35-42
 Scandinavia: **2:** 20-26
 Senegal: **22:** 53-60; **20:** 35-54;
24: 63-69
 Somalia: **7:** 55-62
 South Africa: **14:** 91-98; **17:** 29-33;
 Spain: **11:** 77-82; **17:** 87-93;
19: 69-89; **10:** 35-45; **11:** 69-76;
22: 01-05; **10:** 57-65; **8:** 27-33;
19: 17-26; **12:** 89-95; **19:** 45-56;
7: 29-33; **21:** 23-33; **14:** 73-78;
22: 29-42
 Sri Lanka: **4:** 09-13; **2:** 27-30
 Sub-Saharan French-speaking
 countries: **21:** 01-22
 Tanzania: **6:** 25-28
 Togo: **17:** 35-79
 Uganda: **15:** 23-42
 UK: **4:** 31-36; **5:** 01-05; **6:** 01-09;
25: 45-55
 Ukraine: **18:** 79-91
 Uruguay: **22:** 43-52
 USA: **3:** 33-34
 USSR: **6:** 15-18
 Vietnam: **11:** 21-24
 West and Central Africa:
17: 147-164; **17:** 127-145
 Yemen: **8:** 35-41; **13:** 59-63
 Yugoslavia: **7:** 77-82
 Zanzibar: **23:** 13-20

¹Country: **issue number:** pages

Pages refer to those on the electronic version (CD Rom and Internet), which might not coincide with the hard copy.

Selected secondary citations^{1,2}

- Abathwa chicken: **22**: 61-69
Aberdeen Angus cattle: **12**: 45-57;
18: 65-78; **25**: 45-55
Abondance cattle: **19**: 91-108;
21: 1-22
Acacia ataxacantha: **19**: 91-108
Acacia millifera: **1**: 5-12
Acacia nilotica: **18**: 29-39
Acacia senegal: **1**: 5-12
Acacia tortizis: **1**: 5-12
Acantholippia deserticola: **18**: 7-21
Acremonium coenophialum:
23: 1-11
Adamaoua region: **16**: 47-63
Adamawa Gudali cattle:
17: 147-164
Adamawa mountain: **17**: 147-164
Aegean islands: **22**: 7-13
Aegean sea: **9**: 69-76
African Barb horse: **3**: 35-38
African Star cattle: **4**: 15-17
African Swine Fever: **13**: 3-10
Africander cattle: : **17**: 51-60;
20: 17-26; **23**: 1-11
Agenda 21: **21**: 1-22; **25**: 79-99
Agios Mamas Experimental Station
of Halkidiki: **8**: 13-26
Agrostis spp.: **9**: 37-43
Ahmednagar district: **12**: 77-87
Ajmer: **1**: 29-30
Akhelia region: **4**: 1-9
Akita prefecture: **13**: 11-22
Akou cattle: **21**: 1-22
Al Jawf region: **8**: 35-41
Alantika mountain: **13**: 23-37
Alava: **8**: 13-26
Albumin: **19**: 69-89; **23**: 41-47
Alentejo cattle: **7**: 11-18
Alentejo region: **12**: 89-95
Alonisos island: **9**: 69-76
Alpaca: **14**: 91-98
Alpine goat: **11**: 41-55; **17**: 61-71
Alta Valtellina region: **11**: 41-55
Altai fat-rumped sheep: **20**: 3-8
Altamira (Spain): **16**: 5-12
Amazon region: **10**: 7-26
Amazon river: **9**: 27-35
- American Indian Horse Registry
(AIHR): **3**: 35-38
American Minor Breeds
Conservancy: **3**: 33-34
American Mustang Association
(AMA): **3**: 35-38
Amritmahal cattle: **22**: 79-82
Amrolcs chicken: **17**: 73-86
Ancona chicken: **24**: 41-48
Andalusia region: **10**: 35-45;
12: 89-95; **17**: 87-93; **19**: 69-89
Andalusian horse: **8**: 69-77;
10: 67-73; **22**: 43-52
Andhra Pradesh State: **4**: 27-29;
11: 57-60
Andropogon: **1**: 5-12; **9**: 37-43
Andropogon bicornis: **9**: 27-35
Andropogon gayanus: **17**: 127-145;
17: 147-164
Andropogon lateralis incanus:
6: 19-23; **7**: 11-18
Andropogon tectorum: **17**: 127-145
Anglo-arab horse: **16**: 71-74;
20: 35-54
Anglo-Norman horse: **4**: 19-25;
16: 71-74
Angora goat: **9**: 49-60; **19**: 57-68
Angus cow: **6**: 1-9
Anhui province: **15**: 63-73
Ankole cattle: **17**: 127-145
Aohan sheep: **9**: 49-60
Appaloosa horse: **3**: 35-38
Apullian highlands: **9**: 85-89
Apuileiese region: **10**: 67-73
Arab goat: **21**: 1-22
Arab horse: : **4**: 19-25; **8**: 69-77;
9: 85-89; **10**: 7-26; **16**: 65-70;
22: 29-42; **24**: 31-39
Arab zebu: **9**: 13-21
Arab region: **13**: 23-37; **17**: 127-145
Aragona region: **10**: 57-65
Arbutus unedo: **24**: 31-39
Argentine Creole cattle: **25**: 109-123
Aristida spp.: **1**: 5-12; **9**: 37-43
Arunachal Pradesh state: **12**: 59-67;
18: 41-48
Arundinella spp.: **20**: 9-15
Ascaris spp.: **20**: 35-54
Asian buffalo: **6**: 1-9
Asian zebu: **6**: 25-28
Assam Hill goat: **22**: 79-82
Assiut city: **13**: 47-58
Asturcon horse: **19**: 17-26
Asturia region: **7**: 29-33; **11**: 69-76;
11: 77-82; **22**: 29-42
Asturiana cattle:
Athlassa region: **4**: 1-9; **7**: 23-27
- Atlantic Pyrenees Department
(France): **22**: 1-5
Aurès mountains: **17**: 61-71
Australian Merino sheep: **9**: 49-60
Australian Poll Hereford cattle:
17: 87-93
Avena sativa: **17**: 101-113;
18: 49-57
Avena spp.: **1**: 5-12
Awassi sheep: **4**: 1-9; **7**: 23-27;
16: 37-45
Ayrshire cattle: **1**: 17-20; **6**: 25-28
Babesiosis disease: **17**: 147-164
Bachaur cattle: **22**: 79-82
Bagot goat: **4**: 31-36
Bahia State: **10**: 7-26
Baikal lake: **12**: 45-57
Bako Station: **1**: 5-12
Bakosi cattle: **13**: 23-37
Bakweri cattle: **13**: 23-37
Baladi rabbit: **23**: 49-67
Baladi sheep: **13**: 47-58
Balanites aegyptiaca: **18**: 29-39
Bale region: **8**: 79-82
Bambara cattle: **21**: 1-22
Bank of Genetic Variability: **5**: 1-5
Banyo Gudali cattle: **17**: 147-164
Baoulé cattle: **13**: 23-37
Barb horse: **8**: 69-77
Barbari goat: **22**: 79-82
Bargur cattle: **22**: 79-82
Barroso cattle: **7**: 11-18
Basque country: **8**: 27-33; **22**: 1-5
Batha goat: **21**: 1-22
Bati (Town): **1**: 5-12
Bauhima variegata: **19**: 37-43
B-cells: **23**: 1-11
Bean husk: **14**: 61-71
Beetal goat: **22**: 79-82
Beheira governorate: **13**: 47-58
Beishan goat: **9**: 49-60
Belgian Landrace pig: **16**: 37-45
Belted Galloway cattle: **4**: 31-36
Bengal region: **12**: 69-75
Benin: **14**: 13-25
Bergamasca sheep: **10**: 7-26
Berkshire pig: **4**: 31-36; **12**: 89-95;
19: 91-108
Berrenda cattle: **9**: 27-35
Bezoar wild goat: **8**: 35-41
Bhadwari cattle: **22**: 79-82
Bharatpur (Rajasthan): **1**: 29-30
Bhilwara district: **16**: 75-82
Bhiwani: **1**: 29-30
Bhutan: **19**: 37-43
Bhutia horse: **21**: 71-80
Biga village (Turkey): **14**: 33-39

¹Entries that were not the main concern of the article, but were significantly refereed. Entries represent different types of entries e.g.: breed/type, geographical region, production environment, etc.

²Citation: **issue number**: pages

- Bigawi chicken: **14:** 33-39
 Bikaner district: **22:** 19-27
 Bikaneri camel: **18:** 29-39
 Bikaneri sheep: **16:** 83-89
 Binghu water buffalo: **15:** 75-91
 Bison bonasus: **16:** 5-12
 Bizkaia area: **22:** 1-5
 Black and White cattle: **16:** 65-70;
18: 79-91
 Black and White-chested Danish
 duck: **7:** 45-53
 Black Bengal goat: **22:** 79-82
 Black disease: **8:** 35-41
 Blanco Orejimonos cattle (BOM):
9: 27-35; **24:** 1-11
 Blood antigen: **7:** 55-62
 Blue Albion cattle: **4:** 31-36
 Blue Vienna rabbit: **23:** 49-67
 Bohemian Pied Red cattle: **9:** 7-11
 Bohemian Red cattle: **9:** 7-11
 Bolivia: **14:** 91-98
 Bolivian Chaco forest: **7:** 7-10
 BON cattle: **9:** 27-35
 Bonsmara cattle: **20:** 17-26
 Boophilus microplus: **23:** 1-11
 Boran: **14:** 13-25
 Boran cattle: **6:** 25-28; **23:** 13-20
 Boran zebu: **7:** 55-62
 Bordaleiros sheep: **6:** 19-23
 Border Leicester sheep: **9:** 49-60
 Borgou: **14:** 13-25
 Borneo: **13:** 3-10
 Bos banteng: **16:** 29-35; **18:** 41-48
 Bos brachyceros africanus: **21:** 23-33
 Bos brachyceros: **7:** 71-76
 Bos desertorum: **21:** 23-33
 Bos frontalis: **18:** 41-48; **20:** 9-15;
20: 27-34
 Bos gauris: **20:** 27-34
 Bos grunniens: **12:** 45-57
 Bos indicus: **1:** 17-20; **4:** 27-29;
6: 25-28; **7:** 55-62; **9:** 37-43;
11: 57-60; **13:** 59-63; **15:** 3-22;
15: 75-91; **16:** 13-27; **16:** 47-63;
17: 147-164; **17:** 35-79;
20: 27-34; **21:** 35-42; **23:** 1-11; **25:**
 109-123
 Bos mulus: **19:** 27-36; **6:** 1-9
 Bos primegenius Hahni: **9:** 13-21
 Bos primigenius: **8:** 59-68; **14:** 61-71;
21: 49-60
 Bos primigenius estrepisicerus:
21: 23-33
 Bos silvestris: **8:** 59-68
 Bos taurus: **2:** 1-19; **6:** 1-9;
6: 25-28; **7:** 11-18; **7:** 55-62;
9: 27-35; **9:** 37-43; **12:** 45-57;
14: 41-59; **15:** 59-62; **15:** 75-91;
16: 13-27; **16:** 47-63; **17:** 35-79;
17: 127-145; **19:** 27-36;
21: 35-42; **23:** 1-11; **25:** 109-123
 Bos taurus Bolensz: **9:** 13-21
 Bos Taurus Brachyceros: **14:** 41-59;
17: 147-164
 Bos taurus ibericus: **21:** 23-33
 Bos taurus longifrons: **17:** 147-164
 Bos taurus primigenius: **16:** 5-12
 Bos taurus scythicus: **9:** 37-43
 Bos taurus typicus: **9:** 13-21
 Bos zebu: **14:** 61-71
 Bosnian mountains: **9:** 7-11
 Bougon region: **17:** 95-99
 Boukachmir community: **8:** 55-57
 Bouna region: **19:** 91-108
 Bourgogne rabbit: **23:** 49-67
 Bouscat Giant White rabbit:
23: 49-67
 Bovine spongiform encephalopathy
 (BSE) disease: **23:** 1-11
 Brachiaria mutica: **24:** 1-11
 Brachiaria ruzziziensis: **20:** 55-69
 Brachiaria spp.: **17:** 147-164
 Brachiarias decumbens: **24:** 1-11
 Brachiarias dictyoneura: **24:** 1-11
 Brachiarias spp.: **24:** 1-11
 Brachyceros shorthorn cattle:
17: 147-164
 Brahman cattle: **2:** 1-19;
17: 127-145; **20:** 17-26;
23: 1-11
 Brahman zebu: **13:** 23-37
 Brakna region: **15:** 3-22
 Brasil: **13:** 3-10
 Brassaiopsis mitis: **19:** 37-43
 Brassica campestris: **18:** 49-57
 Brassica spp.: **17:** 101-113
 Bravo cattle: **7:** 11-18
 Brindisi city: **9:** 85-89
 British Lop pig: **4:** 31-36
 British Saanen goat: **9:** 49-60
 British Saddleback pig: **4:** 31-36
 British Welsh Black cattle: **17:** 87-93
 British White bull: **4:** 31-36
 Brittany: **9:** 61-67
 Briza spp.: **9:** 37-43
 Bromus spp.: **9:** 37-43
 Brown bee: **7:** 45-53
 Brown Swiss cattle: **1:** 21-24;
3: 21-32; **7:** 7-10; **7:** 7-10;
10: 7-26; **11:** 61-67
 Brucellosis: **8:** 35-41; **23:** 1-11
 Brune de l'Atlas cattle: **8:** 49-54
 Brune des Alpes cattle: **14:** 13-25
 Bubalos Ameer: **14:** 41-59
 Bubalus bubalis: **12:** 45-57;
19: 7-15
 Buckfast bee: **7:** 45-53
 Buje (Roman) cattle: **7:** 77-82
 Burgos: **8:** 13-26; **19:** 17-26
 Burmese Red: **3:** 17-20
 Busa cattle: **16:** 37-45
 Bush ticks disease: **23:** 1-11
 Cakiel sheep: **9:** 7-11
 Calafate: **9:** 23-26
 Calamagrostis spp.: **9:** 37-43
 Calcutta: **12:** 69-75
 Californian Gray chicken: **17:** 73-86
 Californian rabbit: **23:** 49-67
 Calipuy guanaco reserve (Peru):
14: 91-98
 Camelidae: **14:** 91-98
 Camelus bactrianus: **14:** 91-98
 Camelus dromadairus: **15:** 3-22;
10: 47-56; **14:** 91-98; **18:** 29-39
 Cameroon: **9:** 13-21; **13:** 3-10;
17: 127-145; **17:** 147-164
 Campania region: **12:** 25-43
 Campos Novos region: **9:** 37-43
 Cantabria region: **22:** 29-42
 Cantonese breed: **3:** 17-20
 Cape Toi National Park: **13:** 11-22
 Capparis decidua: **18:** 29-39
 Capra aegagrus hircus: **11:** 41-55
 Capra genus: **7:** 23-27
 Capra hircus: **15:** 3-22; **17:** 61-71
 Capra hircus girgentana: **11:** 41-55
 Capra hircus hircus: **11:** 41-55
 Capra prisca: **11:** 41-55
 Caprine pleuropneumonia: **8:** 35-41
 Caracu cattle: **25:** 109-123
 Carmagnola Grey: **23:** 49-67
 Carpathian Basin: **6:** 29-34;
13: 75-82; **21:** 49-60
 Carpathian goat: **9:** 7-11
 Carpathian region: **11:** 35-40
 Carso region: **10:** 67-73
 Casaldianni: **12:** 25-43
 Casamance area: **24:** 63-69
 Casanare cattle: **9:** 27-35
 Casein: **25:** 57-71
 Cashmere goat: **19:** 27-36
 Cashmere wool: **9:** 49-60
 Caspian horse: **4:** 31-36
 Cassava: **16:** 29-35
 Castille region: **17:** 115-125
 Castlemilk Moorit: **4:** 31-36
 Catalase: **19:** 69-89
 Catalogue of Poultry Stocks held at
 Research and Teaching
 Institutions in Canada: **1:** 27-28
 Cauca river: **9:** 27-35
 Caucasus: **6:** 29-34
 Ceara State: **10:** 7-26
 Celtic ponies: **22:** 29-42
 CENARGEN: **10:** 7-26
 Cenchrus ciliaris: **6:** 25-28
 Central Africa: **1:** 17-20

- Central Italy: **8**: 59-68; **8**: 69-77
 Central Senegal: **24**: 63-69
 Cestoda: **24**: 13-30
 Cévennes (National Park of):
1: 25-26
 Chad lake: **9**: 13-21; **17**: 127-145
 Changthang goat: **22**: 79-82
 Changthangi sheep: **22**: 79-82
 Charolais cattle: **2**: 1-19; **7**: 29-33;
7: 71-76; **8**: 59-68; **9**: 27-35;
10: 7-26; **12**: 45-57; **17**: 51-60; **25**:
 45-55
 Charollais sheep: **9**: 49-60
 Chegu goat: **22**: 79-82
 Chengdu brown goat: **15**: 63-73
 Chianina cattle: **8**: 59-68
 Chianina/Piemontese bull: **4**: 27-29
 Chickpea: **18**: 29-39
 Chifeng (Inner Mongolia): **9**: 45-47
 Chillingham cattle: **4**: 31-36;
6: 1-9
 Chillingham Park cattle: **5**: 9-25
 Chiloé island: **23**: 41-47
 China: **12**: 59-67
 China tropical-subtropical area:
16: 29-35
 Chinchilla rabbit: **23**: 49-67
 Chinese Merino sheep: **9**: 49-60
 Chinese new Merino sheep: **9**: 49-60
 Chino Santandereano cattle:
9: 27-35
 Chios sheep: **7**: 23-27; **23**: 27-39
 Chito dry meat: **21**: 61-70
 Chittorgarh district: **16**: 75-82;
16: 83-89
 Chloris: **6**: 25-28
 Chloris spp.: **9**: 37-43
 Chokla sheep: **22**: 79-82
 Choluteca, Honduras: **4**: 15-17
 Chubotka Crested hen: **9**: 7-11
 Chugoku district: **18**: 65-78
 Chuiquimulilla: **4**: 15-17
 Churra sheep: **6**: 19-23; **10**: 7-26;
17: 115-125
 Churu area: **22**: 19-27
 Cicaria region: **7**: 77-82
 Cicer arietinum: **17**: 101-113
 Cigaja sheep: **9**: 7-11
 Circello: **12**: 25-43
 Cistaceae spp.: **24**: 31-39
 Cleveland Bay horse: **4**: 31-36
 Cloris spp.: **1**: 5-12
 Clydesdale horse: **4**: 31-36
 Coastal Kenya: **24**: 13-30
 Coccidiosis disease: **22**: 61-69
 Colored Bohemian goat: **9**: 7-11
 Commission on Genetic Resources
 for Food and Agriculture:
25: 33-43
 Commission on Plant Genetic
 Resources: **25**: 33-43
 Compositae familia: **9**: 37-43
 Comunidad Autónoma Vasca:
8: 27-33
 Conniphora spp.: **1**: 5-12
 ConSDABI: **12**: 25-43
 Convention on Biological Diversity:
9: 3-6; **25**: 33-43; **25**: 79-99
 Conversano horse family: **9**: 85-89;
10: 67-73
 Cooperia parasite: **23**: 1-11
 Cordillera Cantabrica mountains:
11: 69-76
 Corriedale sheep: **9**: 49-60
 Corsican pony: **24**: 31-39
 Corumba city: **10**: 7-26
 Costeño cattle: **9**: 27-35
 Costeño criollo cattle: **24**: 1-11
 Cotswold sheep: **4**: 31-36
 Cou Nu du Forez chicken: **25**: 73-78
 Cowdriosis disease: **17**: 147-164
 CPAP: **10**: 7-26
 Cracov (Poland): **13**: 3-10
 Creole rabbit: **23**: 49-67
 Criollo rabbit: **23**: 49-67
 Criollo Brazilian cattle: **10**: 7-26; **25**:
 109-123
 Criollo Venezuelan cattle: **7**: 11-18
 Croatia cattle: **7**: 77-82
 Curly chicken: **24**: 63-69
 Cyamopsis tetraganodora: **18**: 49-57
 Cynodon nlemfuensis: **4**: 15-17
 Cynodon spp.: **1**: 5-12
 Cyperaceae familia: **9**: 37-43
 Cyprus: **8**: 13-26
 Cyprus fat-tailed sheep: **7**: 23-27
 Czech Republic: **9**: 7-11
 Dahra mountains: **17**: 61-71
 Daiyunshan goat: **9**: 49-60
 Dala sheep: **7**: 45-53
 Dales Pony horse: **4**: 31-36
 Dandarawi chicken: **14**: 33-39
 Dangi cattle: **22**: 79-82
 Danish Black Pied cattle: **7**: 45-53
 Danish horse: **16**: 65-70
 Danish Land hen: **7**: 45-53
 Danish Red cattle: **7**: 45-53; **7**: 71-76
 Danthonia spp.: **9**: 37-43
 Dar-el-Ramad village: **14**: 33-39
 Dartmoor pony: **22**: 29-42;
24: 31-39
 Dawara zebu: **7**: 55-62
 Deccan plateau: **12**: 77-87
 Deccani goat: **22**: 79-82
 Deccani sheep: **22**: 79-82
 Denmark: **2**: 20-26
 Deoni cattle: **22**: 79-82
 Dermatobia hominis: **2**: 1-19;
9: 27-35
 Dermatophila congolense:
17: 147-164
 Desmodium spp.: **24**: 1-11
 Dexter cattle: **4**: 31-36
 Dian-nan small-ear pig: **20**: 3-8
 Dichantium aristatum: **24**: 1-11
 Dicrostakis gomerata: **19**: 91-108
 Diffa Department: **9**: 13-21
 Digitaria decumbens: **24**: 1-11
 Digitaria spp.: **9**: 37-43
 Dinghai county: **16**: 29-35
 Diougray goat: **21**: 1-22
 Dire Dawa: **1**: 5-12
 Djafoun type cattle: **16**: 47-63
 Djakoré cattle: **21**: 1-22
 DNA storage: **9**: 3-6
 Dongsan Township: **7**: 19-22
 Dosanko pony: **13**: 11-22
 Dubrovnik sheep: **11**: 25-33
 Duroc pig: **12**: 89-95; **16**: 37-45
 Duroc-Jersey pig: **12**: 89-95
 Dynevor cattle: **5**: 1-5;
 Dynevor Raven cattle: **25**: 45-55
 Dzo: **12**: 59-67
 Dzo cattle hybrid: **19**: 27-36
 Dzomo cattle hybrid: **12**: 59-67;
19: 27-36
 EAAP-Animal Genetic Data Bank:
25: 79-99
 East Africa: **15**: 23-42
 East African Small Sheep: **20**: 55-69
 East Europe: **6**: 29-34
 East Friesian sheep: **4**: 1-9
 East Rajasthan region: **16**: 75-82
 Eastern Nepal region: **22**: 15-18
 Ebro river: **10**: 57-65
 Echinocloa polystachia: **4**: 15-17
 Ectoparasites: **3**: 1-9
 Egiptian longhorn cattle:
17: 147-164
 Ehime prefecture: **13**: 11-22
 EMBRAPA: **10**: 7-26; **21**: 43-48; **25**:
 109-123
 Emilia Romagna: **8**: 13-26
 Endocrinology: **8**: 43-47
 English Thoroughbred stallion:
4: 19-25
 Enteropogon: **1**: 5-12
 Equus asinus: **15**: 3-22
 Equus caballus: **15**: 3-22
 Equus ferus: **6**: 29-34
 Equus gmelini: **19**: 45-56
 Equus gracilis: **22**: 29-42
 Equus tarpan: **19**: 45-56
 Eragrostis spp.: **9**: 37-43
 Erduos sheep: **9**: 49-60
 Erianthus longisetosus: **20**: 9-15
 Erica: **1**: 5-12
 Estonian Red cattle: **16**: 65-70
 Etruscans: **8**: 69-77
 European Brown Alpine cattle:
1: 21-24
 European dairy breeds: **3**: 21-32;
7: 7-10
 Exmoor pony: **22**: 29-42
 Extremadura region: **12**: 89-95
 Facial eczema disease: **23**: 1-11

- Faco horse: **19:** 17-26
 FAO Global Data Bank for Domestic Livestock: **11:** 13-20
 Fasciola gigantica: **24:** 13-30
 Fasciola hepatica: **24:** 13-30
 Favory horse family: **10:** 67-73
 Fayoumi Province: **14:** 33-39
 Fell pony: **22:** 29-42; **4:** 31-36
 Fescue toxicosis disease: **23:** 1-11
 Festuca arundinacea: **23:** 1-11
 Ficus curia: **19:** 37-43
 Fighting bull: **17:** 87-93
 Finés sheep: **10:** 57-65
 Finland: **2:** 20-26
 Finn sheep: **17:** 115-125
 Finnish Ayrshire cattle: **7:** 45-53
 Flamande: **9:** 61-67
 Fleckvieh cattle: **19:** 91-108
 Fleischschaf sheep: **10:** 57-65
 Flemish Giant rabbit: **23:** 49-67
 Fleuve horse: **20:** 35-54
 Foamy (pasture) bloat disease: **23:** 1-11
 Forest grasses: **18:** 41-48
 Forest reindeer: **7:** 45-53
 Foulbé zebu: **13:** 23-37
 Foutanké horse: **20:** 35-54
 France (Southern): **8:** 13-26
 French Pyrénées: **13:** 65-73
 French Wedder rabbit: **7:** 45-53
 Friesian cattle: **7:** 71-76; **23:** 1-11
 Fucus serratus: **6:** 1-9
 Fugin goat: **9:** 49-60
 Fuhai Big-tail sheep: **9:** 49-60
 Fulani cattle: **21:** 1-22; **21:** 1-22
 Fulani Gudali cattle: **17:** 147-164
 Fulani zebu: **13:** 39-45
 Fyparrhenia spp.: **1:** 5-12
 Gaddi goat: **19:** 27-36; **22:** 79-82
 Gaddi sheep: **22:** 79-82
 Galao cattle: **22:** 79-82
 Galicia region: **19:** 45-56; **21:** 23-33
 Galician cattle: **2:** 1-19
 Gallic thoroughbre horse: **13:** 65-73
 Gallus gallus: **15:** 3-22
 Gallus gallus domesticus: **22:** 61-69
 Gambia: **14:** 13-25
 Gamo Gofa region: **8:** 79-82
 Ganjia sheep: **9:** 49-60
 Ganqin sheep: **9:** 49-60
 Gansu Alpine sheep: **9:** 49-60
 Gansu Province: **9:** 49-60
 Gargano promontory: **11:** 41-55
 Garrano horse: **19:** 17-26; **22:** 29-42
 Gasara zebu: **7:** 55-62
 Gastrointestinal (GI) nematode parasites: **24:** 13-30
 Gaur cattle: **18:** 41-48; **20:** 27-34
 Gazal horse: **4:** 19-25
 Gemu Gofa (Province of): **1:** 5-12
 Gene pool: **4:** 31-36
 Georgian Brachyceros: **14:** 41-59
 German landrace pig: **16:** 37-45
 Gerola valley: **11:** 41-55
 Gharpala sheep: **22:** 15-18
 Giara plateau: **24:** 31-39
 Ginger: **19:** 37-43
 Gir cattle: **2:** 1-19; **22:** 79-82
 Giza rabbit: **23:** 49-67
 Gliricida sepium: **24:** 1-11
 Global Strategy for the Management of Farm Animal Genetic Resources: **25:** 33-43
 Glossina longipalpis: **17:** 127-145
 Glossina morsitans: **17:** 147-164
 Glossina palpalis: **17:** 147-164
 Glossina tachinoide: **17:** 147-164
 Gloucester cattle: **4:** 31-36
 Gloucester Old Spots pig: **4:** 31-36
 Glucose phosphate isomerase: **22:** 29-42
 Gobi desert: **19:** 27-36
 Gobra cattle: **17:** 127-145
 Gobra zebu: **19:** 91-108; **21:** 1-22
 Godavari District: **4:** 27-29
 Gojam region: **8:** 79-82
 Golden Guernsey goat: **4:** 31-36
 Gongola State: **3:** 1-9
 Gorica cattle: **7:** 77-82
 Goto island: **13:** 11-22
 Goudali zebu: **13:** 23-37; **21:** 1-22
 Gramineae spp.: **15:** 75-91
 Granadina goat: **19:** 69-89
 Grassland Red cattle: **9:** 45-47
 Greek mountain pony: **16:** 71-74
 Grey Danish goose: **7:** 45-53
 Grey Stepland cattle: **14:** 41-59
 Grey Steppe cattle: **16:** 37-45
 Grey-spotted Danish goose: **7:** 45-53
 Grey-spotted Tame goose: **7:** 45-53
 Guadyerbas pig: **12:** 89-95
 Guangdong province: **15:** 75-91
 Guangxi province: **18:** 23-27
 Guanzhong milk goat: **9:** 49-60
 Gudali cattle: **17:** 127-145
 Gudali zebu: **13:** 39-45
 Guéra goat: **21:** 1-22
 Guidimakha region: **15:** 3-22
 Guipuzcoa: **8:** 13-26
 Guizhou province: **18:** 23-27
 Gujarat State: **10:** 47-56; **18:** 29-39; **21:** 71-80
 Gulf of Mexico: **23:** 69-78
 Gumboro disease: **22:** 61-69
 Haemaphysalis longicornis: **23:** 1-11
 Haemoglobin: **19:** 69-89
 Haemonchus contortus: **24:** 13-30
 Haemonchus parasite: **23:** 1-11
 Haemonchus spp.: **24:** 13-30
 Hainan Island: **16:** 29-35
 Haiyuan of Ningxia region: **9:** 49-60
 Haldar community: **12:** 69-75
 Hallikar cattle: **22:** 79-82
 Hamitic cattle: **17:** 147-164
 Hamitic Longhorn cattle: **17:** 127-145
 Hampshire pig: **7:** 63-70; **16:** 37-45
 Hardveld ecological zone: **20:** 17-26
 Hardy-Weinberg equilibrium: **19:** 69-89
 Hariana cattle: **22:** 79-82
 Harungana madagascariensis: **19:** 91-108
 Haryana State: **10:** 47-56; **18:** 29-39; **25:** 27-31
 Hasake sheep: **9:** 49-60
 Hassan sheep: **22:** 79-82
 Hawaii sheep: **5:** 9-25
 Hebridean sheep: **4:** 31-36
 Helminth parasites: **24:** 13-30
 Helminthiasis: **3:** 1-9
 Hereford cattle: **23:** 1-11; **23:** 1-11
 Hereford-like cattle: **2:** 1-19
 Hetian sheep: **9:** 49-60
 Hexi Cashmere goat: **9:** 49-60
 Highland Scots pony: **22:** 29-42
 Himachal Pradesh state: **12:** 59-67
 Himalayan countries: **12:** 59-67
 Himalayan states of India: **12:** 59-67
 Himalayas: **20:** 27-34
 Hiparrhenia rufa: **24:** 1-11
 Hirute: **1:** 5-12
 Hissar (Haryana): **1:** 29-30
 Hissardale sheep: **22:** 79-82
 Histomonas gallinarum: **23:** 69-78
 Histomonas meleagridis: **23:** 69-78
 Hodeidah's province: **8:** 35-41
 Hoislein cattle: **2:** 1-19
 Hokkaido island: **13:** 11-22
 Hokkaido region: **18:** 65-78
 Holstein cattle: **2:** 1-19; **3:** 21-32; **4:** 27-29; **7:** 7-10; **10:** 7-26; **13:** 23-37; **14:** 13-25; **18:** 65-78; **18:** 79-91; **19:** 27-36
 Holstein-Friesian cattle: **7:** 29-33; **8:** 43-47; **11:** 61-67; **17:** 127-145; **23:** 27-39
 Homo sapiens: **16:** 5-12
 Hongtong goat: **9:** 49-60
 Honshu Island: **18:** 65-78
 Hordeum vulgare: **17:** 101-113
 Horn of Africa: **17:** 127-145; **17:** 147-164
 Hortobagy Racka sheep: **11:** 35-40; **11:** 35-40
 Huacaya alpaca: **14:** 91-98

- Huanghe River Valley: **9**: 49-60
Huangyan country: **9**: 45-47
Hubei Province: **15**: 63-73
Humpless longhorn cattle: **9**: 13-21
Hunan Province: **15**: 63-73
Hungarian Podolic cattle: **7**: 77-82
Hungarian White/Silver cattle: **21**: 49-60
Hutsul horse: **9**: 7-11
Huyang sheep: **9**: 49-60
Hybro broiler feathered neck fowl: **7**: 83-88
Hyparrenhia rufa: **9**: 27-35
Hyparrhenia spp.: **17**: 147-164
Iberian cattle: **17**: 87-93
ICAR: **17**: 101-113
Icelandic pony: **22**: 29-42
IGF-I: **8**: 43-47
Ile de France sheep: **10**: 57-65
Ileia horse: **16**: 71-74
Ilubabor Province: **1**: 5-12
Imabari city: **13**: 11-22
Imperata cylindrica: **20**: 9-15
Indian bison: **18**: 41-48
Indo-Gangetic Plains: **1**: 29-30
Internal parasites: **23**: 1-11
International Registry on Poultry Genetic Stocks: **1**: 27-28
Iowa State (USA): **14**: 33-39
Irish Connemara pony: **22**: 29-42
Irish Moyled cattle: **4**: 31-36
Irish pony: **24**: 31-39
Hispaniola island: **25**: 109-123
Istrian peninsula: **7**: 77-82; **16**: 37-45
Istrian Karst breed: **7**: 77-82
Istrian Podolic cattle: **7**: 77-82
Italian saddle horse: **8**: 69-77
Italy (Central-Northern): **8**: 13-26
Jaca horse: **19**: 17-26
Jaffarabadi cattle: **22**: 79-82
Jaipur district: **22**: 19-27
Jaisalmeri camel: **18**: 29-39
Jaisalmeri sheep: **22**: 79-82
Jalori camel: **18**: 29-39
Jammu state: **12**: 59-67
Jamon: **12**: 89-95
Jamunaparii goat: **22**: 79-82
Japanese Large-eared rabbit: **23**: 49-67
Jatsa cattle: **18**: 41-48
Jatsamin cattle: **18**: 41-48
Jersey bull: **7**: 71-76
Jersey cattle: **2**: 1-19; **4**: 27-29; **11**: 61-67; **13**: 23-37; **14**: 13-25; **19**: 27-36; **19**: 37-43
Jersiais cattle: **21**: 1-22
Jhunjhunu area: **22**: 19-27
Jiangsu Province: **7**: 19-22; **15**: 75-91; **20**: 3-8
Jujuy province: **18**: 7-21
Jungle Fowl: **3**: 17-20
Junin sheep: **17**: 115-125
Kaarta cattle: **21**: 1-22
Kacchi camel: **18**: 29-39
Kaduna State: **3**: 1-9
Kaffa Province: **1**: 5-12
Kagoshima prefecture: **13**: 11-22
Kanem goat: **21**: 1-22
Kangayam cattle: **22**: 79-82
Kangra goat: **19**: 27-36
Kankre cattle: **22**: 79-82
Kano State: **3**: 1-9
Karamajong zebu type: **15**: 23-42
Kargil district: **19**: 27-36
Karnah sheep: **22**: 79-82
Karnal: **1**: 29-30
Kashmir sheep: **22**: 79-82
Kashmir state: **12**: 59-67
Kenana cattle: **6**: 25-28
Kenkatha cattle: **22**: 79-82
Kenya: **14**: 13-25
Kenya Milk Records (KMR): **8**: 49-54
Kenyan White rabbit: **23**: 49-67
Kerkoub muton: **6**: 11-13
Kerqin sheep: **9**: 49-60
Kerry cattle: **4**: 31-36
Ketosis disease: **23**: 1-11
Kgalagadi sand: **20**: 17-26
Khemisset (Province of): **8**: 55-57
Kherigarh cattle: **22**: 79-82
Khillari cattle: **22**: 79-82
Kigezi region: **15**: 23-42
Kirdi cattle: **13**: 23-37
Kirdi sheep: **21**: 1-22
Kirgiz chicken: **17**: 73-86
Kiso river: **13**: 11-22
Kladruby horse: **9**: 7-11
Kobe-beef: **18**: 65-78
Kochi cattle strain: **18**: 65-78
Kochi prefecture: **13**: 11-22
Kohafa village: **14**: 33-39
Kohare rabbit: **7**: 45-53
Konik horse: **9**: 7-11
Konnai Adu goat: **22**: 79-82
Korhogo pig: **21**: 1-22
Kraina region: **14**: 27-32
Krainger bee: **7**: 45-53
Kranjska cattle: **7**: 77-82
Krishna river: **4**: 27-29
Krishna Valley cattle: **22**: 79-82
Kumamoto cattle strain: **18**: 65-78
Kuri cattle: **17**: 127-145
Kutchi camel: **18**: 29-39
Kymi sheep: **9**: 69-76
Kyoga zebu type: **15**: 23-42
La Bresse chicken: **24**: 41-48
La Española island: **22**: 43-52
Lacaune sheep: **10**: 57-65
Ladakh plateau: **19**: 27-36
Lagunaire cattle: **13**: 39-45
Lake Rudolph: **1**: 5-12
Lama glama (llama): **14**: 91-98
Lama guanicoe: **14**: 91-98
Lama pacos: **14**: 91-98
Landes de Bretagne ram: **9**: 61-67
Landes de Gascogne (Regional Park of): **1**: 25-26
Landrace pigs: **7**: 45-53
Lankan buffalo: **25**: 33-43
Laoshan goat breed: **9**: 49-60
Large Black pig: **4**: 31-36; **12**: 89-95
Large White pig: **7**: 63-70
Large-eared rabbit: **23**: 49-67
Latium (Northern) Region: **8**: 59-68
Latxa Cara Negra sheep: **8**: 27-33
Latxa Cara Rubia sheep: **8**: 27-33
Latxa sheep: **17**: 115-125
Laze polyantha: **19**: 37-43
Leghorn chicken: **24**: 41-48
Leghorn White chicken: **17**: 73-86
Leguminosae familia: **9**: 37-43
Leicester Longwool sheep: **4**: 31-36
Leicester sheep: **7**: 45-53
Leicester/Dishley sheep: **11**: 25-33
Lepini Mountains: **8**: 59-68
Lequing country: **9**: 45-47
Leucosis disease resistance: **14**: 33-39
Leukosis disease: **23**: 1-11
Lialgshan county: **16**: 29-35
Liaoning Cashmere goat: **9**: 49-60
Liguria: **8**: 13-26
Limusin cattle: **7**: 29-33; **25**: 45-55
Lincoln Longwool sheep: **4**: 31-36
Lincoln sheep: **9**: 49-60
Lipizza: **10**: 67-73
Lobelia: **1**: 5-12
Lohi goat: **22**: 79-82
Lohi sheep: **25**: 27-31
Lolium perenne L.: **23**: 1-11
Longhorned zebu: **17**: 127-145
Lonjsko Polje Natural Park: **14**: 27-32
Los Glaciares National Park: **9**: 23-26; **15**: 49-57
Loudetia arundinacea: **17**: 147-164
Luberon (Regional Park of): **1**: 25-26
Lugo Provence: **14**: 73-78
Lute hen: **7**: 45-53
Lymphoid leucosis disease: **22**: 61-69
M'Bororo zebu: **13**: 23-37
Macedonia region: **14**: 79-90
Macina goat: **21**: 1-22
Madagascar zebu: **17**: 127-145
Madhya Pradesh State: **17**: 101-113; **18**: 29-39
Madras Red sheep: **22**: 79-82
Maestoso horse family: **10**: 67-73
Magnisia region: **9**: 69-76
Magra sheep: **22**: 79-82
Maharashtra state: **12**: 77-87
Mahel Meda: **1**: 5-12
Mahendragarh: **1**: 29-30

- Maize: **19:** 37-43
Malabari goat: **22:** 79-82
Malawi: **14:** 13-25
Mali: **14:** 13-25; **17:** 147-164
Malpura sheep: **22:** 79-82
Maltese goat: **11:** 41-55; **17:** 61-71
Malvi cattle: **22:** 79-82
Malay Game chicken: **25:** 73-78
Malwa (Malva) region: **18:** 29-39
Manchega sheep: **10:** 57-65
Mandara mountain: **13:** 23-37;
16: 47-63
Mandya sheep: **22:** 79-82
Manipur State: **18:** 41-48
Manipuri horse: **21:** 71-80
Mantiqueira cattle: **25:** 109-123
Manx Loghtan sheep: **4:** 31-36
Maradi goat: **21:** 1-22
Marais Poitevin (Regional Park of):
1: 25-26
Marchigiana cattle: **8:** 59-68
Mareb region: **8:** 35-41
Marek's disease: **22:** 61-69
Maremna region: **8:** 59-68; **8:** 69-77
Maremmana cattle: **7:** 77-82
Mariadvori cattle: **7:** 77-82
Marwari camel: **18:** 29-39
Marwari goat: **22:** 79-82
Marwari horse: **21:** 71-80
Marwari sheep: **16:** 75-82; **22:** 79-82
Massakori goat: **21:** 1-22
Mastitis disease: **23:** 1-11
Mato Grosso do Sul: **10:** 7-26;
21: 43-48; **25:** 109-123
Matsuzaka-beef: **18:** 65-78
Mauna Loa sheep: **5:** 9-25
Maure sheep: **21:** 1-22
Mawanga chicken: **22:** 61-69
Mayo Kebbi goat: **21:** 1-22
Mayo-Tsanaga department:
13: 23-37
Mbarara area: **15:** 23-42
Mbororo zebu: **13:** 39-45
Mbuuyé cattle: **13:** 23-37
Medicago sativa: **17:** 101-113
Mediterranean buffalo: **14:** 79-90
Mediterranean Pig Network:
12: 25-43
Mediterranean region: **12:** 25-43
Mediterranean sea coast: **13:** 47-58
Mehsana cattle: **22:** 79-82
Melanin pigments: **14:** 33-39
Meleagris gallopavo gallopavo:
23: 69-78
Meleagris gallopavo intermedia:
23: 69-78
Melica spp.: **9:** 37-43
Melka Werer: **1:** 5-12
Menshat Abdalla village: **14:** 33-39
Merens horse: **19:** 17-26
Merino sheep: **10:** 57-65; **11:** 35-40;
16: 37-45; **17:** 115-125;
19: 69-89; **22:** 79-82
Merinos stavisfol sheep: **11:** 25-33
Merinos turc sheep: **11:** 25-33
Mersuch horse: **4:** 19-25
Mesopotamia: **6:** 29-34
Mewari camel: **18:** 29-39
Mewati (Kosi) cattle: **22:** 79-82
Mewati camel: **18:** 29-39
Mezohegyes horse: **4:** 19-25
Middle Atlas: **8:** 55-57
Middle Awash Valley (Eastern
Ethiopia): **1:** 5-12
Midi-Pyrénées Region: **10:** 27-33
Millet: **19:** 37-43
Mimosa pigra: **19:** 91-108
Minas Gerais State: **10:** 7-26; **25:** 109-
123
Mithun cattle: **20:** 27-34
Miyako island: **13:** 11-22
Mizoram State: **18:** 41-48
Mmeta khosi chicken: **22:** 61-69
MoDAD: **25:** 33-43
Mogadishu: **7:** 55-62
Monastiri region: **22:** 7-13
Monbeliard cattle: **13:** 23-37;
14: 13-25; **18:** 79-91; **21:** 1-22
Mongolia: **12:** 59-67
Mongolian type cattle: **9:** 45-47
Montecristo island: **11:** 41-55
Montemaggiore: **10:** 67-73
Montgomery District of Pakistan:
1: 13-16
Moors pony: **24:** 31-39
Moravia: **6:** 29-34
Moravian Red cattle: **9:** 7-11
Moroccan Atlantic Coast: **6:** 11-13
Mossi sheep: **21:** 1-22
Mountain Abudemade: **1:** 5-12
Moxoto valley: **10:** 7-26
Mpar pony: **20:** 35-54
Mubende region: **15:** 23-42
Muffah group of buffalo: **14:** 79-90
Multibreed trial: **4:** 31-36
Murge region: **9:** 85-89
Murrah buffalo: **15:** 75-91;
18: 49-57; **25:** 33-43
Mutton type sheep: **25:** 27-31
Muturu cattle: **13:** 39-45;
17: 127-145
Muzzafarnagri sheep: **22:** 79-82
Mycoplasmosis: **3:** 1-9
Mycotoxic diseases: **23:** 1-11
Myrtus communis: **24:** 31-39
N'Dama cattle: **13:** 23-37; **13:** 39-45;
14: 13-25; **17:** 127-145;
21: 1-22
Nagaland State: **18:** 41-48
Nagori cattle: **22:** 79-82
Nagoya breed: **3:** 17-20
Nali sheep: **22:** 79-82; **25:** 27-31
Nan-yang cattle: **20:** 3-8
Napolitano horse: **10:** 67-73
Narougor horse: **20:** 35-54
National Committee of Danish
Cattle Husbandry: **25:** 101-108
National Council on Gene
Resources: **3:** 33-34
National Dairy Cattle Breeding
Program (NDCBP): **8:** 49-54
National Park of Hortobágy:
13: 75-82
Navarra: **8:** 13-26
Neimenggu Cashmere goat: **9:** 49-60
Neimenggu Region: **9:** 49-60
Neimenggu sheep: **9:** 49-60
Nellor sheep: **22:** 79-82
Nellore cattle: **10:** 7-26; **25:** 109-123
Nellore District: **4:** 27-29
Nematoda: **24:** 13-30
Nematodirus spp.: **24:** 13-30
Neolithic period: **6:** 29-34
Nepal: **12:** 45-57
Netherland Dwarf rabbit: **23:** 49-67
New Forest pony: **22:** 29-42
New Hampshire chicken: **14:** 33-39
New Hampshire hen: **7:** 45-53
New Zealand: **23:** 1-11
New Zealand White rabbit:
23: 49-67
Newcastle disease: **22:** 61-69
Nganda cattle: **15:** 23-42
Nganda cattle: **17:** 127-145
Ngaundere Gudali cattle:
17: 147-164
Niaux caves: **13:** 65-73
Nicosia: **4:** 1-9
Niger: **9:** 13-21
Nigeria: **9:** 13-21; **17:** 127-145;
17: 147-164
Nile valley: **9:** 13-21; **13:** 47-58
Nilgiri sheep: **22:** 79-82
Nili-Ravi buffalo: **15:** 75-91;
18: 49-57
Nili-Ravi cattle: **22:** 79-82
Nimari cattle: **22:** 79-82
Ningxia area: **20:** 3-8
Ningxia Hui Autonomous Region:
9: 49-60
Ninh Thuan Province: **11:** 21-24
Nordic Animal-Gene Banks
(NAGBs): **7:** 45-53
Nordic cattle breeds: **7:** 45-53
Nordic horse breeds: **7:** 45-53
Norfolk Horn cattle: **4:** 31-36
Norfolk Horn sheep: **4:** 31-36

- North Africa: **15**: 43-78
 North America: **1**: 21-24
 North eastern region of India: **18**: 41-48
 North India: **18**: 49-57
 North Ronaldsay sheep: **4**: 31-36
 Northeast China sheep: **9**: 49-60
 North-eastern India: **20**: 9-15
 Northern Burma: **20**: 9-15
 Northern coast of Colombia: **9**: 77-83
 Northern Colombia: **24**: 1-11
 Northern Spain: **8**: 13-26
 Norway: **2**: 20-26
 Norwegian pony: **24**: 31-39
 Norwegian Red cattle: **7**: 45-53
 Nothofagus betuloides: **9**: 23-26
 Nothofagus pumpilio: **9**: 23-26
 Notro: **9**: 23-26
 Nouakchott-Rosso chicken: **15**: 3-22
 Nova Gradiska: **14**: 27-32
 Nubin buck goat: **15**: 63-73
 Oaxaca city: **23**: 69-78
 Oaxaca State: **21**: 61-70
 Oesophagostomum spp.: **24**: 13-30
 Ogaden: **1**: 5-12
 Oland goose: **7**: 45-53
 Olaroz-Cauchari Park: **18**: 7-21
 Oldenburg/Ostfriesland region: **14**: 99-105
 Ongole cattle: **22**: 79-82
 Orinoco river: **9**: 27-35
 Orites region: **4**: 1-9
 Orkney Islands: **6**: 1-9
 Orlov horse: **16**: 65-70
 Osmanabadi goat: **22**: 79-82
 Ossimi valley and delta: **13**: 47-58
 Ostertagia spp.: **24**: 13-30
 Oulmès community: **8**: 55-57
 Overo allele: **3**: 35-38
 Ovis ammon Arkal: **13**: 75-82
 Ovis aries: **7**: 23-27; **15**: 3-22
 Ovis aries ligeriensis: **10**: 57-65
 Ovis musimon: **7**: 23-27
 Ovis orientalis: **7**: 23-27
 Oxford Down sheep: **4**: 31-36; **7**: 45-53
 Pahari camel: **10**: 47-56
 Paint horse: **3**: 35-38
 Palencia: **8**: 13-26
 Palomino horse: **3**: 35-38
 Pantaneiro cattle: **25**: 109-123
 Pamir: **12**: 45-57
 Pampas of Rio Grande do Sul: **25**: 109-123
 Panicum maximum: **6**: 25-28; **17**: 127-145; **24**: 1-11
 Panicum phragmitoides: **17**: 147-164
 Panicum spp.: **1**: 5-12
 Pannonian type cattle: **7**: 77-82
 Paraná State: **6**: 19-23; **10**: 7-26
 Pashmina (Cashmere) goat: **22**: 79-82
 Pashmina goat: **19**: 57-68
 Paspalum spp.: **9**: 37-43
 Paspalum spp.: **17**: 147-164
 Paspalum virgatum spp: **9**: 27-35
 Pasture bloat disease: **23**: 1-11
 Pasturellosis: **3**: 1-9
 Patagonia: **15**: 49-57
 Patagonia Argentina: **9**: 23-26
 Patanwadi sheep: **22**: 79-82
 Pavo creollo turkey: **23**: 69-78
 Pawindah camel: **10**: 47-56
 Peanuts: **16**: 29-35
 Peel Neck chicken: **24**: 63-69
 Pemba island: **23**: 13-20
 Península Herminita: **9**: 23-26
 Penna river: **4**: 27-29
 Penniselum typhoides: **18**: 49-57
 Pennisetum spp.: **17**: 147-164; **20**: 9-15
 Pennisetum typhoides: **17**: 101-113
 Pernambuco State: **10**: 7-26
 Peru: **14**: 91-98
 Peruvian Andes: **17**: 115-125
 Peshin camel: **10**: 47-56
 Peste bovine: **13**: 23-37
 Peuhl zebu cattle: **17**: 147-164
 Peul cattle: **17**: 147-164
 Peul Houda sheep: **21**: 1-22
 Peul sheep: **21**: 1-22
 Peul Toronké sheep: **21**: 1-22
 Peul zebu: **13**: 23-37; **21**: 1-22; **22**: 53-60
 Phacochoerus aethiopicus: **7**: 63-70
 Phaseolus spp.: **24**: 1-11
 Piétain pig: **16**: 37-45
 Pinto horse: **3**: 35-38
 Pinus khasiana: **20**: 9-15
 Piper horse: **10**: 67-73
 Piptochaetium spp.: **9**: 37-43
 Pirenaica cattle: **7**: 29-33
 Pithomyces chartarum: **23**: 1-11
 Pitt Island, (New Zealand): **5**: 9-25
 Plevin Black-Face sheep: **14**: 41-59
 Plovdiv: **14**: 41-59
 Pluto horse family: **10**: 67-73
 Plymouth Rock chicken: **17**: 51-60
 Plymouth Rock White chicken: **17**: 73-86
 Poços de Caldas region: **10**: 7-26
 Podgorska cattle: **9**: 7-11
 Podolian cattle: **21**: 49-60
 Podolic cattle: **8**: 59-68
 Poephagus grunniens: **12**: 45-57; **12**: 59-67; **18**: 41-48
 Poephagus mutus Przewalski: **12**: 59-67
 Poitou Charentes region: **17**: 95-99
 Poland: **9**: 7-11
 Polish Prick eared pig: **9**: 7-11
 Polish Whitebacked cattle: **9**: 7-11
 Polled Criollo cattle: **9**: 37-43
 Ponte Alta city: **15**: 59-62
 Ponte Alta region: **9**: 37-43
 Ponwar cattle: **22**: 79-82
 Portland sheep: **4**: 31-36
 Potamochoerus porcus: **7**: 63-70
 Pottoka horse: **19**: 17-26; **22**: 29-42
 Poulfoulo cattle: **17**: 147-164
 Pox goat disease: **8**: 35-41
 PPR diseases: **3**: 1-9
 Prakasam District: **4**: 27-29
 Prealpes sheep: **10**: 57-65
 Prisca goat: **19**: 69-89
 Pro Specie Rara: **9**: 7-11; **14**: 27-32
 Prosopis cineraria: **18**: 29-39
 Ptzevalsky horse: **6**: 29-34
 Pune district: **12**: 77-87
 Punjab State: **10**: 47-56; **18**: 29-39; **18**: 49-57; **25**: 27-31
 Putuo county: **16**: 29-35
 Pyrenean cattle: **22**: 1-5
 Qinghai Province: **9**: 49-60
 Qinghai sheep: **9**: 49-60
 Qingzang plateau: **15**: 75-91
 Qinshan goat: **9**: 49-60
 Qinzang Plateau Areas: **9**: 49-60
 Quarter horse: **3**: 35-38
 Quintana Roo region: **23**: 69-78
 Rabicano allele: **3**: 35-38
 Rajasthan: **10**: 47-56; **16**: 83-89; **16**: 75-82; **21**: 71-80; **22**: 19-27; **18**: 29-39; **18**: 49-57; **25**: 27-31
 Rajputana Maharajahs: **10**: 47-56
 Ramsar wetland sites: **14**: 79-90
 Rank grass: **15**: 75-91
 Rare Breeds Survival Trust: **4**: 31-36; **5**: 1-5
 Rath cattle: **22**: 79-82
 Rato munda sheep: **22**: 19-27
 Rebari camel: **18**: 29-39
 Red African sheep: **10**: 7-26
 Red cattle: **15**: 75-91
 Red Cattle Breeders' Association: **7**: 71-76
 Red (Rahaji) Fulani cattle: **17**: 127-145; **21**: 1-22
 Red Mbororo cattle: **21**: 1-22
 Red Poll cattle: **4**: 31-36
 Red Sindhi cattle: **3**: 21-32; **6**: 25-28; **21**: 1-22; **22**: 79-82; **11**: 21-24
 Red Steppe cattle: **21**: 1-22
 Red-and-White cattle: **7**: 71-76
 Research Animals in Canada: **1**: 27-28
 Rhode Island Red chicken: **7**: 45-53; **14**: 33-39; **15**: 3-22; **17**: 51-60; **17**: 73-86
 Rhodope mountains: **14**: 41-59
 Rice: **16**: 29-35; **19**: 37-43
 Rice bran: **14**: 61-71
 Rio Grande do Sul: **6**: 19-23
 Rivas, Nicaragua: **4**: 15-17

- River buffalo: **14**: 79-90
River type buffalo: **15**: 75-91
Robel: **1**: 5-12
Rohtak: **1**: 29-30
Romagnola cattle: **7**: 77-82; **8**: 59-68
Romanov sheep: **10**: 57-65
Romney Maxsh sheep: **9**: 49-60
Romney sheep: **23**: 1-11
Romosinuano cattle: **9**: 27-35
Rottboellia spp.: **9**: 37-43
Round worms: **8**: 35-41
Rous Sarcoma virus: **14**: 33-39
Roxa cattle: **11**: 77-82
Rubia Gallega cattle: **7**: 29-33
Rubio andaluz pig: **12**: 89-95
Rubio cattle: **7**: 11-18
Russia: **12**: 59-67
Ruta de los Tehuelches: **9**: 23-26
Ryegrass staggers disease: **23**: 1-11
Ryeland sheep: **4**: 31-36
Rygja sheep: **7**: 45-53
Saanen goat: **11**: 41-55; **17**: 61-71
Saaremaa island: **16**: 65-70
Sabino allele: **3**: 35-38
Sahel goat: **21**: 1-22
Sahel Region: **3**: 1-9; **17**: 11-28
Sahiwal cattle: **3**: 21-32; **6**: 25-28; **21**: 1-22; **22**: 79-82
Sahiwal District: **1**: 13-16
Saint Nazaire region: **9**: 61-67
Sakarska sheep: **14**: 41-59
Salvador Malgar: **4**: 15-17
San Clemente goat: **5**: 9-25
Sana'a province: **8**: 35-41
Sanchori camel: **18**: 29-39
Sandomierska goat: **9**: 7-11
Sandveld ecological zone: **20**: 17-26
Sanga cattle: **9**: 13-21; **17**: 127-145; **21**: 35-42
Sangamneri goat: **22**: 79-82
Sanmartinero cattle: **9**: 27-35
Santa Catalina State: **7**: 11-18
Santa Catarina State: **6**: 19-23; **10**: 7-26; **15**: 59-62
Santa Cecilia region: **9**: 37-43
Santa Cruz (City of): **3**: 21-32
Santa Cruz Province: **9**: 23-26; **7**: 7-10
Santa Gertrudis cattle: **9**: 27-35; **10**: 7-26; **17**: 51-60
Sardinia island: **8**: 13-26; **24**: 31-39
Sardinia valley: **11**: 41-55
Sardinian sheep: **16**: 37-45
Sava River: **9**: 7-11; **16**: 37-45
SAVE Foundation: **14**: 27-32
Schyzacchirium spp.: **9**: 37-43
Segureña sheep: **10**: 57-65
Semi fat-tailed Ruda type sheep: **22**: 7-13
Senegal: **14**: 13-25
Senegal river: **17**: 127-145
Senegalese Fulani (Gobra) Cattle: **17**: 127-145
Serbian Sumadija pig: **4**: 19-25
Serere zebu type: **15**: 23-42
Setaria spp.: **9**: 37-43; **20**: 9-15
Shaanbei sheep: **9**: 49-60
Shamos goat: **13**: 11-22
Shandong Province: **15**: 63-73
Shanghai: **16**: 29-35
Shekhawati camel: **18**: 29-39
Shekhawati/Chapper sheep: **22**: 19-27
Shetland cattle: **4**: 31-36
Shetland pony: **22**: 29-42; **24**: 31-39
Shetland sheep: **4**: 31-36
Shewa region: **8**: 79-82
Shimane prefecture: **13**: 11-22
Shizuoka prefecture: **13**: 11-22
Shkodra plain: **16**: 91-103
Shoa Province: **1**: 5-12
Shorthorn cattle: **4**: 31-36; **13**: 3-10; **17**: 51-60; **25**: 45-55
Shorthorn sires: **9**: 45-47
Shorthorned zebu: **17**: 127-145; **17**: 147-164
Shropshire sheep: **4**: 31-36; **7**: 45-53
Sichuan Province: **15**: 63-73; **15**: 75-91; **18**: 23-27
Sichuan sheep: **9**: 49-60
Sicily: **8**: 13-26; **11**: 41-55
Sidamo region: **8**: 79-82
Sierra de la Carba: **14**: 73-78
Sierra del Xistral: **14**: 73-78
Sierra region: **17**: 115-125
Siglavy horse: **4**: 19-25; **10**: 67-73
Sikar area: **22**: 19-27
Sikkim district: **19**: 37-43; **22**: 15-18; **12**: 59-67
Silver Campine chicken: **14**: 33-39
Simbruini Mountains: **8**: 59-68
Simmental cattle: **7**: 71-76; **17**: 51-60; **18**: 79-91; **21**: 1-22
Sindhi camel: **18**: 29-39
Sinú river: **9**: 77-83; **24**: 1-11
Siri cattle: **22**: 79-82
Sirohi camel: **18**: 29-39
Sirohi goat: **22**: 79-82
Sisa: **14**: 27-32
Skaane goose: **7**: 45-53
Skiathos island: **9**: 69-76
Slavonian Grey Steppe type cattle: **16**: 37-45
Slavonki Podolaz type cattle: **16**: 37-45
Slovakian Red cattle: **9**: 7-11
Slovakian region: **9**: 7-11
Smaalens goose: **7**: 45-53
Soay sheep: **4**: 31-36
Sokoto State: **3**: 1-9
Solannum verbacifolium: **19**: 91-108
Somali cattle: **23**: 13-20
Somba: **14**: 13-25
Somba cattle: **13**: 39-45
Sonadi sheep: **16**: 75-82; **22**: 79-82
Sondrio and Varese Provinces: **11**: 41-55
Sopravissana sheep: **8**: 13-26; **11**: 25-33
Sorghum: **18**: 29-39
Sorghum vulgare: **17**: 101-113; **18**: 49-57
South America: **2**: 1-19; **3**: 35-38
South Devon cattle: **17**: 51-60
South West Spanish Mustang Association (SSMA): **3**: 35-38
Southdown sheep: **11**: 25-33
Southern India: **18**: 59-63
Southern Mexico: **23**: 69-78
Southern Saharan region: **15**: 3-22
South-West Asian Region: **8**: 35-41
Southwestern China: **18**: 23-27
Soybean: **19**: 37-43
Spain: **11**: 77-82
Spain (Central plateau): **8**: 13-26
Spain (Northern): **8**: 13-26
Spanish Barb Breeders Association (SBBA): **3**: 35-38
Spanish chicken: **24**: 41-48
Spanish Faco pony: **22**: 29-42
Spanish Giant rabbit: **23**: 49-67
Spanish horse: **16**: 65-70; **22**: 29-42
Spanish Mustang Registry (SMR): **3**: 35-38
Speckled Hungarian fowl: **7**: 83-88
Spel sheep: **7**: 45-53
Spiti horse: **21**: 71-80
Sporades islands: **9**: 69-76
Sporobolus pyramidalis: **20**: 55-69
Sporobolus spp.: **9**: 37-43
Stara Zagora city: **14**: 41-59
Stara Zagora sheep: **8**: 13-26
Steigar sheep: **7**: 45-53
Stipa spp.: **9**: 37-43
Stockholm International Conference on Environment: **12**: 5-23
Strongylus spp.: **20**: 35-54
Sudanese Fulani cattle: **17**: 127-145
Suffolk horse: **4**: 31-36
Suffolk sheep: **10**: 57-65; **16**: 37-45
Sugar cane: **16**: 29-35
Sumava sheep: **9**: 7-11
Sumavsky cattle: **9**: 7-11
Sundarbans: **12**: 69-75
Suri alpaca: **14**: 91-98
Surti cattle: **22**: 79-82
Surti goat: **22**: 79-82

- Sus barbatus barbatus*: **13**: 3-10
Sus scrofa palustris Rutt: **14**: 41-59
 Susques department: **18**: 7-21
 Sussex cattle: **17**: 51-60
 Swamp buffalo: **14**: 79-90; **15**: 75-91
 Sweden: **2**: 20-26
 Swedish Blue duck: **7**: 45-53
 Swedish Mountain cattle: **9**: 27-35
 Swedish Red and White cattle:
 7: 45-53
 Sweet potato: **14**: 61-71; **16**: 29-35
 Swiniarka sheep: **9**: 7-11
 Swiss cattle: **7**: 29-33
 Tai Lake: **7**: 19-22
 Taihu lake: **16**: 29-35
 Taiz area cheese: **8**: 35-41
 Tall fescue disease: **23**: 1-11
 Tamworth pig: **4**: 31-36; **12**: 89-95
 Tan sheep: **9**: 49-60
 Tangshan goat breed: **9**: 49-60
 Tanyang sheep: **20**: 3-8
 Taranto city: **9**: 85-89
 Targhee sheep: **17**: 115-125
 Tarpan horse: **6**: 29-34
 Tattabareji cattle: **17**: 147-164
 Testa Rossa sheep: **8**: 13-26
 Texas Longhorn cattle: **2**: 1-19
 Texel sheep: **7**: 45-53
 Thal camel: **10**: 47-56
 Tharparkar cattle: **22**: 79-82
 Thessaly region: **8**: 13-26
 Thieldon horse: **19**: 17-26
 Thimphu district: **20**: 27-34
 Thoka gene: **7**: 45-53
 Thoroughbred English horse:
 22: 29-42
 Thoroughbred horse: **3**: 35-38;
 10: 7-26
 Thrace region: **14**: 79-90; **22**: 7-13
Thysanolaena agrostis: **20**: 9-15
Thysanolaena maxima: **20**: 9-15
 Tibetan (Glang) cattle: **19**: 27-36
 Tibetan plateau: **12**: 45-57;
 12: 59-67
 Tiergarten Schoenbrunn: **14**: 27-32
 Tla pseke cattle: **13**: 23-37
 Toggenburg goat: **9**: 49-60;
 11: 41-55
 Tokara Island: **13**: 11-22
 Tongxin region: **9**: 49-60
 Tonkinese Red: **3**: 17-20
 Torbiscal pig: **12**: 89-95
 Toronké cattle: **21**: 1-22
 Toscana: **8**: 13-26
 Touabire sheep: **21**: 1-22
 Transferrin: **19**: 69-89; **22**: 29-42;
 23: 41-47
 Transilvania: **7**: 83-88
 Transylvanian buffaloes: **4**: 19-25
 Transylvanian Naked Neck
 chicken: **25**: 73-78
 Trarza region: **15**: 3-22
Trema guineensis: **19**: 91-108
 Trematoda: **24**: 13-30
 Trichostrongyles parasites:
 24: 13-30
Trichostrongylus colubriformis:
 24: 13-30
Trichostrongylus spp.:
 24: 13-30
Trifolium alexandrinum:
 17: 101-113; **18**: 49-57
Trigonella foenum graecum:
 18: 49-57
Triticum aestivum: **17**: 101-113
Triticum vulgare: **18**: 49-57
 Tropoja mountain: **16**: 91-103
 Trypanosomiasis: **1**: 17-20
 Trypanosomiasis resistance:
 17: 127-145; **19**: 91-108
 Trypanotolerance: **1**: 17-20;
 14: 13-25; **16**: 47-63; **17**: 35-79
 Trypanotolerant cattle: **13**: 39-45
 Tsampa: **12**: 45-57
 Tsigai sheep: **9**: 7-11; **9**: 49-60
 Tsuru cattle: **18**: 65-78
 Tsushima island: **13**: 11-22
 Tuberculosis disease: **23**: 1-11
 Tuli cattle: **20**: 17-26
 Tundra reindeer: **7**: 45-53
 Turrialba region: **3**: 21-32
 Tuscany: **8**: 69-77; **8**: 59-68
 Tzigay sheep: **14**: 41-59
 Uberaba region: **9**: 37-43
 UNCED Conference: **9**: 3-6
 UNEP: **12**: 5-23
 Unguja island: **23**: 13-20
 Urate nephritis disease: **22**: 61-69
 Usuku zebu type: **15**: 23-42
 Uttar Pradesh: **17**: 101-113;
 18: 49-57
 Val di Forno: **8**: 13-26
 Val Verzasca (Switzerland):
 11: 41-55
 Valachian Dwarf cattle: **9**: 7-11
 Valachian sheep: **9**: 7-11
 Valdarno chicken: **24**: 41-48
 Valdesequera pig: **12**: 89-95
 Valladolid: **8**: 13-26
 Valle de Losa region: **19**: 17-26
 Vasco-navarro pony: **22**: 29-42
 Verbenaceae familia: **9**: 37-43
Veronia patauus: **9**: 27-35
Vicugna vicugna: **14**: 91-98
Vicugna vicugna mensalis: **18**: 7-21
Vicugna vicugna vicugna: **18**: 7-21
 Vietnam: **14**: 33-39
 Vlorra hillside: **16**: 91-103
 Volcans d'Auvergne (Regional Park
 of): **1**: 25-26
 Wagyu cattle: **18**: 65-78
 Walachian sheep: **9**: 7-11
 Water buffalo: **25**: 33-43
 Wellega Province: **1**: 5-12
 Wello region: **8**: 79-82
 Wenling county: **9**: 45-47
 Wenling Humped cattle: **15**: 75-91
 Wensleydal sheep: **4**: 31-36
 Wessex Saddleback pig: **12**: 89-95
 West Africa: **1**: 17-20; **7**: 63-70
 West African Dwarf goat: **20**: 55-69
 West African pig breed: **7**: 63-70
 West African Shorthorn cattle:
 16: 47-63
 West Bengal: **19**: 37-43
 West Macedonia region: **22**: 7-13
 Western Bhutan region: **22**: 15-18
 Western coastal belt of Sri Lanka:
 4: 9-13
 Western Himalaya region: **19**: 27-36
 Western Egypt: **14**: 33-39
 Western Peloponnese: **16**: 71-74
 Western Turkish coast: **8**: 13-26
 Westphalian Red cattle: **17**: 87-93
 Wheat: **18**: 29-39
 White Bengal goat: **22**: 79-82
 White Fulani cattle: **21**: 1-22
 White Galloway cattle: **4**: 31-36
 White Hungarian fowl: **7**: 83-88
 White Land rabbit: **7**: 45-53
 White Leghorn chicken: **7**: 45-53;
 14: 33-39; **17**: 51-60
 White Mbororo cattle: **21**: 1-22
 White Norwegian goose: **7**: 45-53
 White Park cattle: **4**: 31-36; **9**: 27-35
 White Pele de Boi of Bahia sheep:
 10: 7-26
 White zebu: **21**: 1-22
 Whiteheaded marsk sheep: **7**: 45-53
 Wight City (Jilin): **9**: 45-47
 Wild Goat of Crete: **11**: 41-55
 Wild White cattle: **9**: 27-35
 Wiltshire Horn sheep: **4**: 31-36
 Wollo: **1**: 5-12
 World Watch List: **9**: 3-6;
 13: 3-10; **21**: 1-22
 Wrzosowka sheep: **9**: 7-11
 Wu County: **7**: 19-22
 Württemberg Brown cattle:
 17: 87-93
 Wuzhumuqin sheep: **9**: 49-60
 WWL-DAD: **13**: 3-10
 Xiahe country: **9**: 49-60
 Xinjiang sheep: **9**: 49-60
 Xizang (Tibet) sheep: **9**: 49-60
 Xizang Tibetan Autonomous
 Region: **9**: 49-60
 Yellow Hungarian fowl: **7**: 83-88
 Yellow Italian bee: **7**: 45-53
 Yiecheng sheep: **9**: 49-60
 Yola Gudali cattle: **17**: 147-164
 Yonaguni island: **13**: 11-22
 Yorkshire Large White pig:
 19: 91-108

-
- Yorkshire pig: **7:** 45-53
Yucatan Peninsula: **23:** 69-78
Yuhuan country: **9:** 45-47
Yungui Semi-fine wool sheep:
 9: 49-60
Yunnan province: **18:** 23-27
Zackel sheep: : **11:** 35-40; **14:** 41-59;
 16: 37-45; **22:** 7-13; **23:** 27-39
Zagreb: **14:** 27-32
Zanskari horse: **21:** 71-80
Zaupel sheep: **11:** 35-40
- Zea mais: **17:** 101-113
Zebu: **1:** 17-20; **2:** 1-19; **11:** 21-24;
 13: 23-37; **13:** 39-45; **16:** 29-35;
 17: 35-79; **19:** 91-108; **25:** 109-123
Zebu (Brahman): **9:** 27-35
Zebu cattle: **18:** 41-48; **21:** 35-42
Zemmour sheep: **6:** 11-13
Zhangjiakon (Hebei): **9:** 45-47
Zhejiang Province: **9:** 45-47;
 9: 49-60; **15:** 75-91
- Zhenhai county: **16:** 29-35
Zhongwei Lambskin White goat:
 9: 49-60
Zhongwei region: **9:** 49-60
Zizyphus glabrata: **18:** 29-39
Zlotnicka fowl: **9:** 7-11
Zubr: **16:** 5-12
-

Editorial Policies and Procedures

The mission of the Animal Genetic Resources Information Bulletin (AGRI) is the promotion of information on the better use of animal genetic resources of interest to food and agriculture production, under the Global Strategy for the Management of Farm Animal Genetic Resources. All aspects of the characterization, conservation and utilization of these resources are included, in accordance with the Convention on Biological Diversity. AGRI will highlight information on the genetic, phenotypic and economic surveying and comparative description, use, development and maintenance of animal genetic resources; and on the development of operational strategies and procedures which enable their more cost-effective management. In doing this AGRI will give special attention to contributions dealing with breeds and procedures capable of contributing to the sustainable intensification of the world's medium to low input production environments (agro-ecosystems), which account for the substantial majority of the land area involved in livestock production; the total production of food and agriculture from livestock; and of our remaining farm animal genetic resources.

Views expressed in the paper published in AGRI represent the opinions of the author(s) and do not necessarily reflect those of the institutions which the authors are affiliated, FAO or the Editors.

The suitability of manuscripts for publication in AGRI is judged by the Editors and reviewers.

Electronic publication

AGRI is available in full electronically on the Internet, in addition to being published in hard copy, at:
<< <http://www.fao.org/dad-is>>>

Types of Articles

The following types of articles are published in AGRI.

Research articles

Findings of work on characterization, conservation and utilization of farm animal genetic resources (AnGR) in well described production environments, will be considered for publication in AGRI. Quality photographs of these genetic resources viewed in the primary production environment to which they are adapted, accompanying the manuscripts are encouraged.

Review articles

Unsolicited articles reviewing agro-ecosystems, country-level, regional or global developments on one or more aspects of the management of animal genetic resources, including state-of-the-art review articles on specific fields in AnGR, will be considered for publication in AGRI.

Position papers

Solicited papers on topical issues will also be published as deemed required.

Other published material

This includes book reviews, news and notes covering relevant meetings, training courses and major national, regional and international events and conclusions and recommendations associated with the outcomes of these major events. Readers are encouraged to send such items to the editors.

Guidelines for Authors

Manuscript submission

Manuscripts prepared in English, French or Spanish with an English summary and

another summary in either French or Spanish, should be submitted to AGRI Editor, AGAP, FAO, Viale delle Terme di Caracalla, 00100 Rome, Italy. Alternatively a manuscript may be sent as a WinWord Electronic Mail attachment to < agri@fao.org >. Photographs, coloured or black and white, and figures must be always sent by mail.

Manuscripts should be typed double-spaced and with lines numbered in the left margin. All pages, including those of references, tables etc., must be consecutively numbered. The corresponding author is notified of the receipt of a manuscript.

For manuscripts that are accepted after revision, authors are encouraged to submit a last version (3½" disc format) in Word 6.0 for Windows of their revised manuscript along with the printed copy.

Preparation of the manuscript

The first page of the manuscript must include the running head (abbreviated title), title, names of authors, institutions, full addresses including postal codes and telephone number and other communication details (fax, e-mail, etc.) of the corresponding author. The running head not exceeding 45 characters plus spaces, should appear at the top of page 1 of the manuscript entirely in capital letters. The title of the manuscript is typed in upper and lower case letters. The title should be as brief as possible not exceeding 150 characters (including spaces) with species names when applicable. Authors, institutions and addresses are in upper and lower case italics. There is one blank line between the title and the authors. Addresses are typed as footnotes to the authors after leaving one blank line. Footnotes are designated numerically. Two lines are left below the footnotes.

Headings

Headings of sections, for example Summary, Introduction, etc., are left-justified. Leave two blank lines between addresses footnotes and Summary and between the heading Summary and its text. Summary should not exceed 200

words. It should be an objective summary briefly describing the procedures and findings and not simply stating that the study was carried on such and such and results are presented, etc. Leave one line between the summary text and Keywords which is written in italics as well as the keywords themselves. All headings of sections (14 regular) and sub-sections (12 regular) are typed bold and preceded and succeeded by one blank line and their text begins with no indentation. The heading of a sub-subsection is written in italics, and ends with a dot after which the text follows on the same line. Keywords come immediately after the summaries. They should be no more than six, with no "and" or "&".

Tables and figures

Tables and figures must be enclosed with the paper and attached at the end of the text according their citation in the document. Photos will not be returned

Tables

Tables, including footnotes, should be preceded and succeeded by 2 blank lines. Table number and caption are written, above the table, in italics (12) followed by a dot, then one blank line. For each column or line title or sub-title, only the 1st letter of the 1st word is capitalized. Tables should be numbered consecutively in Arabic numerals. Tables and captions should be left justified as is the text. Use horizontal or vertical lines only when necessary. Do not use tabs or space-bar to create a table but only the appropriate commands.

Figures

Figures including titles and legends should be preceded and succeeded by two blank lines. Figure number and title are written, below the figure, in italics (12) and end with a dot. The term figures includes photos, line drawings, maps, diagrams etc.

All the submitted diagrams, must be

.....

accompanied with the original matrix of the data used to create them. It is strongly advised to submit diagrams in Word 6.0 or Excel 5.0. Figures should be numbered consecutively in Arabic numerals.

References

Every reference cited in the text should be included in the reference list and every reference in the reference list should have been mentioned in the text at least once. References should be ordered firstly alphabetically by the first author's surname and secondly by year.

Example for reference in a periodical is:

Köhler-Rollefson, I., 1992; The camel breeds of India in social and historical perspective. *Animal Genetic Resources Information* 10, 53-64.

When there are more than one author:

Matos, C.A.P., D.L. Thomas, D. Gianola, R.J. Tempelman & L.D. Young, 1997; Genetic analysis of discrete reproductive traits in sheep using linear and nonnlinear models: 1. Estimation of genetic parameters 75, 76-87.

For a book or an ad hoc publication, e.g., reports, theses, etc.:

Cockril, W.R., (Ed), 1994; *The Husbandry and Health of the Domestic Buffalo*. FAO, Rome, Italy, pp 993.

For an article in the proceedings of a meeting:

Hammond, K., 1996; FAO's programme for the management of farm animal genetic resources. In C. Devendra (Ed.) *Proceedings of IGA/FAO Round Table on the Global Management of Small Ruminant Genetic Resources*, Beijing, May 1996, FAO, Bangkok, Thailand, 4-13.

Where information included in the article has been obtained or derived from a World Wide Web site, then quote in the text, e.g. "derived from FAO. 1996" and in the References quote the URL standard form:

FAO, 1996; *Domestic Animal Diversity Information System* <<http://www.fao.org/dad-is/>>, FAO, Rome

.....

Normes et règles éditoriales

L'objectif du Bulletin d'Information sur les Ressources Génétiques Animales (AGRI) est la vulgarisation de l'information disponible sur la meilleure gestion des ressources génétiques animales d'intérêt pour la production alimentaire et agricole, d'après les recommandations de la Stratégie Mondiale pour la Gestion des Ressources Génétiques des Animaux Domestiques. Tous les aspects relatifs à la caractérisation, la conservation et l'utilisation de ces ressources seront pris en considération, suivant les normes de la Convention pour la Biodiversité.

AGRI désire diffuser de l'information sur la génétique, les enquêtes phénotypiques et économiques et les descriptions comparatives, l'utilisation et la conservation des ressources génétiques animales, ainsi que toute information sur le développement de stratégies opérationnelles et de normes qui puissent permettre une meilleure gestion de la relation coût/efficacité. C'est pour cela que AGRI prendra spécialement en considération toutes les contributions référées aux races et aux normes capables de permettre une intensification durable des milieux (agroécosystèmes) à revenus moyens et bas dans le monde; qui comprennent la majeure partie des terres consacrées à l'élevage, à la production totale des aliments et l'agriculture provenant de l'élevage; et tout ce qui reste comme ressources génétiques des animaux domestiques.

Les opinions exprimées dans les articles publiés dans AGRI appartiennent seulement aux auteurs et donc ne représentent pas nécessairement l'opinion des instituts pour lesquels ils travaillent, la FAO ou les éditeurs.

L'opportunité ou non de publier un article dans AGRI sera jugée par les éditeurs et les réviseurs.

Publication électronique

En plus de sa version imprimée, la version totale de AGRI se trouve disponible sur Internet, sur le site:

<<<http://www.fao.org/dad-is/>>>

Types d'articles

Les articles suivants pourront être publiés sur AGRI:

Articles de recherche

Seront prises en considération pour leur publication sur AGRI les études sur la caractérisation, la conservation et l'utilisation des ressources génétiques des animaux domestiques (AnGR) accompagnées d'une bonne description du milieu. On encourage les auteurs à envoyer des photographies de bonne qualité qui montrent les races en question dans leur milieu naturel de production.

Révisions

Occasionnellement, des articles contenant une révision des agroécosystèmes, au niveau national, régional ou mondial, avec un ou plusieurs aspects se rapportant à la gestion des ressources génétiques animales, y comprises les mises à jour des différentes zones de AnGR, seront pris en considération.

Articles spécifiques

Ponctuellement, des articles sur des thèmes spécifiques pourront être demandés pour la publication d'éditions spéciales.

Autre matériel pour publication

Ceci comprend la révision de livres, nouvelles et notes de réunions importantes, cours de formation et principaux événements nationaux, régionaux et internationaux; ainsi que les conclusions et recommandations par rapport aux objectifs de ces principaux événements. Les auteurs sont priés d'envoyer ce genre de matériel aux éditeurs.

Guide pour les auteurs

Présentation du manuscrit

Les articles se présenteront en anglais, français ou espagnol, avec un résumé en anglais et sa traduction en français ou en espagnol; et seront envoyés à l'éditeur de AGRI, AGAP, FAO, Viale delle Terme di Caracalla, 00100 Rome, Italie. L'autre possibilité est d'envoyer l'article par courrier électronique avec le document adjoint en version WinWord à <agri@fao.org>. Les photographies, en couleur ou en blanc et noir, seront toujours envoyées par courrier normal.

Les manuscrits se présenteront à double interligne et avec le numéro correspondant à chaque ligne sur la marge gauche. Toutes les pages seront numérotées, y comprises celles avec les références bibliographiques, les tableaux, etc. L'auteur recevra une lettre lui donnant bonne réception de son document.

Lorsqu'un article, après sa révision, sera accepté, on demandera à l'auteur d'envoyer la version finale révisée sur disquette (format 3 1/2") en Word 6.0 x Windows, ainsi qu'une copie sur papier.

Préparation du manuscrit

Sur la première page du manuscrit on indiquera le titre de l'article en abrégé, le titre et noms des auteurs, des institutions, les adresses complètes (y compris code postal et numéro de téléphone); ainsi que tout autre moyen de contact tel que fax, e-mail, etc. avec l'auteur principal. Le titre abrégé ne devra pas dépasser les 45 caractères, plus les espaces nécessaires, et s'écrira sur la partie supérieure de la page 1 du manuscrit en majuscules. Le titre en entier du manuscrit sera écrit en majuscules et minuscules; il devra être aussi bref que possible, sans dépasser les 150 caractères (y compris les espaces nécessaires), et avec l'indication des noms des espèces. Les noms des auteurs, des institutions et les adresses seront en italique et en lettres majuscules et minuscules. On laissera un espace en blanc entre le titre et les noms des auteurs. Les adresses seront indiquées comme

des notes à pied de page pour chacun des auteurs après avoir laissé un espace en blanc après les noms. Chaque note de pied de page sera numérotée. On laissera deux espaces en blanc après les adresses.

Titres

Les titres de chaque chapitre, par exemple Résumé, Introduction, etc. seront alignés à gauche. Laisser deux espaces en blanc entre les notes de pied de page avec les adresses et le Résumé, et entre le titre Résumé et le texte qui suit. Le résumé ne devra pas dépasser les 200 mots. Il s'agira d'un résumé objectif qui fasse une brève description des processus utilisés et des résultats obtenus, et non pas une simple présentation du travail réalisé avec une description générale des résultats. Laisser un espace en blanc entre la fin du texte du résumé et les mots-clés, qui seront écrits en italique ainsi que le titre Mots-clés. Les mots-clés seront au maximum six et il ne devra pas y avoir de "et" ou "&". Tous les titres principaux de chapitre (14 regular) et sous-chapitre (12 regular) seront en gras avec un espace en blanc avant et après. Le texte commencera sans retrait. Un titre à l'intérieur d'un sous-chapitre s'écrira en italique, suivi d'un point, avec le texte à continuation.

Tableaux et figures

Les tableaux et les figures iront à la fin du texte en suivant l'ordre d'apparition dans le texte. Les photographies ne seront pas dévolues aux auteurs.

Tableaux

Les tableaux, y compris les notes de pied de page, devront avoir un espace en blanc avant et après. Le numéro du tableau et le titre s'écriront sur la partie supérieure en italique (12) avec un point à la fin et un espace en blanc en dessous. Sur chaque colonne, titre d'en-tête ou sous-titre, seulement la première lettre du premier mot sera en majuscule. Les tableaux et leur titre seront alignés à gauche, ainsi que le texte. Les lignes verticales et

horizontales seront utilisées seulement si nécessaires. Ne pas utiliser les tabs ou la barre de séparation pour créer un tableau.

Figures

Les figures, y compris les titres et les légendes, seront précédés et suivis de deux espaces en blanc. Le numéro de la figure et le titre s'écriront sur la partie supérieure en italique (12) avec un point à la fin. Sous la rubrique figure on trouvera les photographies, les graphiques, les cartes, les diagrammes, etc. Dans le cas des diagrammes, la matrice originale avec les données utilisées pour son élaboration devra être envoyée. On recommande l'utilisation de Word 6.0 ou Excel 5.0 pour la présentation des diagrammes.

Références

Toute référence présente dans le texte devra apparaître sur la liste des références, et chaque référence de la liste aura été citée au moins une fois dans le texte. Les références iront en ordre alphabétique du nom de l'auteur, suivi de l'année. Exemple dans le cas d'une référence sur une revue:

Köhler-Rollefson, I., 1992; The camel breeds of India in social and historical perspective. *Animal Genetic Resources Information* 10, 53-64.

Lorsqu'il s'agit de plus d'un auteur:

Matos, C.A.P., D.L. Thomas, D. Gianola, R.J. Tempelman & L.D. Young, 1997; Genetic analysis of discrete reproductive traits in sheep using linear and non-linear models: 1. Estimation of genetic parameters 75, 76-87.

Dans le cas d'un livre ou d'une publication ad hoc, par exemple un rapport, une thèse, etc.:

Cockril, W.R., (Ed), 1994; *The Husbandry and Health of the Domestic Buffalo*. FAO, Rome, Italy, pp 993.

S'il s'agit d'un acte d'une réunion:

Hammond, K., 1996; FAO's programme for the management of farm animal genetic resources. In C. Devendra (Ed.) *Proceedings of IGA/FAO Round Table on the Global Management of Small Ruminant Genetic Resources*, Beijing, May 1996, FAO, Bangkok, Thailand, 4-13.

Lorsque l'information contenue dans l'article ait été obtenue ou dérive d'un site World Wide Web, il faudra mettre le texte entre guillemets; par exemple "tiré de la FAO. 1996" et indiquer dans les Références la forme standard URL:

FAO, 1996; Domestic Animal Diversity Information System <<http://www.fao.org/dad-is/>>, FAO, Rome

Reglas y normas editoriales

El objetivo del Boletín de Información sobre Recursos Genéticos Animales (AGRI) es la divulgación de la información sobre una mejor gestión de los recursos genéticos animales de interés para la producción alimentaria y agrícola, siguiendo la Estrategia Mundial para la Gestión de los Recursos Genéticos de los Animales Domésticos. Todos los aspectos referidos a la caracterización, la conservación y el uso de estos recursos serán tomados en consideración, de acuerdo con la Convención sobre la Biodiversidad.

AGRI publicará información sobre genética, encuestas fenotípicas y económicas y descripciones comparativas, uso, desarrollo y conservación de los recursos genéticos animales, así como sobre el desarrollo de estrategias operacionales y normas que permitan una gestión más eficaz de la relación costo/eficacia. Por ello, AGRI prestará especial atención a las contribuciones referidas a razas y normas capaces de contribuir a la intensificación sostenible de los medios (agroecosistemas) con ingresos medio y bajos en el mundo, que comprenden casi la mayor parte de las tierras dedicadas a la producción ganadera; la producción total de alimentos y agricultura provenientes de la ganadería; y el resto de los recursos genéticos de animales domésticos.

Los puntos de vista expresados en los artículos publicados en AGRI son solamente las opiniones de los autores y, por tanto, no reflejan necesariamente la opinión de las instituciones para las cuales trabajan dichos autores, de la FAO o de los editores.

La oportunidad o no de publicar un artículo en AGRI será juzgada por los editores y revisores.

Publicación electrónica

Además de su publicación impresa, la versión íntegra de AGRI se encuentra disponible electrónicamente sobre Internet, en el sitio: <<<http://www.fao.org/dad-is/>>>

Tipos de artículos

Serán publicados en AGRI los siguientes tipos de artículos:

Artículos sobre investigación

Se tomarán en consideración para su publicación en AGRI los estudios sobre la caracterización, conservación y uso de los recursos genéticos de los animales domésticos (AnGR) con una buena descripción del entorno. Se agradecerá el envío de fotografías de calidad que presenten a las razas en cuestión en su ambiente natural de producción.

Artículos de revisión

Se podrán tener en consideración ocasionalmente aquellos artículos que presenten una revisión de los agroecosistemas, a nivel nacional, regional o mundial, con el desarrollo de uno o más aspectos referidos a la gestión de los recursos genéticos animales, incluidas las revisiones sobre el estado actual de las distintas áreas de AnGR.

Artículos específicos

Se solicitarán puntualmente artículos sobre temas específicos para ediciones especiales.

Otro material para publicación

Incluye la revisión de libros, noticias y notas referidas a reuniones importantes, cursos de formación y principales eventos nacionales, regionales e internacionales, así como conclusiones y recomendaciones relacionadas con los objetivos de estos principales eventos. Se invita a los lectores a enviar este tipo de material a los editores.

Guía para los autores

Presentación del manuscrito

Los artículos se presentarán en inglés, francés o español, junto con un resumen en inglés y su traducción en francés o español, y se enviarán al editor de AGRI, AGAP, FAO, Viale delle Terme di Caracalla, 00100 Roma, Italia. Otra posibilidad es enviar el artículo por correo electrónico adjuntando el documento en versión WinWord a <agri@fao.org>. Las fotografías, a color o en blanco y negro, se enviarán siempre por correo normal.

Los manuscritos se presentarán con doble espacio y con el número correspondiente a cada línea en el margen izquierdo. Todas las páginas serán numeradas, incluidas las de las referencias bibliográficas, cuadros, etc. El autor recibirá una notificación sobre la recepción de su documento.

En el caso de aceptación de un artículo después de su revisión, se solicitará al autor una versión final de su artículo revisado en disquete (formato 3¹/₂" en Word 6.0 x Windows, así como una copia impresa del mismo.

Preparación del manuscrito

En la primera página del manuscrito se indicará el título abreviado del artículo, títulos y nombres de los autores, instituciones, direcciones completas (incluido código postal y número de teléfono); así como otros medios de contacto tales como fax, e-mail, etc., del autor principal. El título abreviado no deberá sobrepasar los 45 caracteres más los espacios correspondientes, y aparecerá en la parte superior de la página 1 del manuscrito en mayúsculas. El título entero del manuscrito viene escrito en mayúsculas y minúsculas. Dicho título debe ser lo más breve posible y no sobrepasar los 150 caracteres (incluidos los espacios necesarios), con los nombres de las especies, si necesario. Los nombres de los autores, instituciones y direcciones se escribirán en cursiva y en letras mayúsculas y minúsculas. Se dejará una línea en blanco

entre el título y los nombres de los autores. Las direcciones se escribirán como notas de pie de página de cada autor después de dejar una línea en blanco entre los nombres y éstas. Cada nota de pie de página con la dirección vendrá indicada numéricamente. Se dejarán dos líneas en blanco después de las direcciones.

Títulos

Los títulos de cada sección, por ejemplo Resumen, Introducción, etc., vienen alineados a la izquierda. Dejar dos líneas en blanco entre las notas de pie de página con las direcciones y el Resumen y entre el título Resumen y el texto que sigue. El resumen no deberá exceder de 200 palabras. Deberá ser un resumen objetivo que describa brevemente los procesos y logros obtenidos, y no una presentación de cómo se ha llevado a cabo el estudio y una descripción genérica de los resultados. Dejar una línea en blanco entre el final del texto del resumen y las palabras clave, que se escribirán en cursiva así como el título Palabras clave. No deberán ser más de seis y no deberán contener "y" o "&". Todos los títulos principales de capítulo (14 regular) y subcapítulo (12 regular) serán en negrita e irán precedidos y seguidos de una línea en blanco. El texto correspondiente empezará sin sangrado. Un título dentro de un subcapítulo se escribirá en cursiva e ira seguido de un punto con a continuación el texto correspondiente.

Cuadros y figuras

Los cuadros y las figuras se incluirán al final del texto siguiendo el orden de cita dentro del mismo. Las fotografías no serán devueltas a sus autores.

Cuadros

Los cuadros, incluidas las notas de pie de página, deberán ir precedidos y seguidos por dos líneas en blanco. El número del cuadro y su título se escribirán en la parte superior en cursiva (12) con un punto al final y seguido

de una línea en blanco. En cada columna o título de encabezamiento o subtítulo, sólo la primera letra de la primera palabra irá en mayúscula. Los cuadros irán numerados de forma consecutiva con números árabes. Los cuadros y sus títulos se alinearán a la izquierda, así como el texto. Se utilizarán líneas horizontales o verticales sólo cuando sea necesario. No utilizar tabuladores o la barra espaciadora para crear un cuadro.

Figuras

Las figuras, incluidos los títulos y leyendas, irán precedidas y seguidas de dos líneas en blanco. El número de la figura y el título se escribirán en la parte superior en cursiva (12) con un punto al final. La palabra figura incluye las fotografías, los gráficos, los mapas, los diagramas, etc. En el caso del diagrama se enviará la matriz original con los datos utilizados para crearlo. Se recomienda encarecidamente la utilización de Word 6.0 o Excel 5.0 para la presentación de los diagramas.

Referencias

Toda referencia presente en el texto deberá aparecer en la lista de referencias y, de la misma manera, cada referencia de la lista deberá haber sido citada por lo menos una vez en el texto. Las referencias deben ir en orden alfabético del apellido del autor, seguido por el año.

Ejemplo en el caso de una referencia de una revista:

Köhler-Rollefson, I., 1992; The camel breeds of India in social and historical perspective. *Animal Genetic Resources Information* 10, 53-64.

Cuando se trata de más de un autor:

Matos, C.A.P., D.L. Thomas, D. Gianola, R.J. Tempelman & L.D. Young, 1997; Genetic analysis of discrete reproductive traits in sheep using linear and nonnlinear models: 1. Estimation of genetic parameters 75, 76-87.

En el caso de un libro o de una publicación ad hoc, por ejemplo informes, tesis, etc.:

Cockril, W.R., (Ed), 1994; *The Husbandry and Health of the Domestic Buffalo*. FAO, Rome, Italy, pp 993.

Cuando se trate de un artículo dentro de las actas de una reunión:

Hammond, K., 1996; FAO's programme for the management of farm animal genetic resources. In C. Devendra (Ed.) *Proceedings of IGA/FAO Round Table on the Global Management of Small Ruminant Genetic Resources*, Beijing, May 1996, FAO, Bangkok, Thailand, 4-13.

Cuando la información contenida en el artículo haya sido obtenida o derive de un sitio World Wide Web, poner el texto entre comillas; por ejemplo "sacado de la FAO. 1996" e indicar en las Referencias la forma estándar URL:

FAO, 1996; Domestic Animal Diversity Information System <<http://www.fao.org/dad-is/>>, FAO, Rome