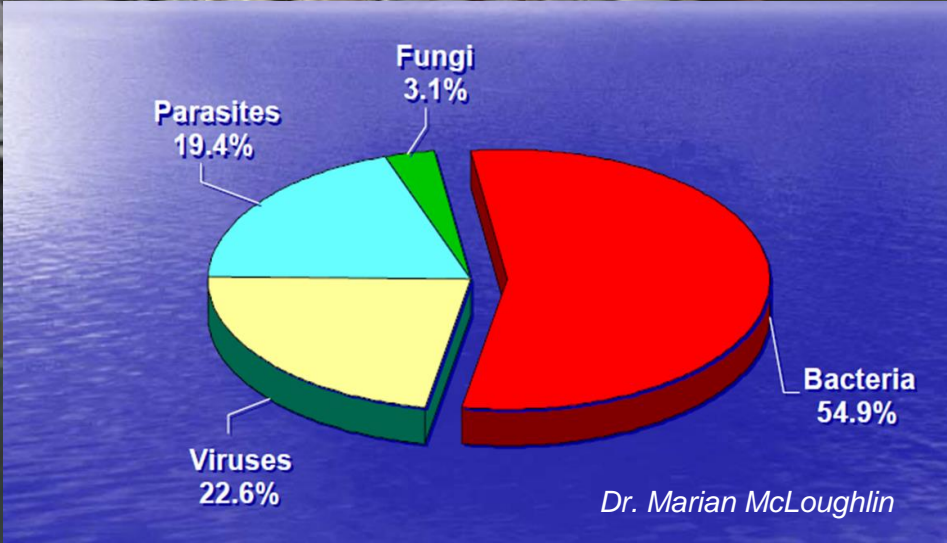
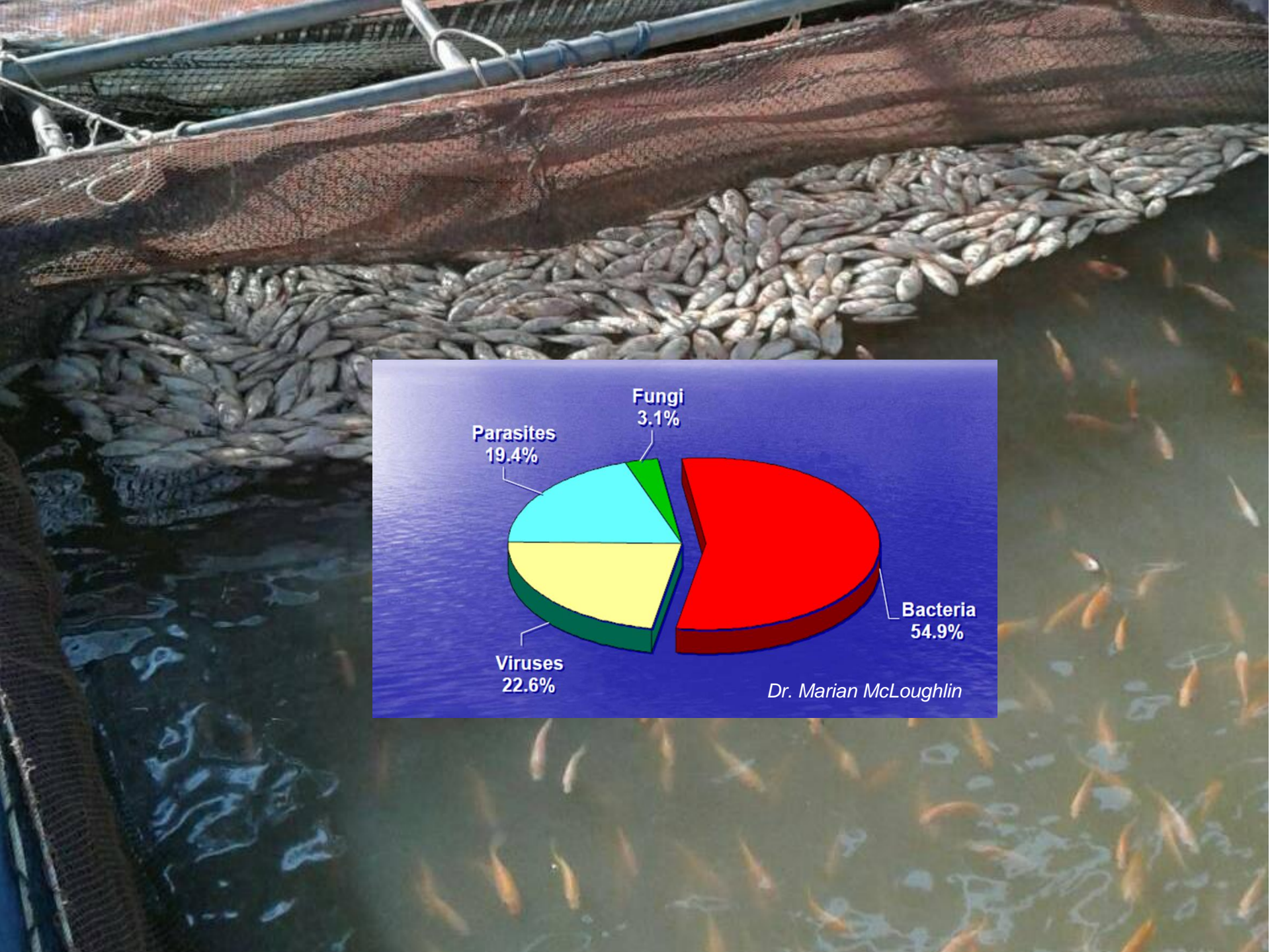


Bacterial Diseases of Tilapia

Ha Thanh Dong

Faculty of Science and Technology, Suan Sunandha Rajabhat University
Fish Health Platform, CENTEX Shrimp, BIOTEC/Mahidol University



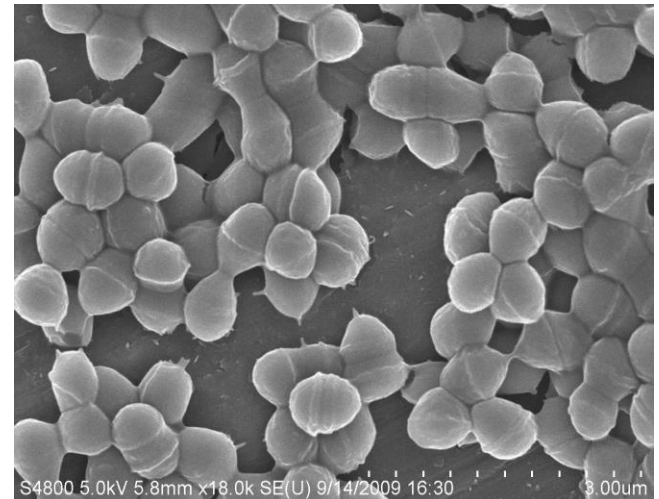
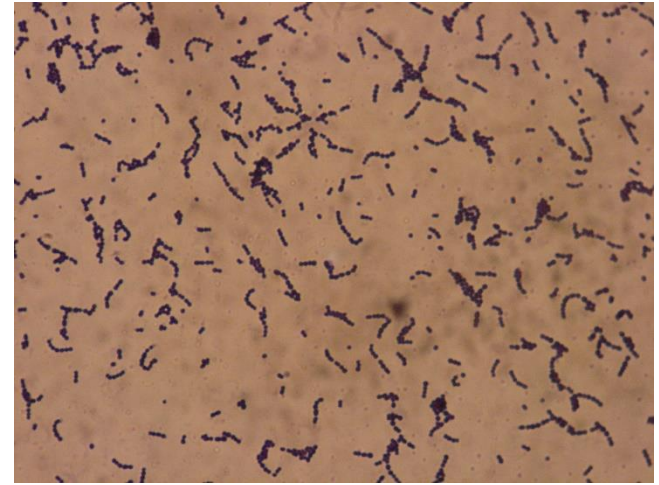
Bacterial Diseases in Farmed Tilapia

- ❖ Streptococcosis
- ❖ Columnaris
- ❖ Francisellosis
- ❖ Edwardsiellosis
- ❖ Hemorrhagic septicemia caused by motile aeromonads
- ❖ Hahellosis (red egg disease)
- ❖ Epitheliocystis
- ❖ Miscellaneous disease

Streptococcosis

❖ Causative agents

- *Streptococcus agalactiae*
 - *Streptococcus iniae*
 - *Streptococcus dysagalactiae*
 - *Lactococcus garviae*
 - *Aerococcus viridans* (?)
-
- Gram positive bacteria
 - GBS have been classified to 10 serotypes (Ia, Ib, II–IX)
 - In aquatic animals: 4 serotypes Ia, Ib, II and III
 - Serotype IX is newly reported in tilapia (China)

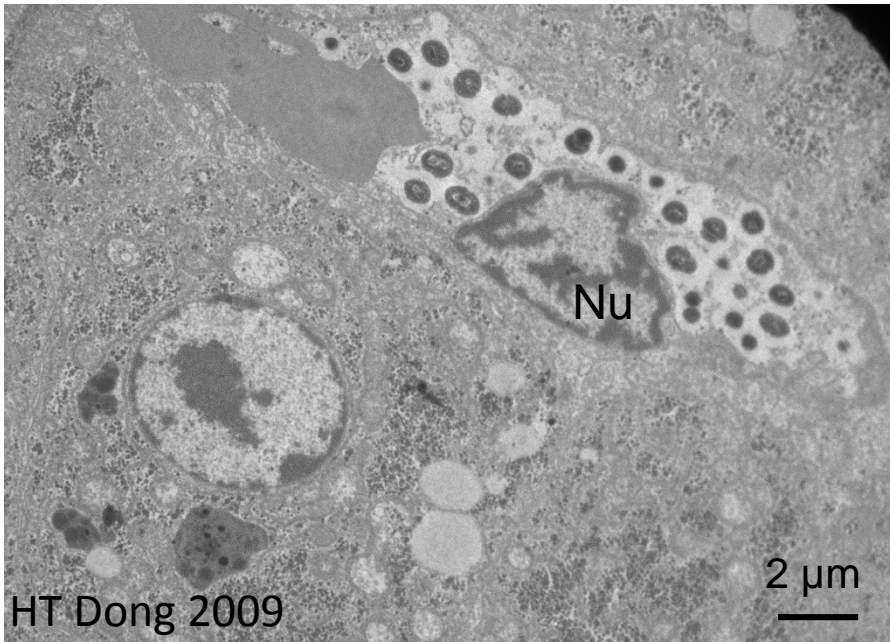


Streptococcus agalactiae

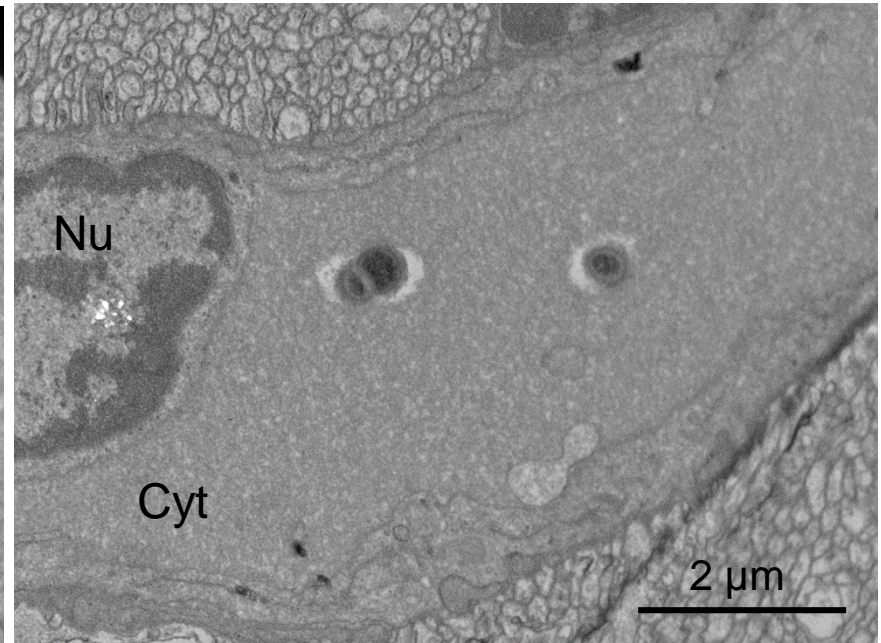
Streptococcosis

❖ Causative agents

Intracellular living style of *S. agalactiae*



Liver



Brain

Streptococcosis

❖ Clinical signs (Level 1)

- erratic swimming
- pop eyes (exophthalmia)
- darkening
- lost of appetite

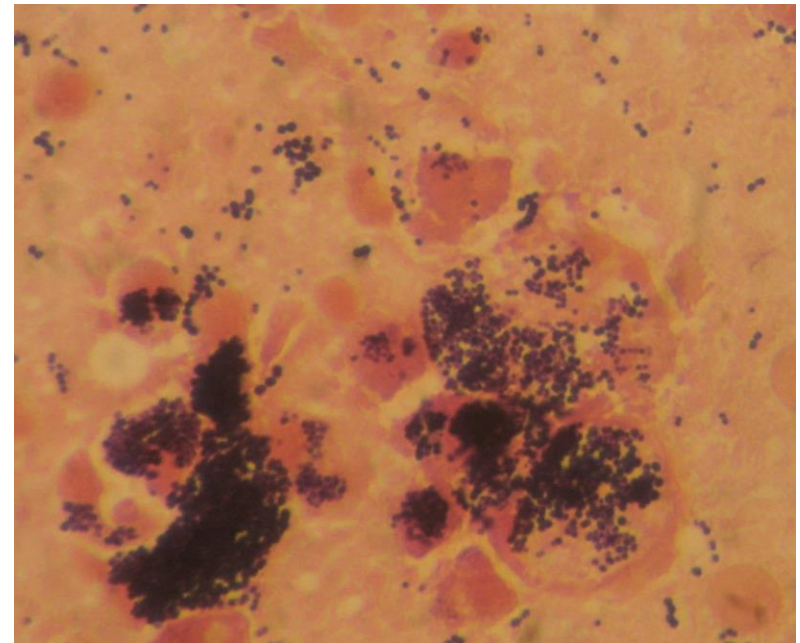
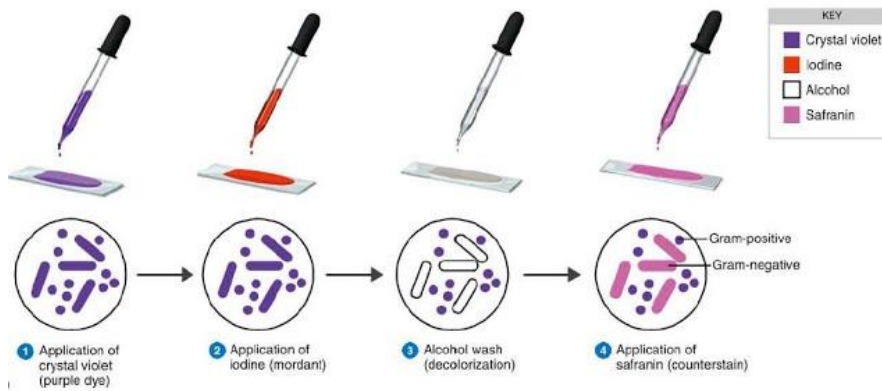


- swollen gallbladder
- brown/dark areas in the liver
- ascites

Streptococcosis

Diagnostics (Level 2)

- Rapid Gram staining of smeared tissue e.g. blood, kidney, liver, brain

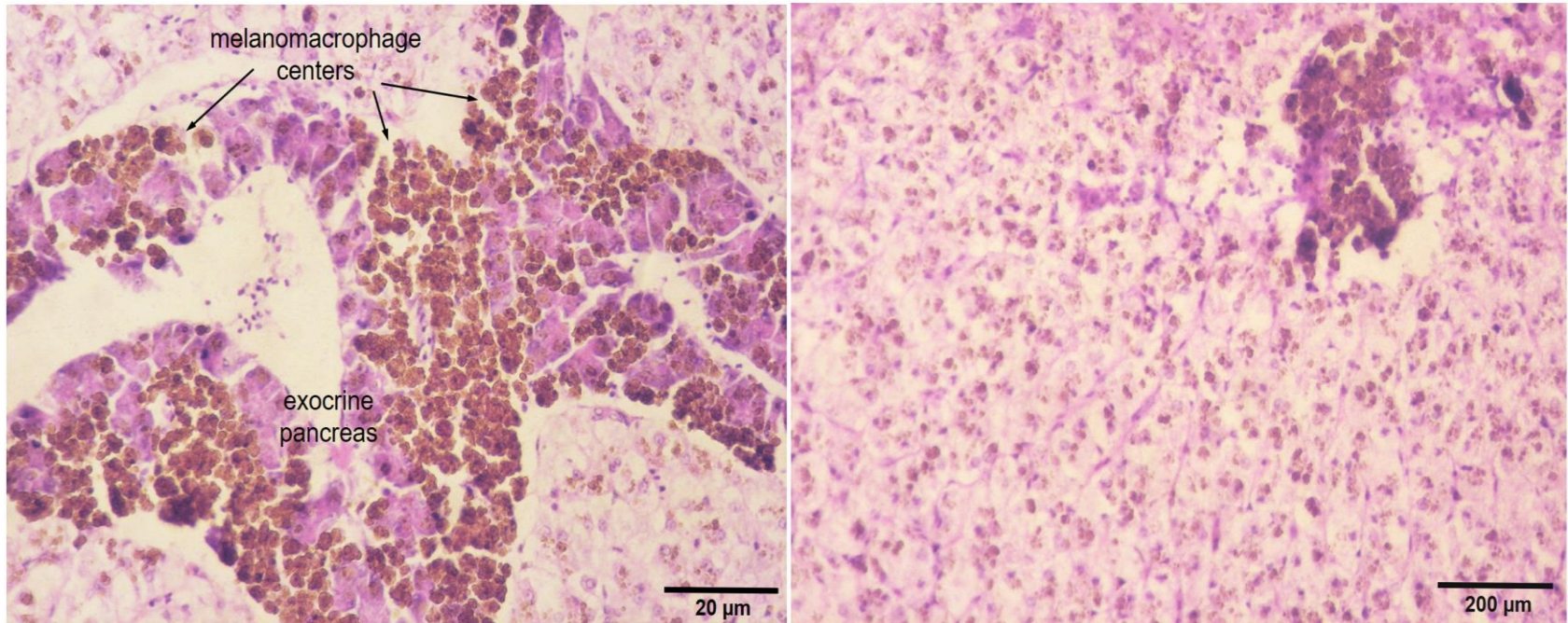


Gram staining of smeared kidney

Streptococcosis

Diagnostics (Level 2)

- histopathology

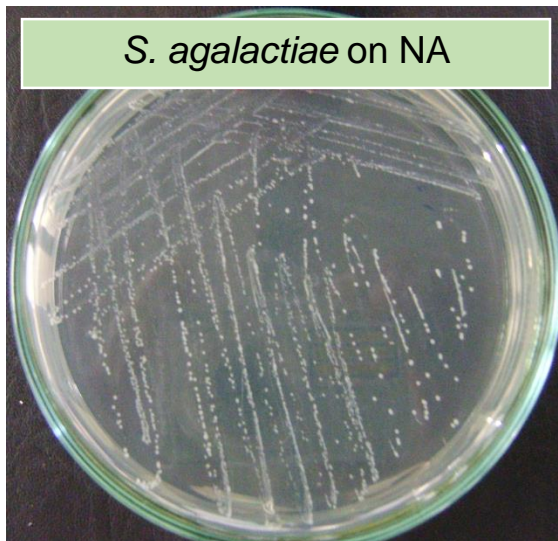


Liver, Nile tilapia infected with *S. agalactiae* showing hyperactivation of melanomacrophage centers with overloaded melanophores in the pancreas, severe hepatocyte degeneration and accumulation of melanophores.

Streptococcosis

Diagnostics (Level 2)

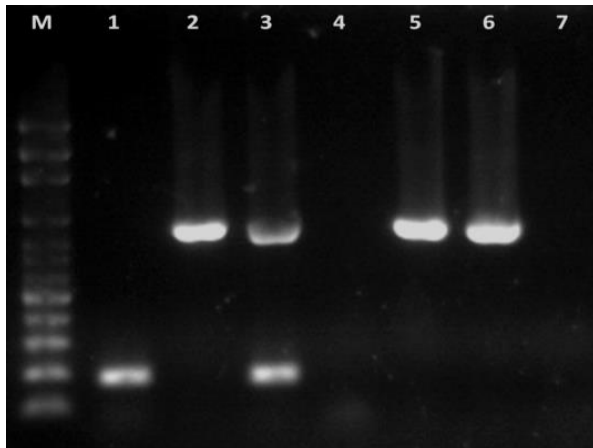
- Bacterial culture + biochemical tests
 - culture: e.g. TSA, NA, blood agar
 - incubate at 28-33 °C for 24-48 hrs. → pinpoint colonies
 - Gram positive, oval/round shape, catalase (-) → *Streptococcus*
 - API 20 strep kit



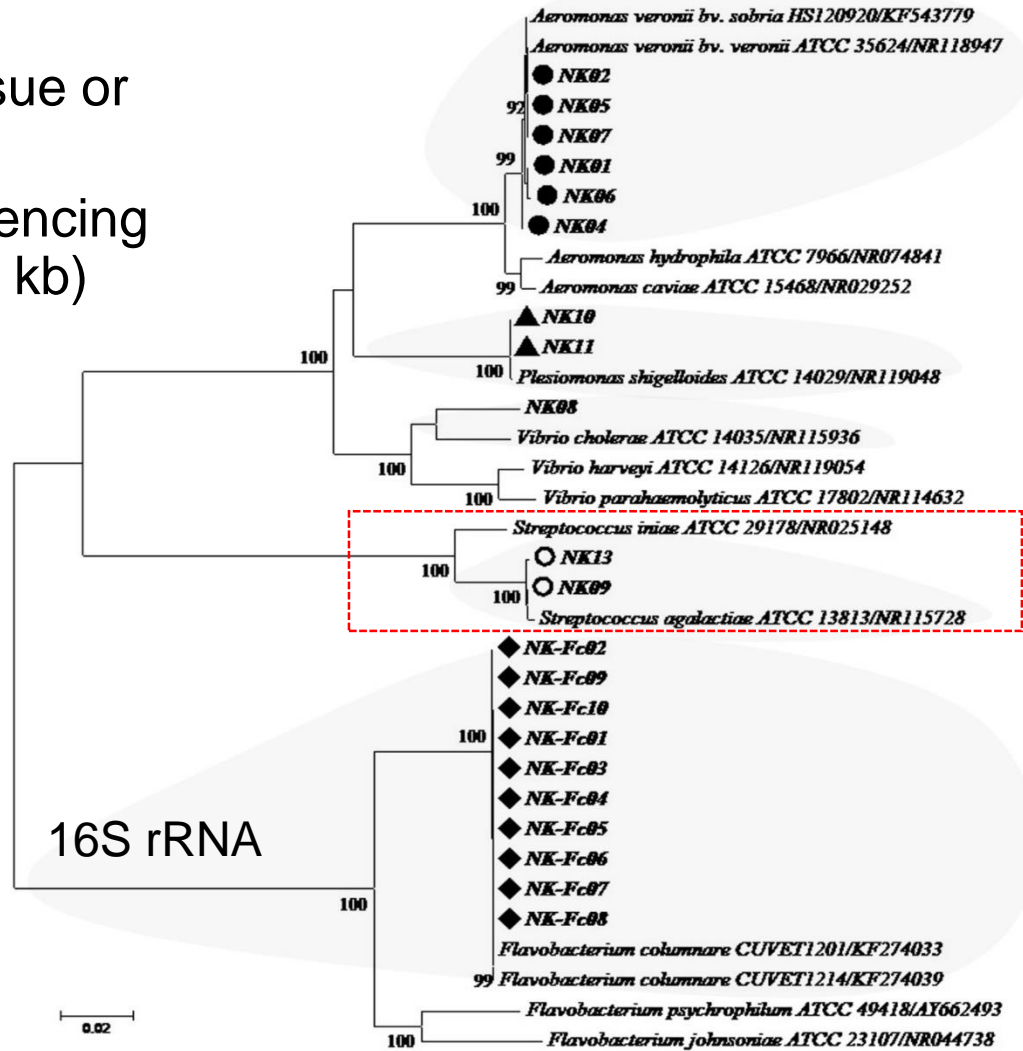
Streptococcosis

• Diagnostics (Level 3)

- Specific PCR (either tissue or pure culture)
- Bacterial culture + sequencing of 16S rRNA gene (>1.3 kb)



Duplex PCR for detection of *S. agalactiae* & *S. iniae* (Rodkhum et al. 2012)



Streptococcosis

- ❖ **Mortality:** variable, up to 90%
- ❖ **Geographical distribution:** worldwide
- ❖ **Risk factors:** high temperature (31-33 °C), poor water quality
- ❖ **Prevention**
 - Treat water with disinfectants
 - Control water temperature (e.g. increased water level in the pond)
 - Supply more oxygen (e.g. increased aeration)
 - Improve fish immunity (e.g. vitamin C, immunostimulants)
 - **Vaccine is available**
 - Injectable vaccine (e.g. MSD, Dr. Nontawith's team at KU, Thailand)
 - Immersion vaccine (China)
 - Vaccine incorporated with feed for oral route (DOF, Malaysia)

Columnaris

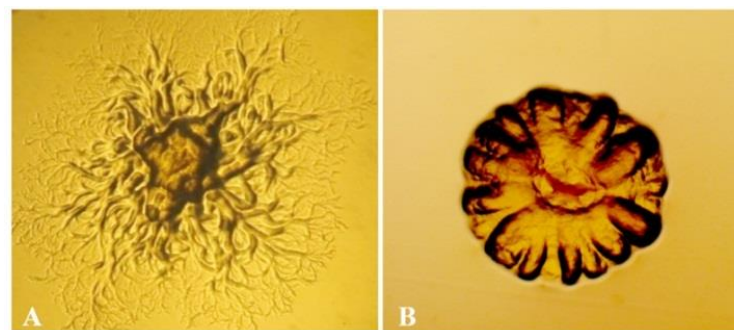
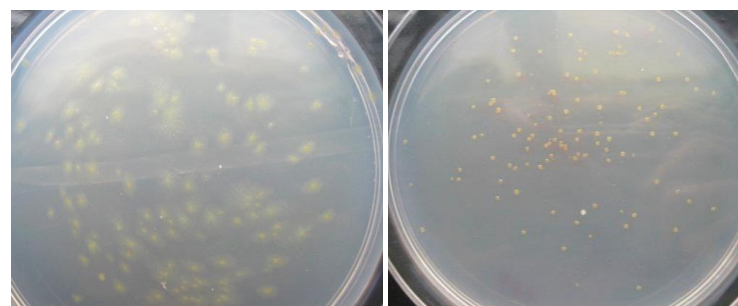
Columnaris

❖ Causative agent

- *Flavobacterium columnare*
- Gram negative, long rod-shaped bacterium
- Gliding motility
- Form strong biofilm
- Rhizoid morphotype: pathogenic
- Non-Rz morphotype: non-pathogenic



Gram negative, long rod-shape bacterium



Rhizoid morphotype

Non-Rhizoid morphotype

Biofilm formation

Columnaris

❖ Clinical signs (Level 1)

- fin rot
- necrotic gills
- muscle necrosis
- “saddle back” lesion
- pale skin



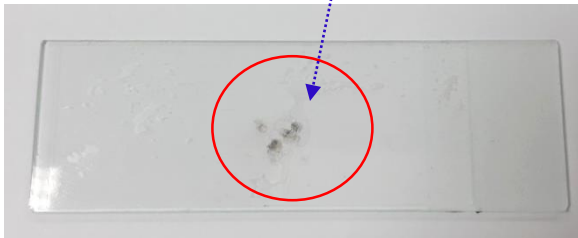
“saddle back”



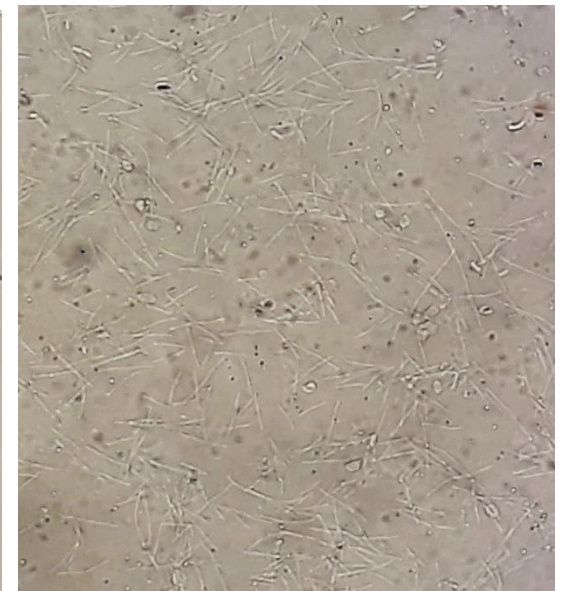
Columnaris

Diagnostics (Level 2)

- wet-mount



Wet mount

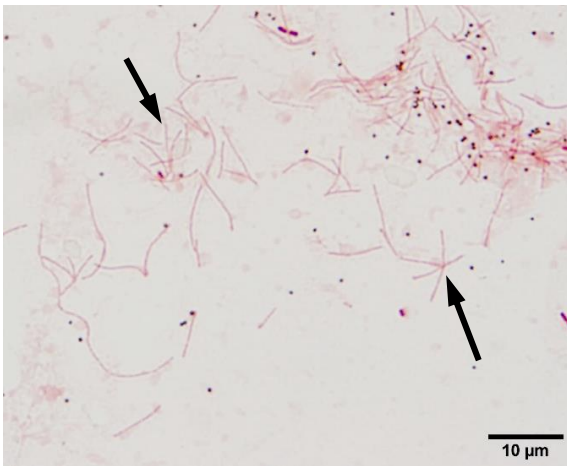


Wet mount of the saddle-back lesion revealed clumps of long rod-shaped bacteria

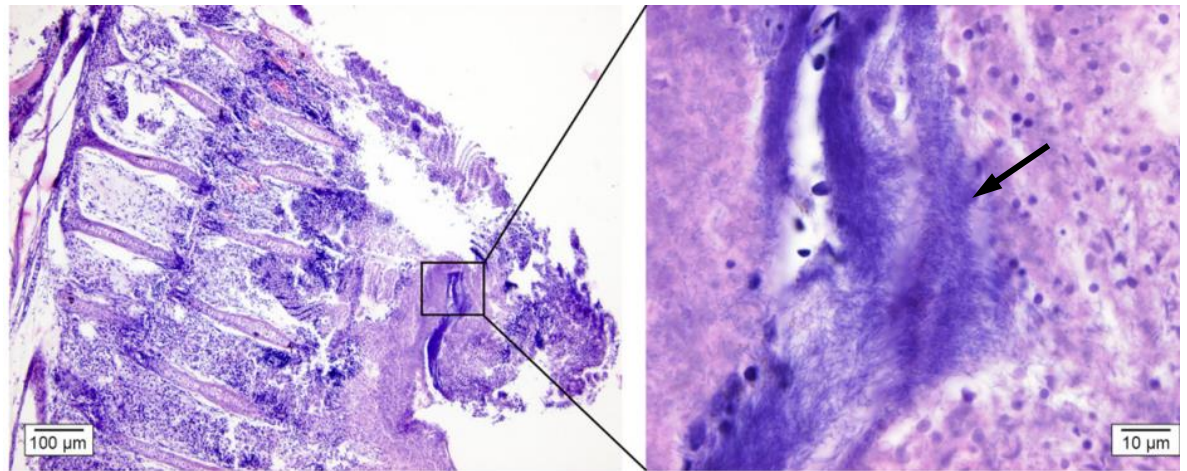
Columnaris

Diagnosics (Level 2)

- Rapid Gram staining of smeared tissue e.g. gills, skin lesion
- Histopathology



Gram staining of smeared tissue

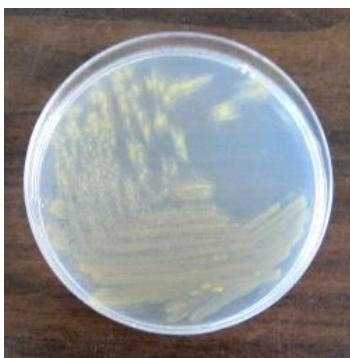


Histopathology of fish gills infected with *F. columnare* (Declercq et al. Vet Res 2013, 44:27)

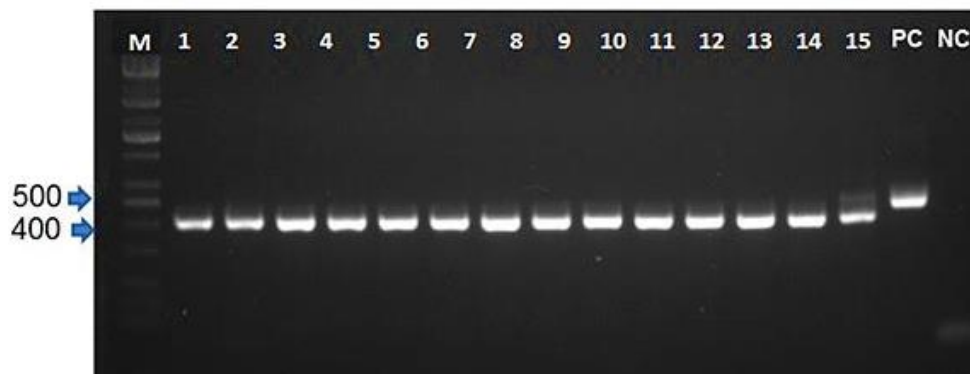
Columnaris

Diagnostics (Level 3)

- Specific PCR (for either fish tissue or pure isolate)
 - ✓ Welker et al. 2005
 - ✓ Triyanto et al. (1999)
 - ✓ Darwish et al. (2004)
- Bacterial culture + sequencing of 16S rRNA
 - Anaker and Ordal's medium (AOA)
 - Modified Shield medium



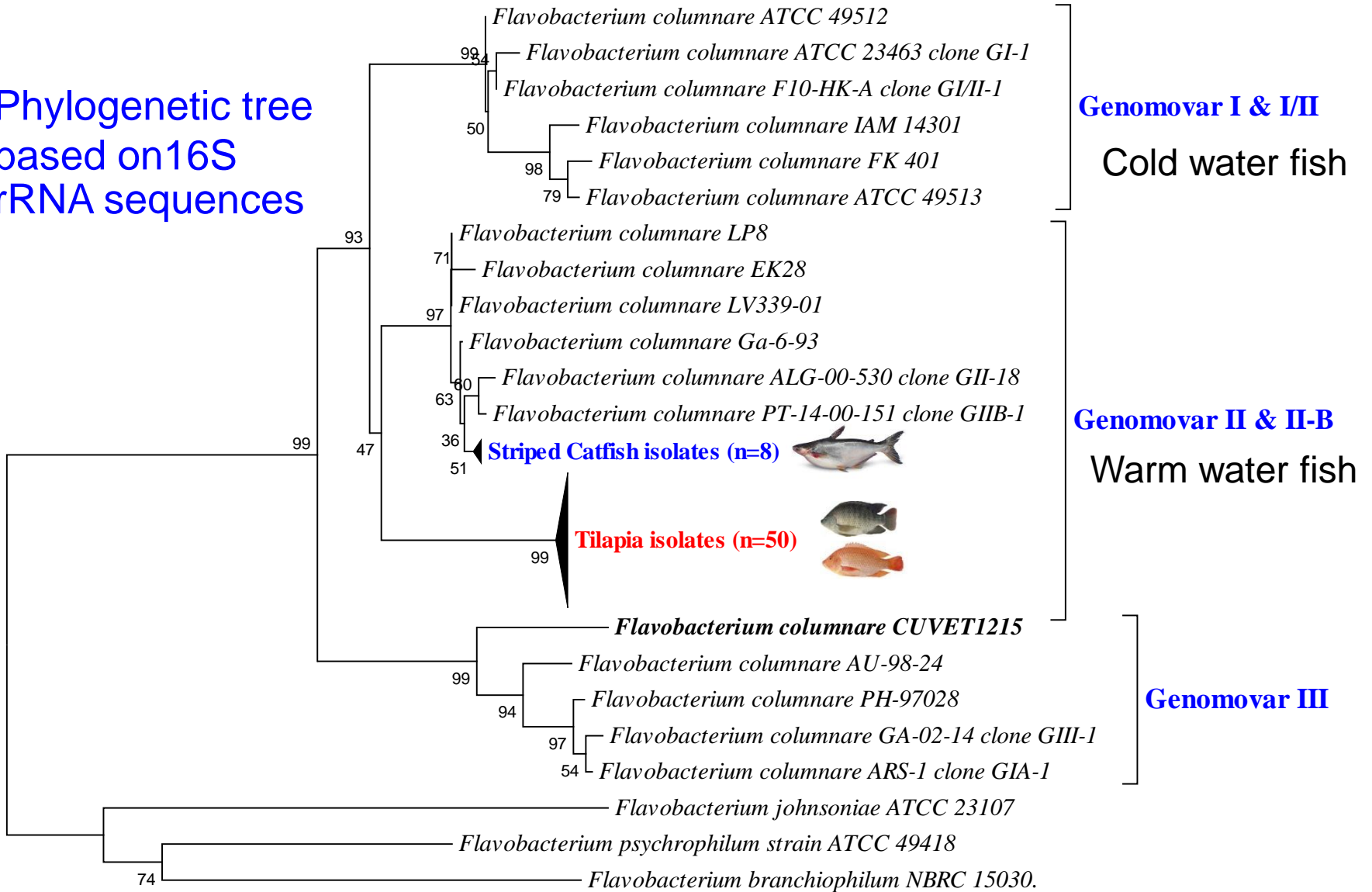
F. columnare on AOA medium



F. columnare-specific PCR according to Welker et al. 2005

Columnaris

Phylogenetic tree based on 16S rRNA sequences



0.01

Columnaris

Mortality: variable, reached up to 100% in challenge experiments

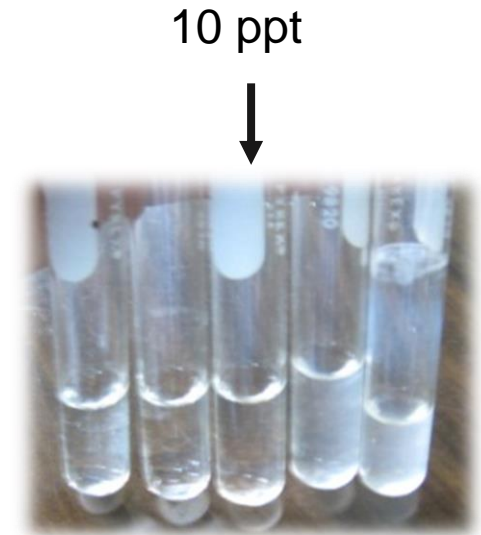
Geographical Distribution: worldwide

Risk factors

- Disease usually occurs after transportation
- Cage culture is more susceptible than pond culture

Prevention

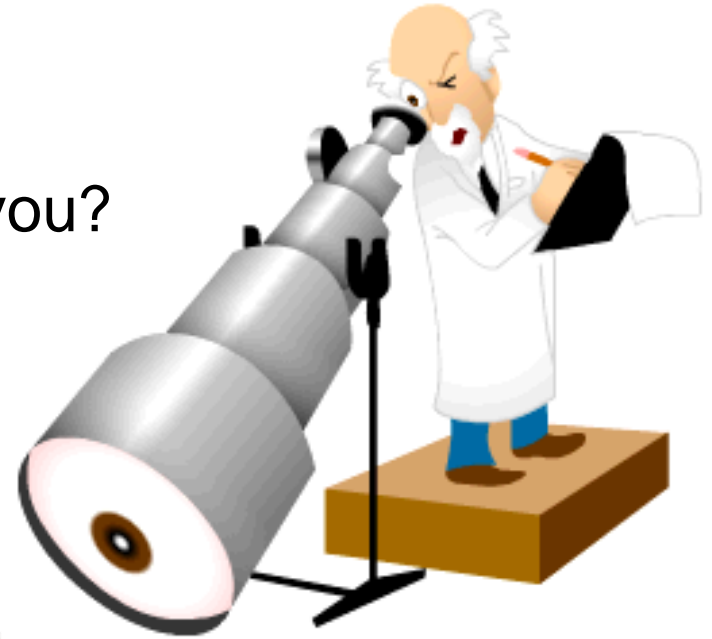
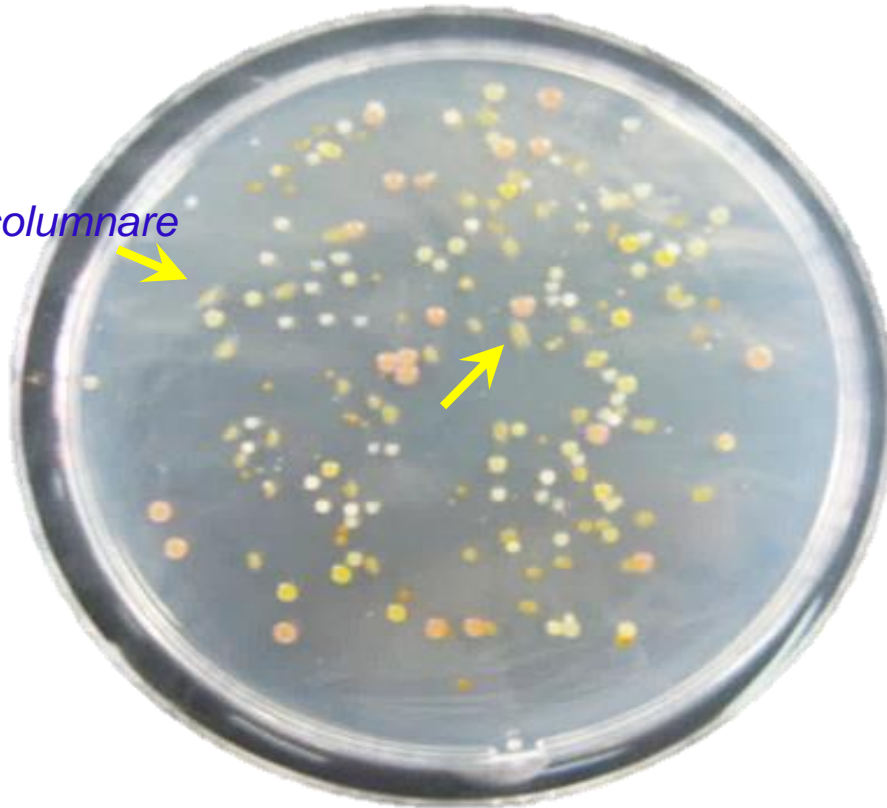
- Sodium chloride 10 ppt can prevent/control the disease
- No commercial vaccine available for tilapia



Ignore OR Find Answer?

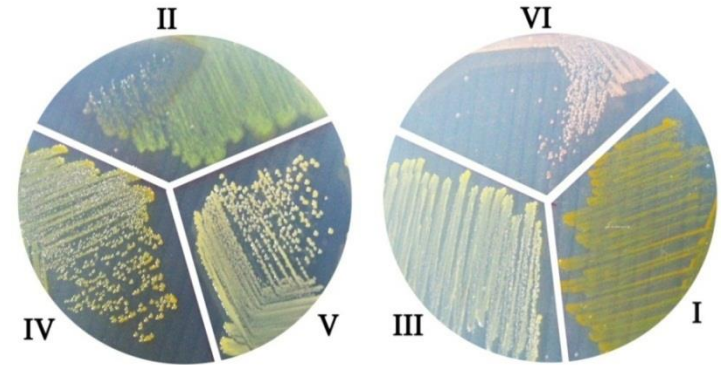
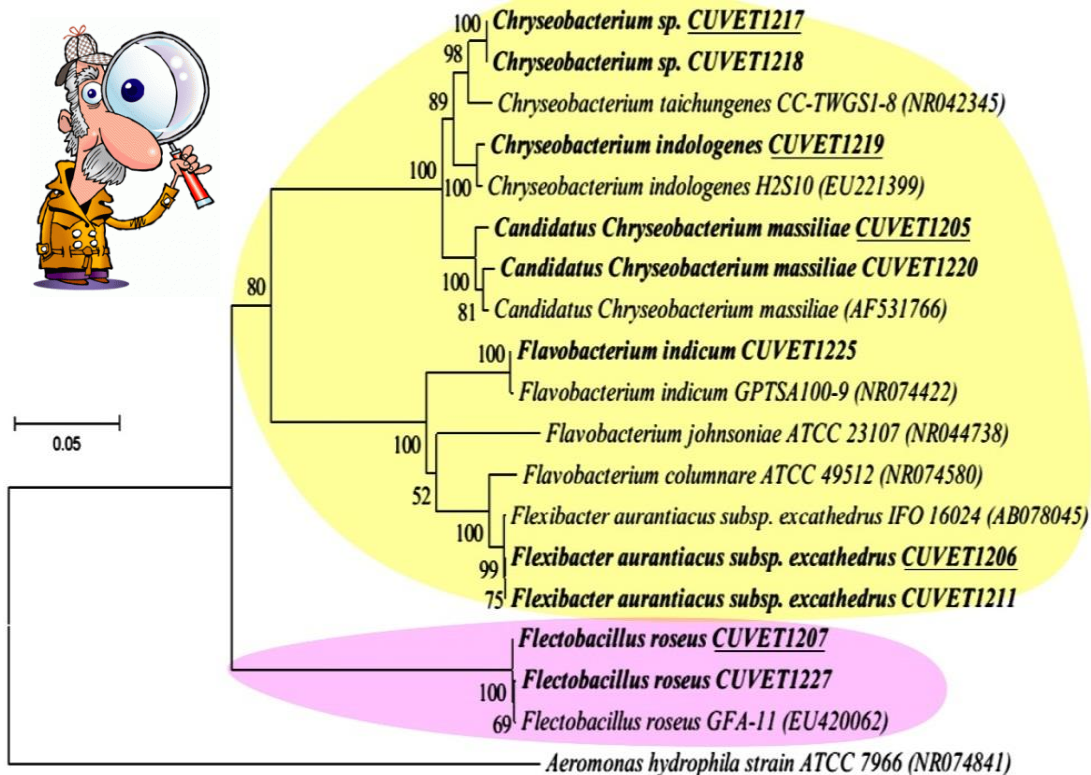
Who are you?

F. columnare



Columnaris

Diversity of *Non-F. columnare* associated with Columnaris diseased fish



- ✓ Most bacteria first found in tilapia
- ✓ Experimental challenge (I.M.) showed 0-20% mortality
- ✓ May serve as opportunistic pathogens

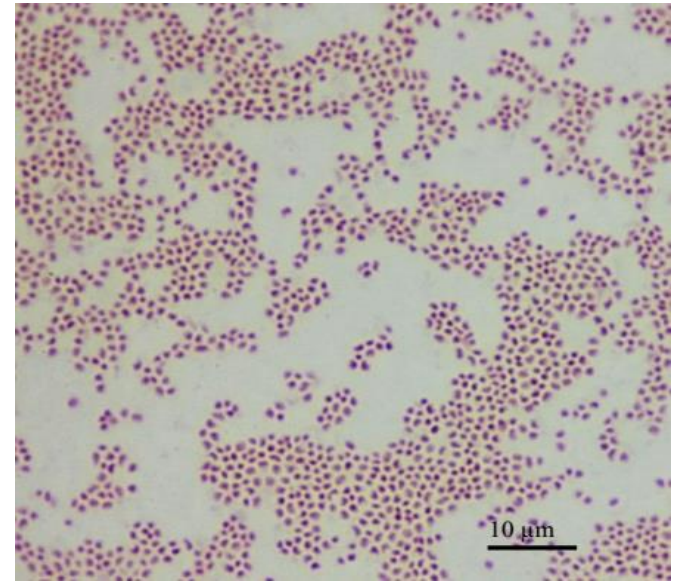
Phylogenetic analysis based on 16S rRNA

Francisellosis

Francisellosis of tilapia

❖ Causative agent

- *Francisella noatunensis* subsp. *orientalis* (*Fno*)
- Previously known as Rickettsia-like organism, RLO or Piscirickettsia-like organism
- Fastidious intracellular bacterium
- Gram negative, oval shape
- Require cysteine for growth
- Optimum temperature for *Fno* (25-28 °C)

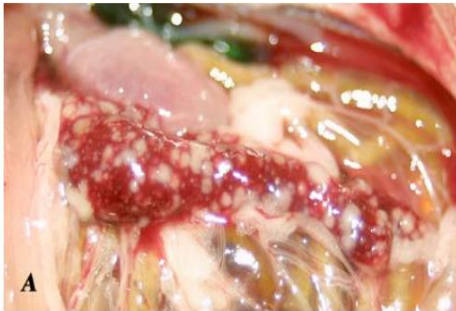


Fno, Gram staining (Photo: VV Nguyen)

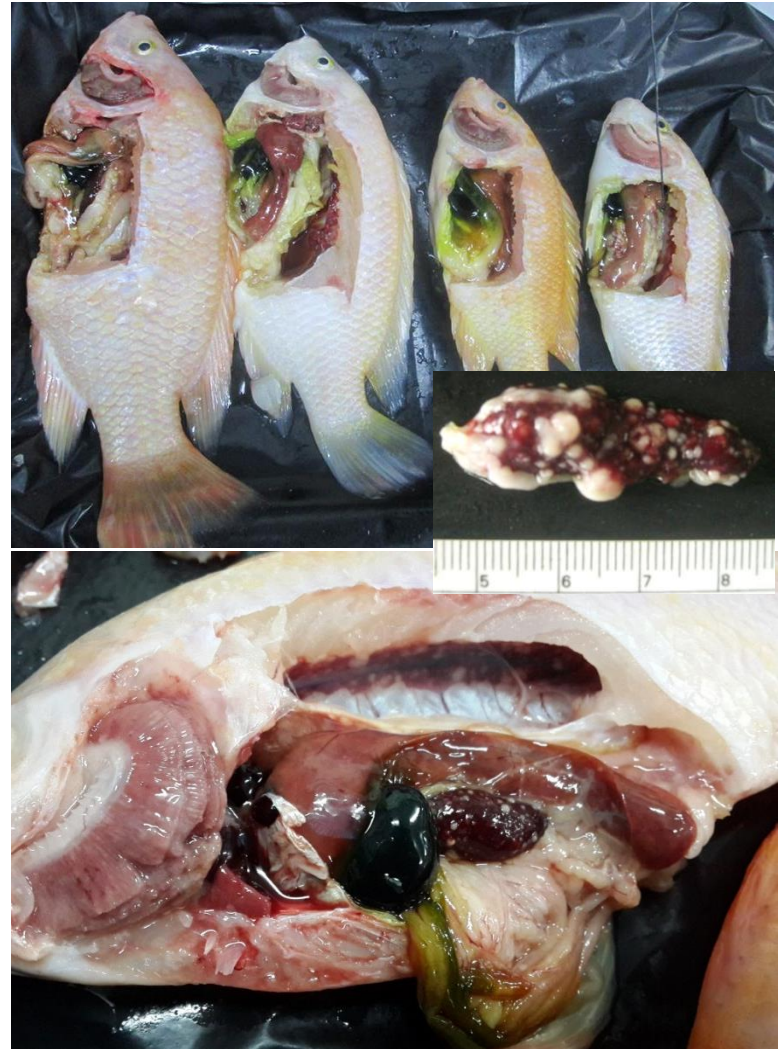
Francisellosis of tilapia

❖ Clinical signs (Level 1)

- Pale body
- White spots/white nodules on the spleen, head kidney, trunk kidney, gills
- Lost of appetites



Images by Michael J. Mauel

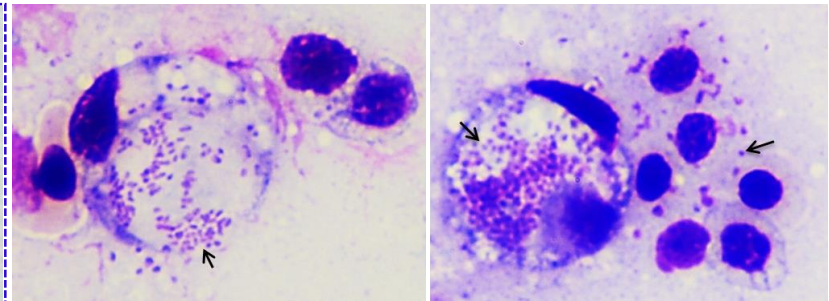
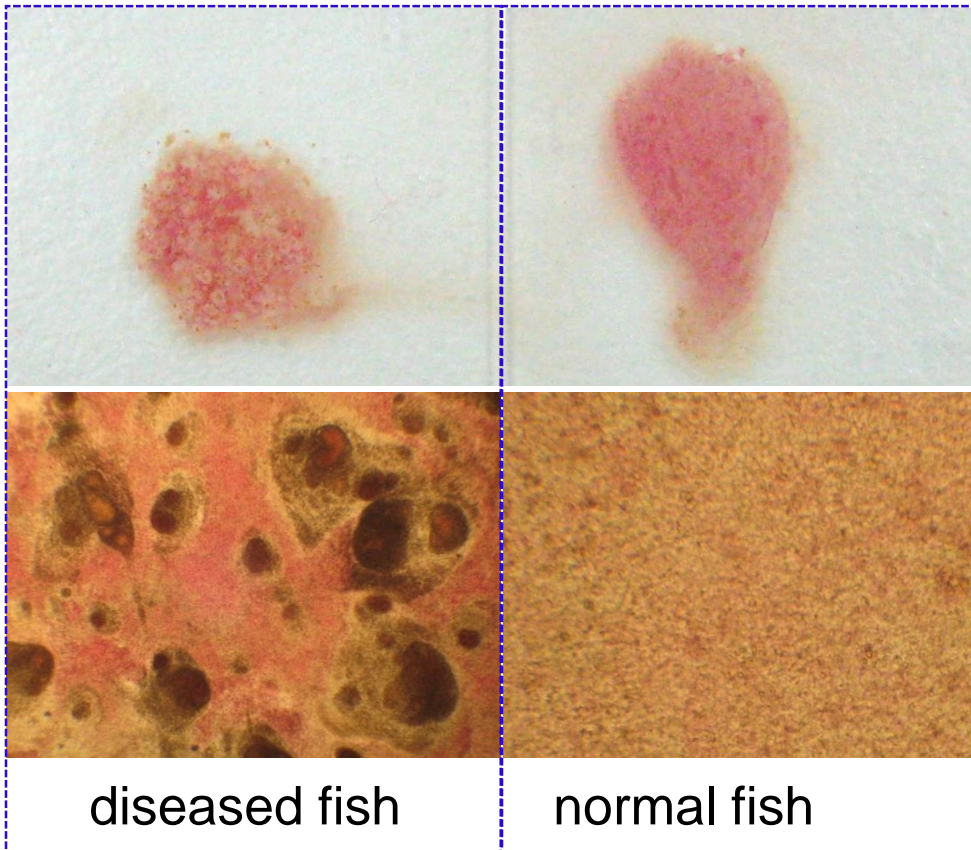


Photographs were taken in conjunction with the outbreaks described in Nguyen et al. 2015. Aquac Res. ²⁴

Francisellosis of tilapia

Diagnostics (Level 2)

- Wet-mount (e.g. spleen, gills)
- Rapid staining

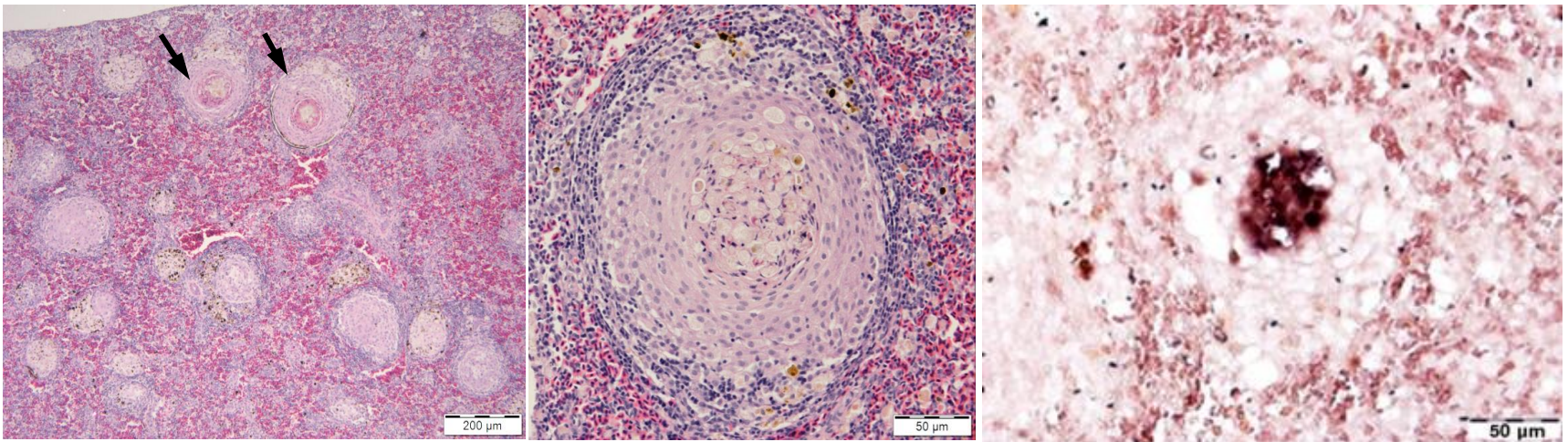


Rapid staining of smeared-head kidney with **Giemsa** revealed presence of both intra- and extra-cellular bacteria (Nguyen et al. 2015)

Francisellosis of tilapia

Diagnosics (Level 2)

- Histopathology: presence of granulomas in multiple internal organs
- *In situ* hybridization (ISH) using Fno-specific probe



Micrographs of H&E stained sections of the spleen showed typical granulomas.

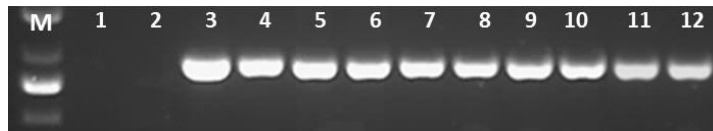
ISH using specific probe revealed location of Fno in the spleen

Francisellosis of tilapia

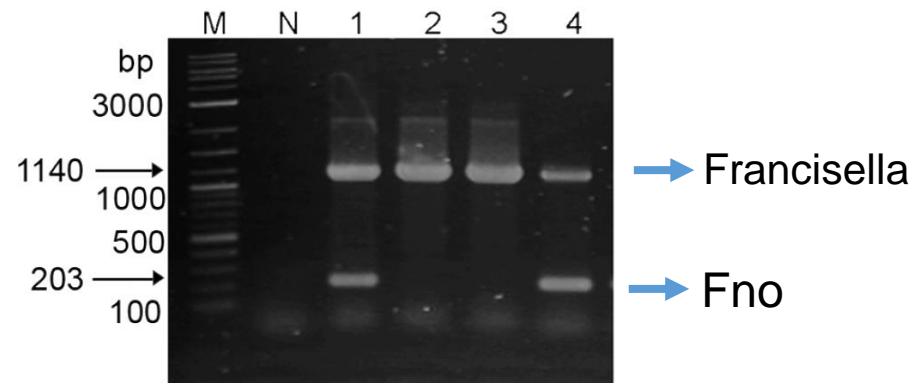
Diagnostics (Level 3)

❖ Detect Fno from fish tissue

- Genus specific PCR (Forsman et al. 1994)
- Real-time PCR (Duodu et al. 2012)
- ISH, genus-specific (Hsieh et al. 2007)
- Immunohistochemistry (Soto et al. 2012)
- Duplex PCR and ISH (Dong et al. 2016)
- Colorimetric LAMP (Pradeep et al. 2016)
- Recombinase polymerase amplification (RPA) (Shahin et al. 2018)



Genus-specific PCR (Forsman et al. 1994)



Duplex PCR (Dong et al. 2016)

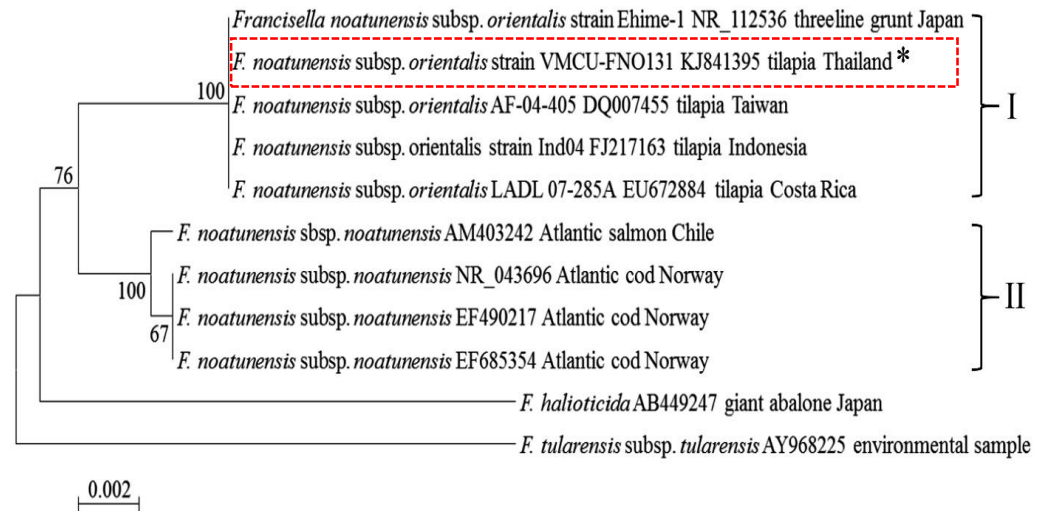
Francisellosis of tilapia

Diagnostics (Level 3)

- Culture + specific PCR
- Culture + sequencing of 16S rRNA



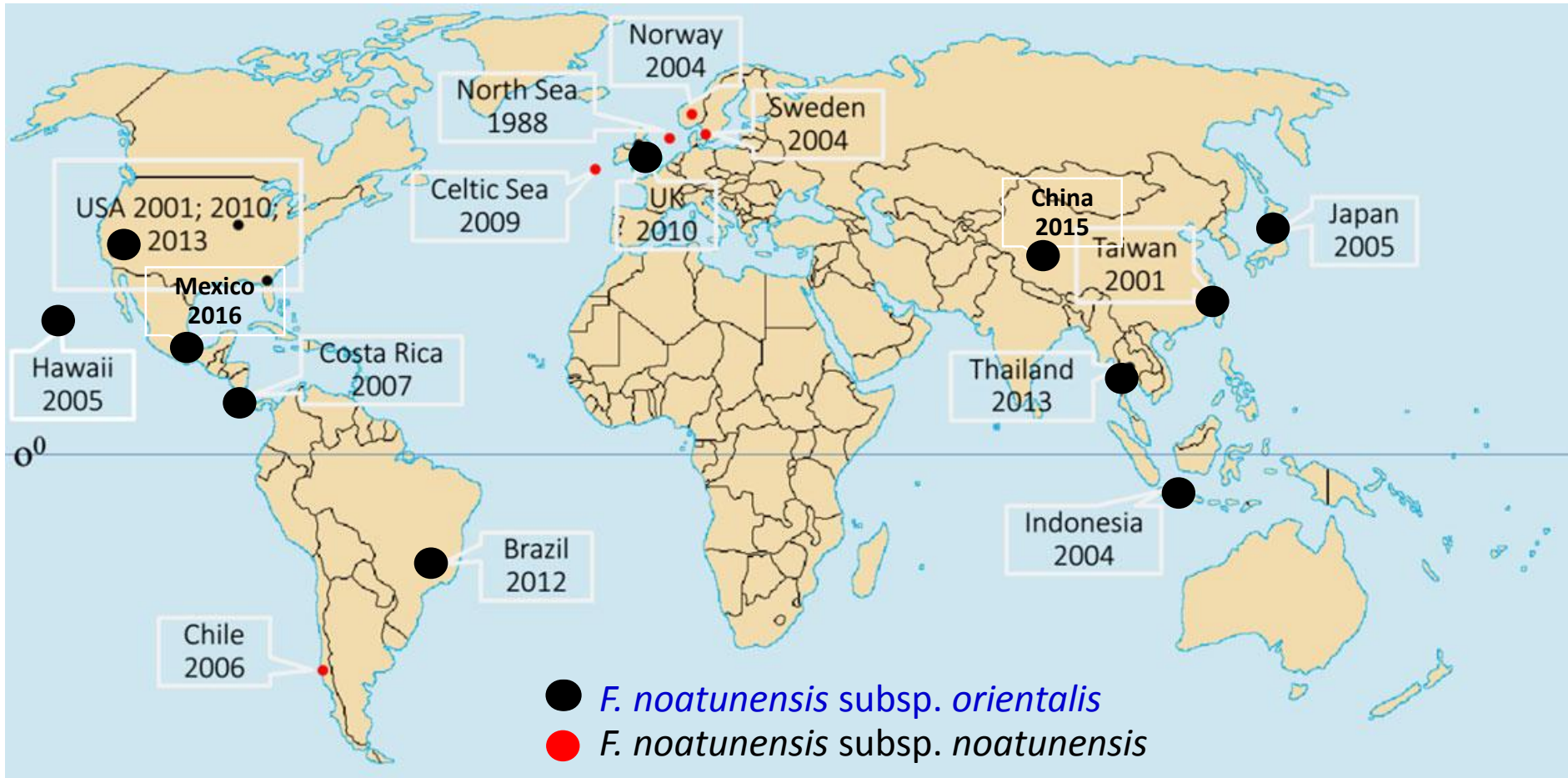
Fno on Cysteine heart agar (CHA)
(Photo: VV Nguyen)



Nguyen et al. 2015. Aquac Res.

Francisellosis of tilapia

Geographical distribution



Adapted from Nguyen et al. 2015 Aquac Res. doi:10.1111/are.12802

Francisellosis of tilapia

Mortality: 40-50%

Risk factors:

- Disease outbreaks were associated with cool season (<28⁰ C)
- Very chronic, disease outbreaks depend on environmental factors

Prevention/treatment

- ❖ Use Fno-free fish
- ❖ Antibiotics have been used for treatment
- ❖ Temperature >30⁰ C, no mortality occur
- ❖ Commercial vaccine is NOT available

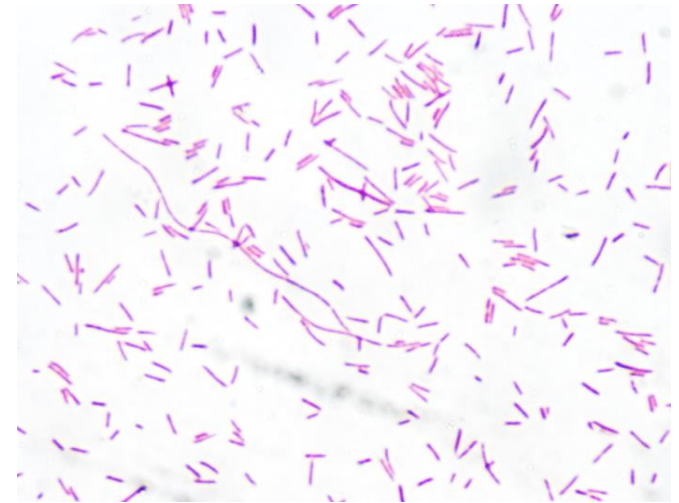
Edwardsiellosis of tilapia

Edwardsiellosis of tilapia

Causative agent:

- *Edwardsiella ictaluri*
- *Edwardsiella tarda*
- *Edwardsiella anguillarum*
- *Edwardsiella piscicida* (?)

- Pinpoint colonies on culture media
- Rod-shaped Gram negative
- Oxidase negative



Rod-shaped Gram negative

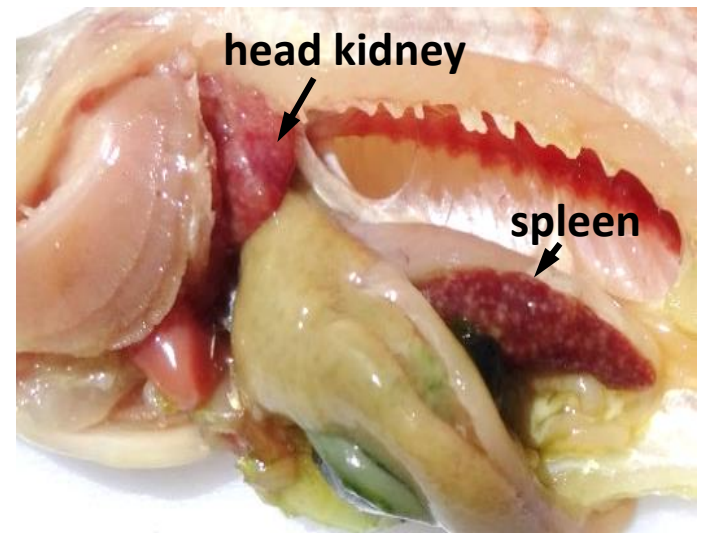
Edwardsiellosis caused by *E. ictaluri*

- Common in catfish
- Not common in non-catfish
- Does not kill tilapia in striped catfish ponds (personal observation)
- 2012: first report of *E. ictaluri* in Nile tilapia in Western Hemisphere (Soto et al. 2012)

Edwardsiellosis of tilapia

Recent cases in Southeast Asia

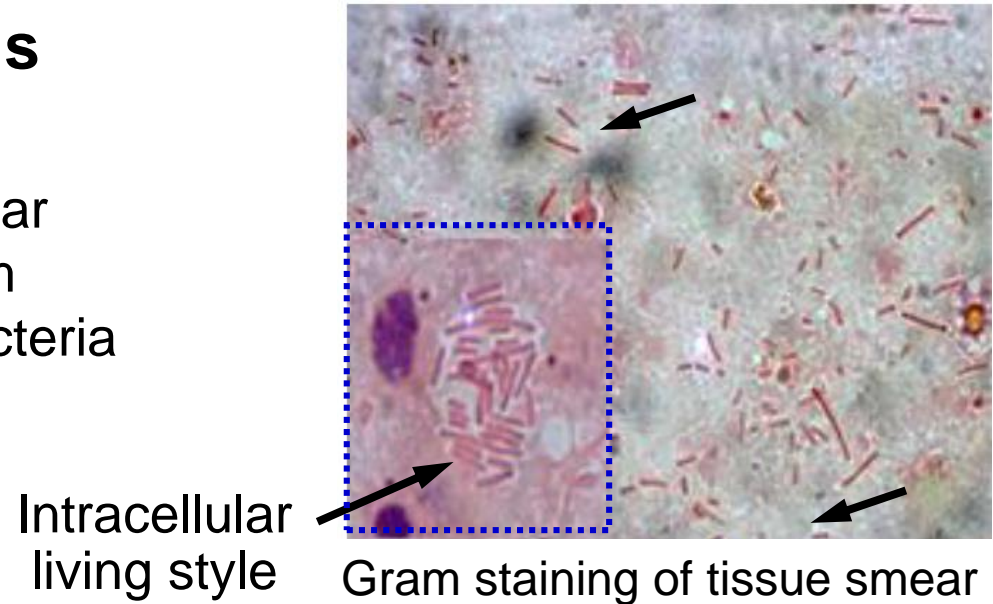
- Red tilapia juveniles
- Killed 40-50% fish in the first month after stocking
- Presence of white spots in multiple internal organs
- Presumptive diagnosis based on clinical sign (**Level 1**): **Francisellosis**
- PCR negative for *Fno*



Edwardsiellosis of tilapia

Presumptive diagnosis (Level 2)

- ❖ Gram stained tissue smear revealed numerous Gram negative, rod-shaped bacteria

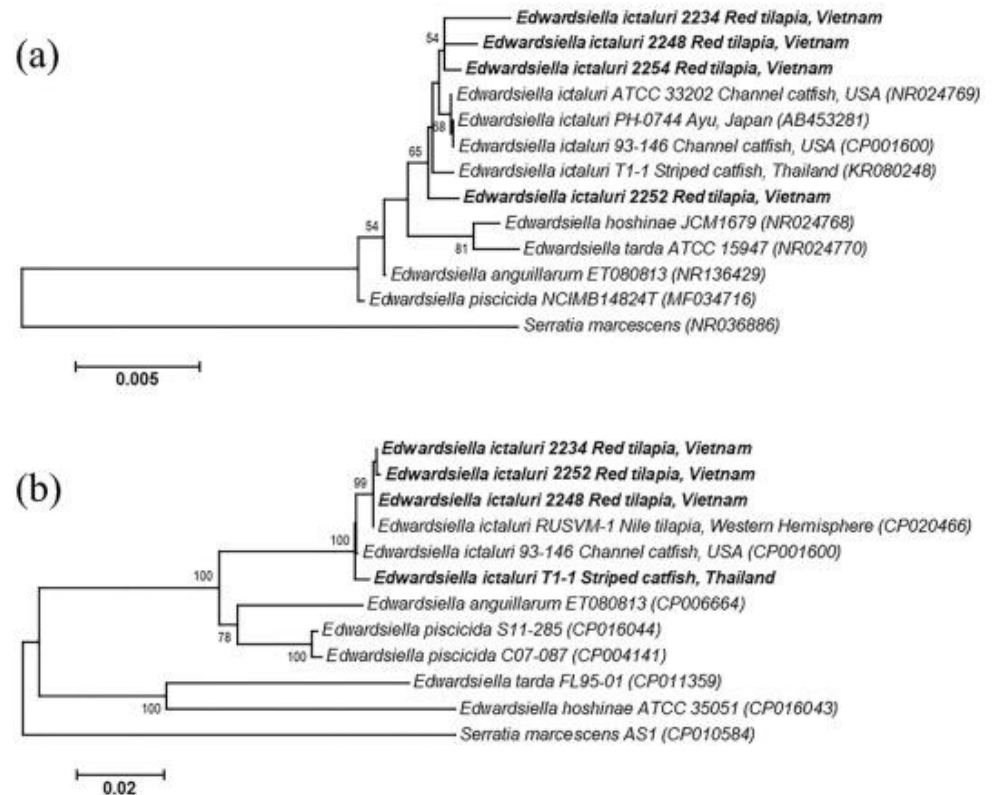
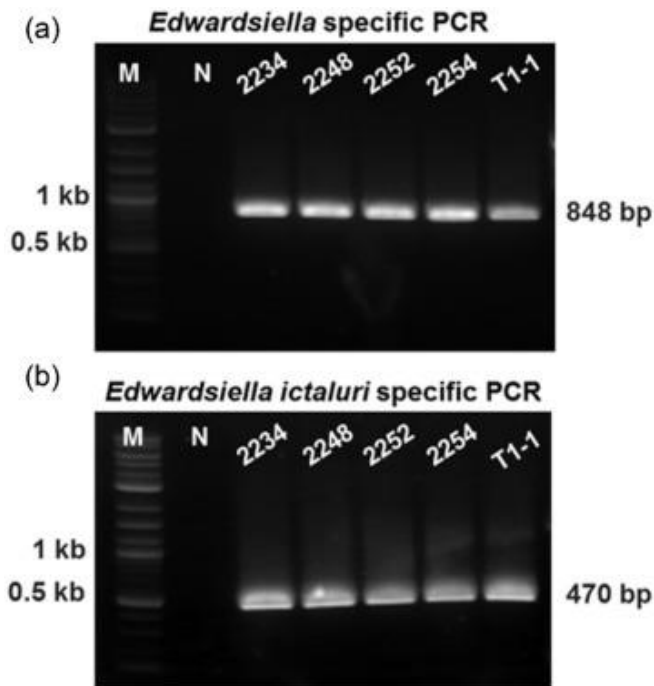


- ❖ **Bacterial isolation:** pure pinpoint colonies on TSA or NA
- ❖ Gram negative, rod-shaped bacteria



Edwardsiellosis of tilapia

❖ Confirmed diagnosis (Level 3)

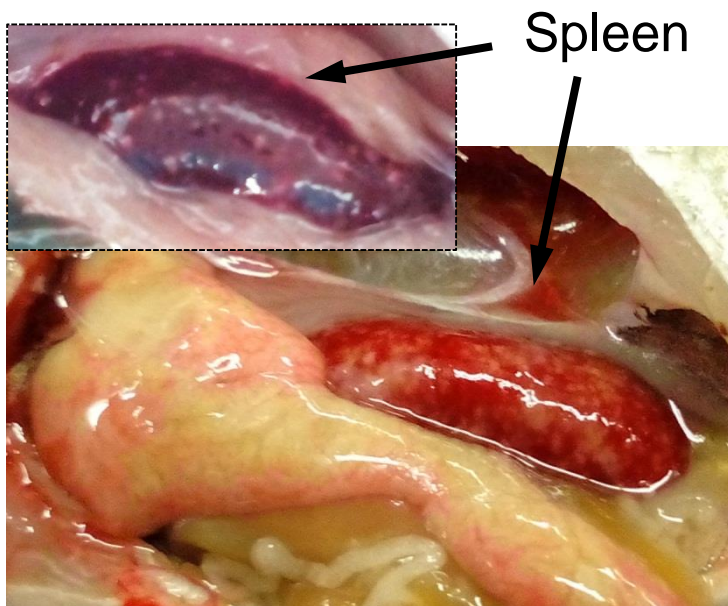


Phylogenetic tree based on 16S rRNA (a) and *gyB* (b)

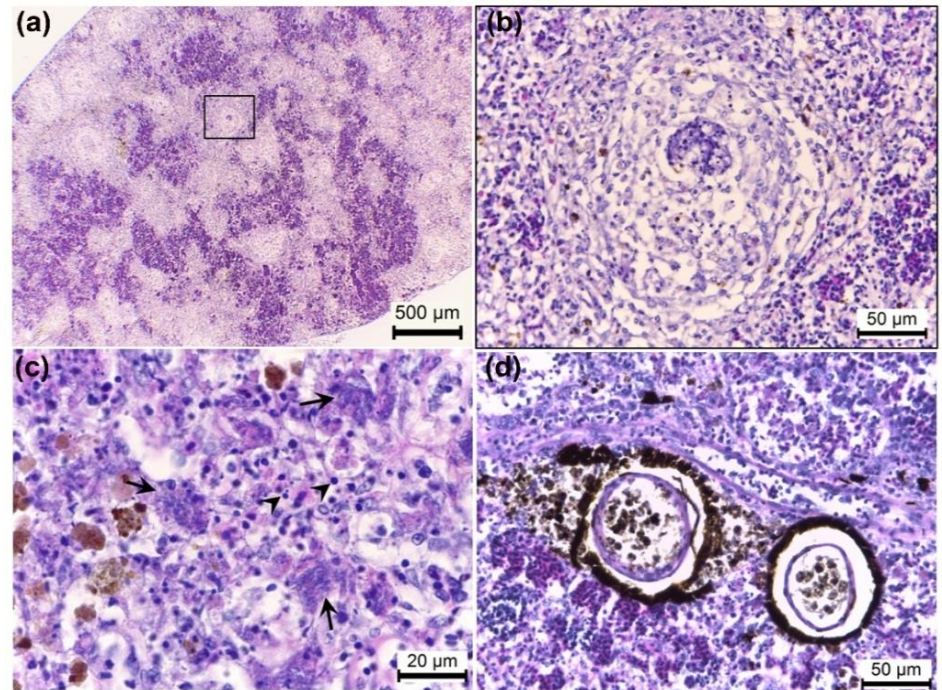
Edwardsiellosis of tilapia

Challenged experiments fulfilled Koch's postulates

- Fish reproduce the same clinical signs
- 95-100% mortality in 3-9 days (dose-dependent)



Experimental fish

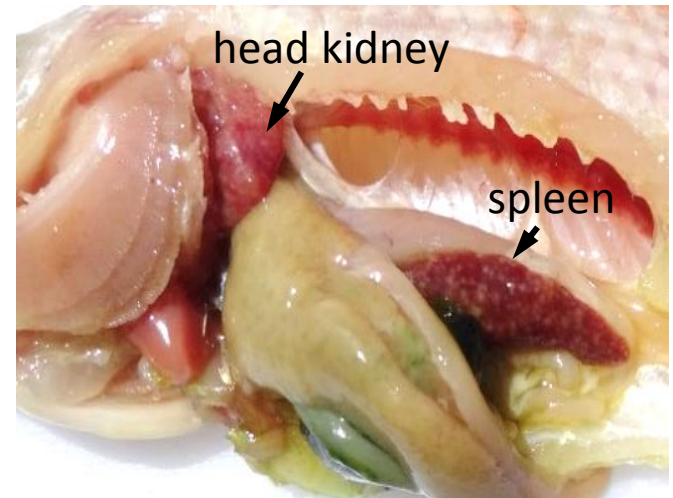


Histopathological features of edwardsiellosis in the experimental fish

Edwardsiellosis of tilapia

Comments

- *E. ictaluri* is an emerging pathogen of tilapia aquaculture in Southeast Asia
- *E. ictaluri* infections in tilapia may have been overlooked due to similar clinical signs between Francisellosis & Edwardsiellosis
- Should be put on disease watchlist in tilapia farming countries



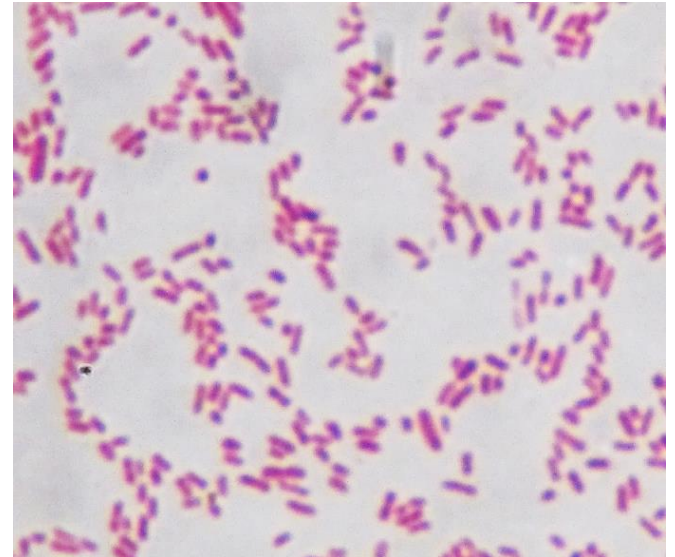
Same same...but different...

Hemorrhagic septicemia caused by motile aeromonads

❖ Causative agent

- *Aeromonas hydrophila*
- *Aeromonas veronii*
- *Aeromonas jandaei*
- *Aeromonas shuberti*
- *Aeromonas dhakensis*

- Gram negative bacteria
- Rod or oval shaped
- Non-spore forming
- Oxidase (+), catalase (+)



Aeromonas sp. (Gram stain)

Hemorrhagic septicemia caused by motile aeromonads

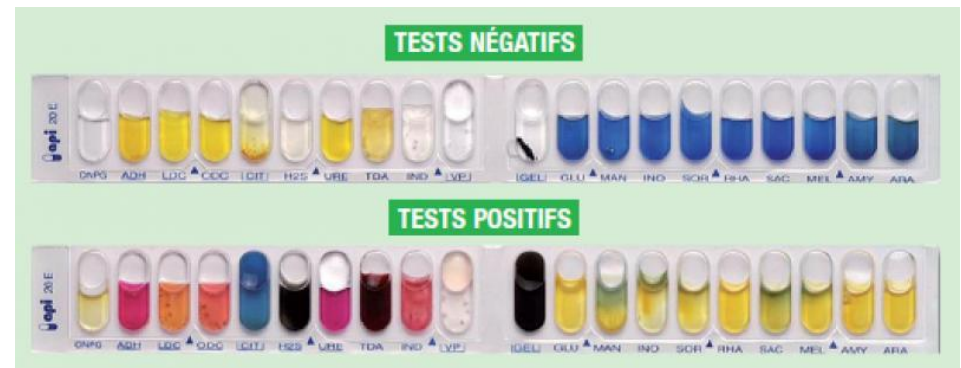
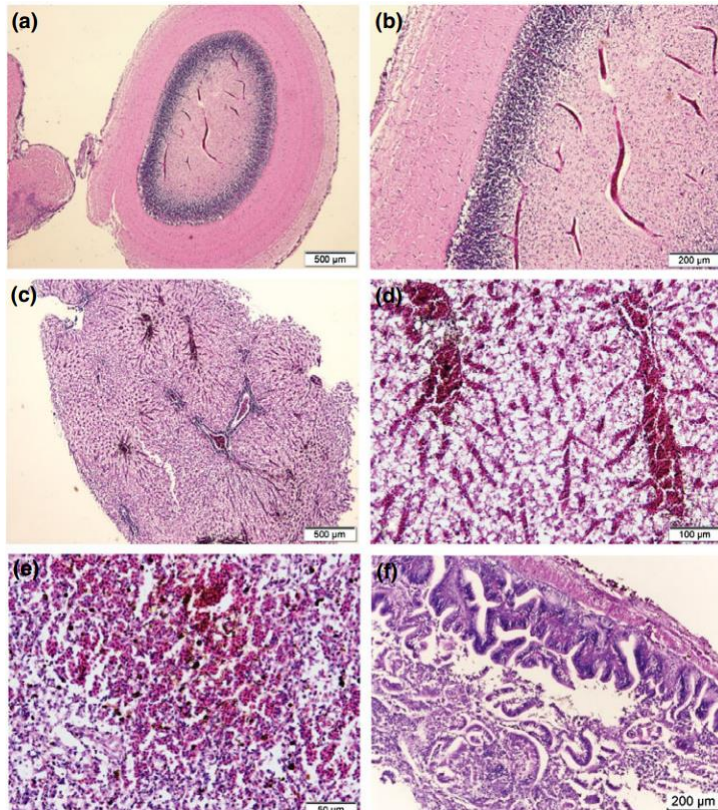
❖ Clinical signs (Level 1)



Hemorrhagic septicemia caused by motile aeromonads

Diagnostics (Level 2)

- Histopathology
- Bacterial culture + biochemical tests

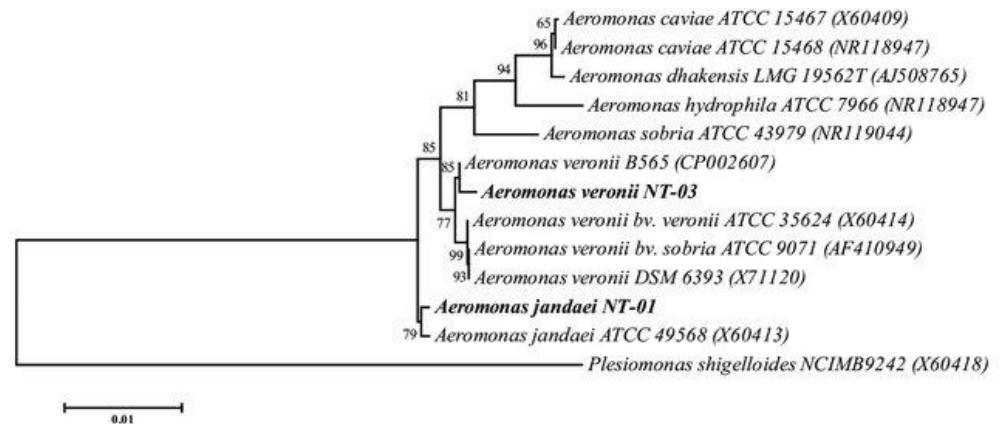
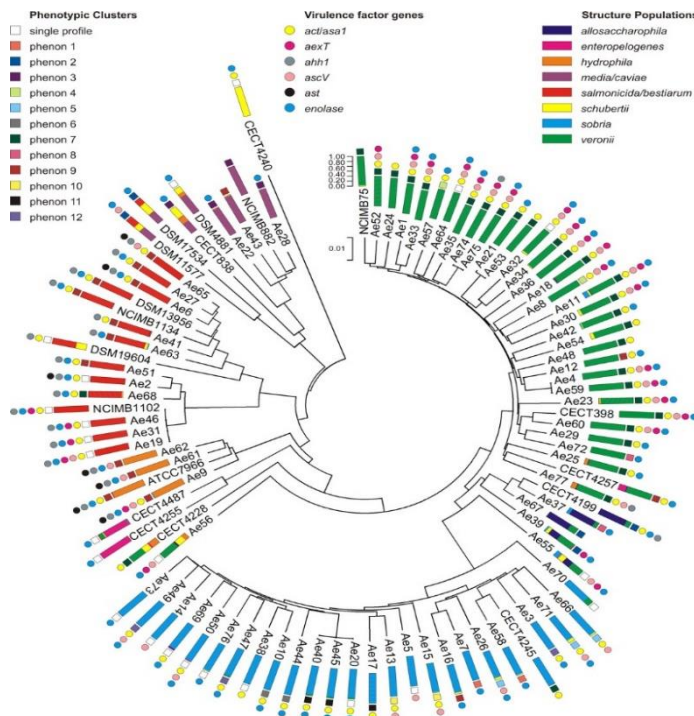


API 20E kit

Hemorrhagic septicemia caused by motile aeromonads

Diagnostics (Level 3)

- ❖ Specific PCR (for either fish tissue or pure isolate)
 - Misidentification is common in *Aeromonas* group
 - Bacterial culture + sequencing of 16S rRNA or MLST is highly recommended



MLST (Martino et al. 2011)

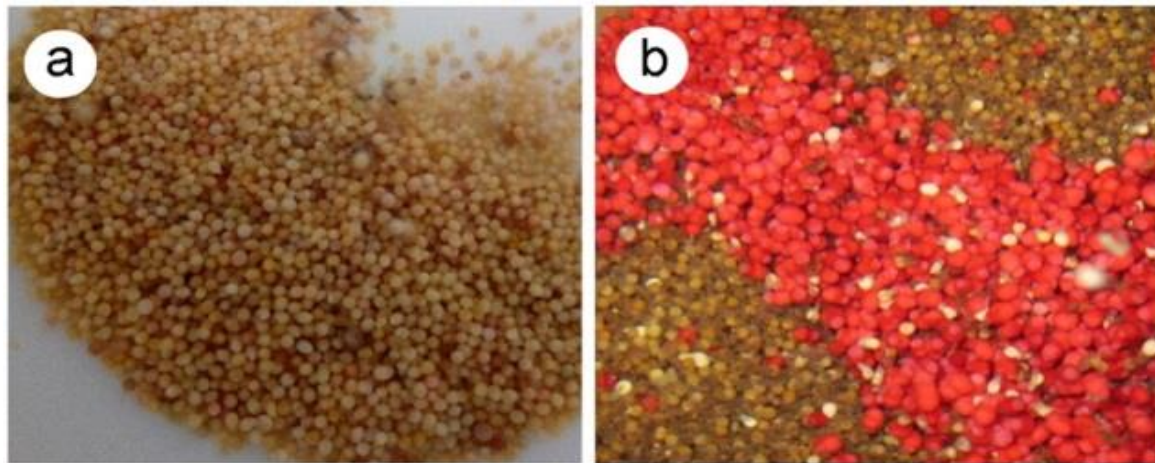
Hahellosis (red egg disease)

Hahellosis/Red egg disease

Causative agent

- ❑ *Hahella chejuensis*
- ❑ Gram negative, rod-shaped, red pigmented bacteria
- ❑ Marine bacterium

Diagnostics (Level 1)



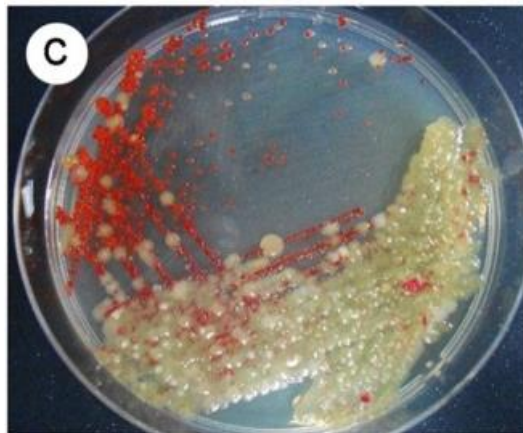
Normal eggs

Red eggs

Hahellosis/Red egg disease

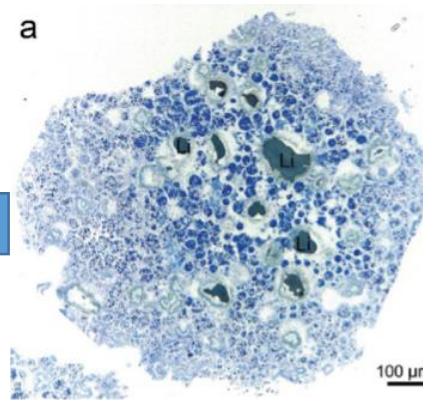
Diagnostics (Level 2)

- Histology
- Bacterial isolation

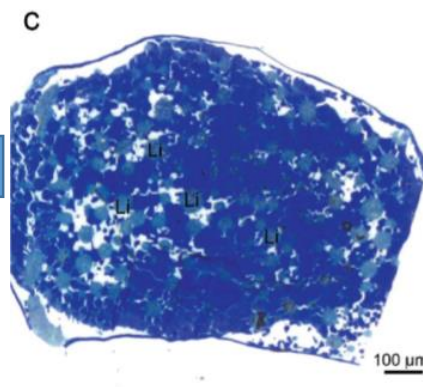


Bacterial isolation using TSA

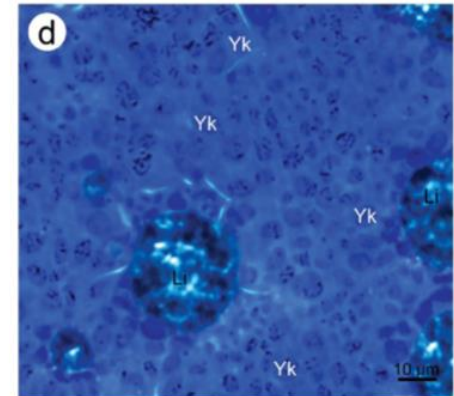
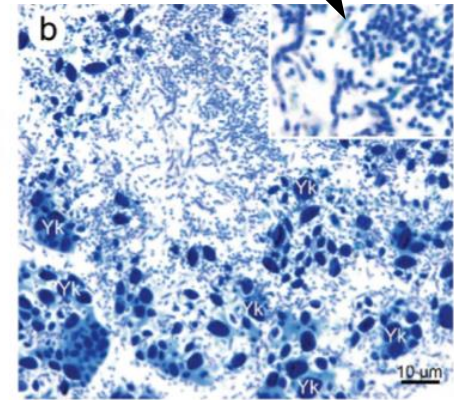
Red egg



Normal egg



Rod bacterial cells

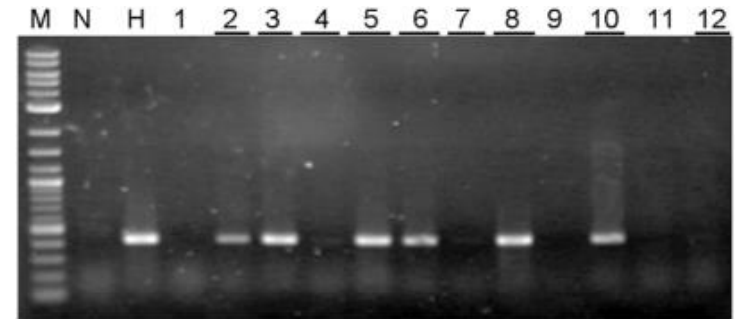
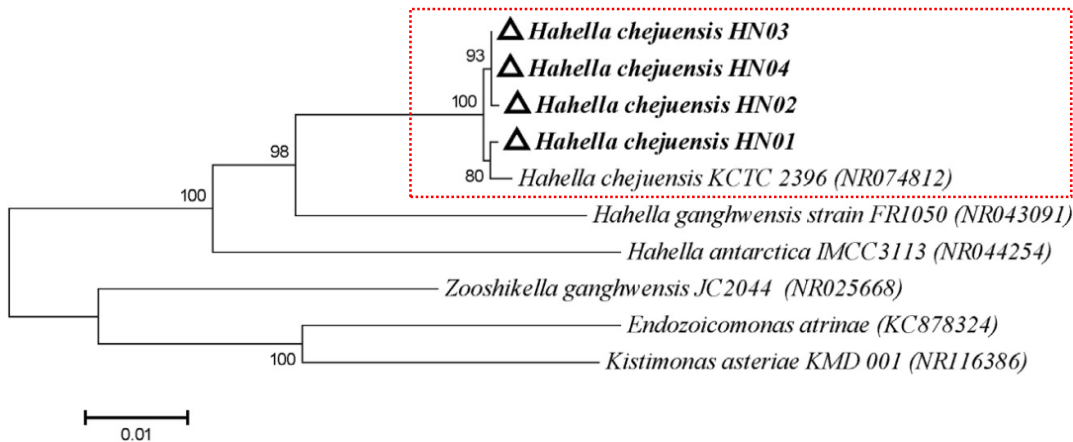


Semi-thin section (1 µm), stained with toluidine blue

Hahellosis/Red egg disease

Diagnostics (Level 3)

- ✓ Sequencing of 16S rRNA
- ✓ Genus & species specific PCR (Senapin et al. 2016)



Specific PCR methods were developed targeting 16S rRNA

Red pigmented bacteria was identified using 16S rRNA

Hahellosis/Red egg disease

Geographical Distribution

- ✓ Tilapia hatcheries in Thailand since 2010
- ✓ Recently found in rabbitfish in Vietnam (unreported)

Mortality: 10-50%

Risk factors:

- ✓ Occur during cold season (<24 °C)
- ✓ Occurred in hatcheries using 7 ppt NaCl water

Hahellosis/Red egg disease

Disease control

- ✓ Reduce salinity from 7 ppt to 4 ppt
- ✓ Expose sand from the filter system to sunlight
- ✓ Wrap the hatcheries with plastic to increase temperature (30 °C)



- Reduction of loss: ~ \$ 600,000 /year
- Calculation based on 30% mortality (range from 10-50%)

Epitheliocystis Disease

- Causative agent: *Chlamydia-like* organisms (CLOs)
- Affect mainly gills
- Sometime associated with mortality in tilapia fry and fingerlings
- Diagnostics: wet-mount of gills filaments or histology

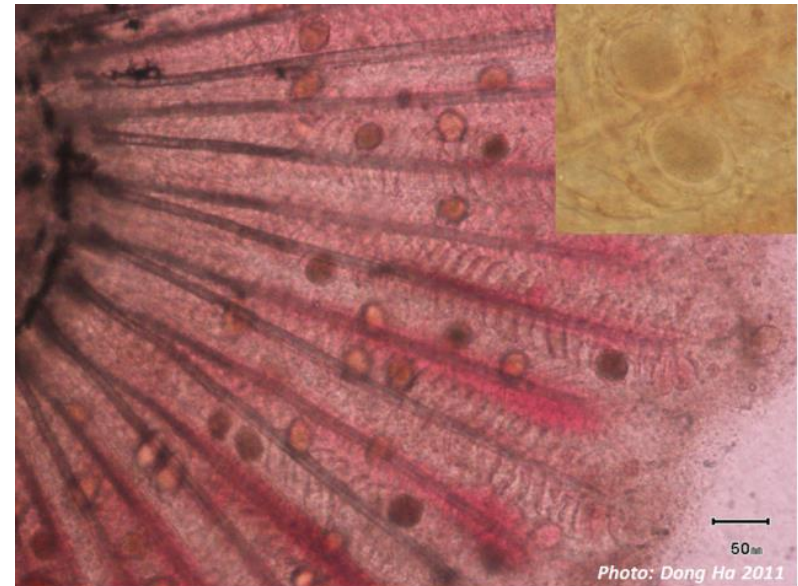
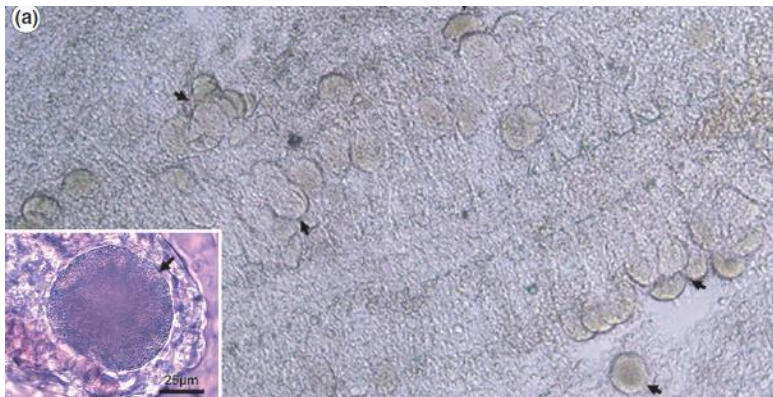
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Short communication

A survey of epitheliocystis disease in farmed Nile tilapia (*Oreochromis niloticus* Linnaeus, 1758) in Brazil

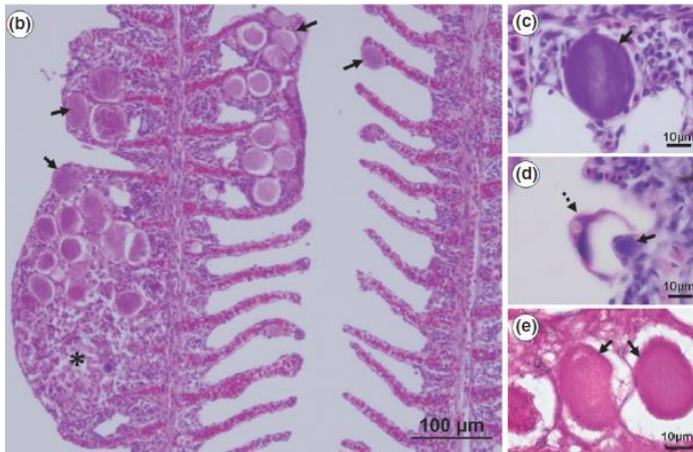
By S. B. Pádua¹, R. N. Menezes-Filho¹, M. L. Martins², M. A. A. Belo³, M. M. Ishikawa⁴, C. A. Nascimento⁵, K. C. Saturnino⁶ and J. R. Carrijo-Mauad⁷



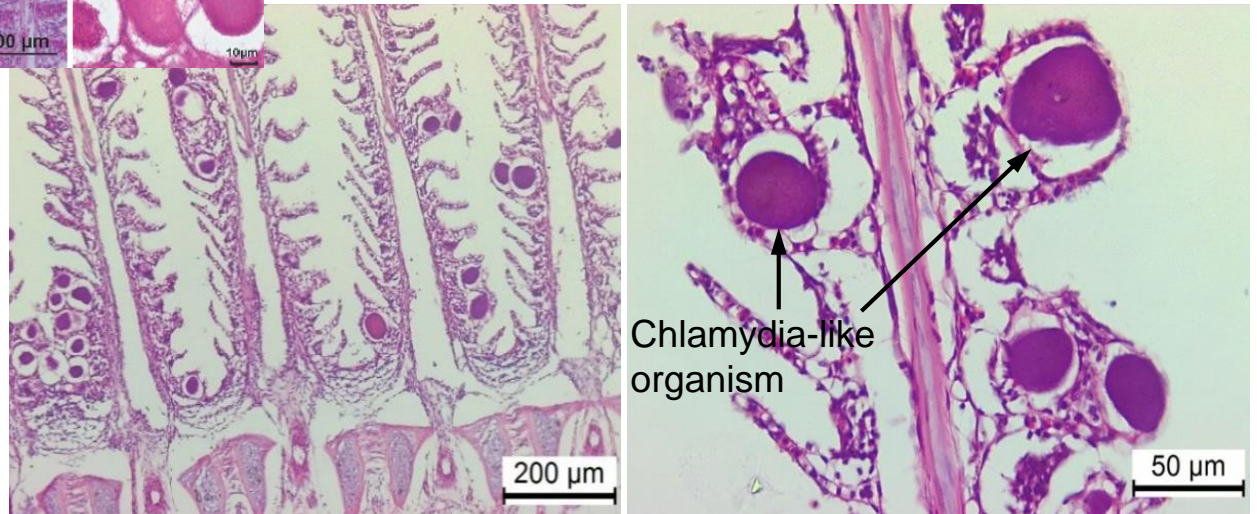
Fresh mount showing heavy infection of CLOs

Epitheliocystis Disease

- Diagnostics: histopathology



Nile tilapia, Brazil
(Padua et al. 2014)



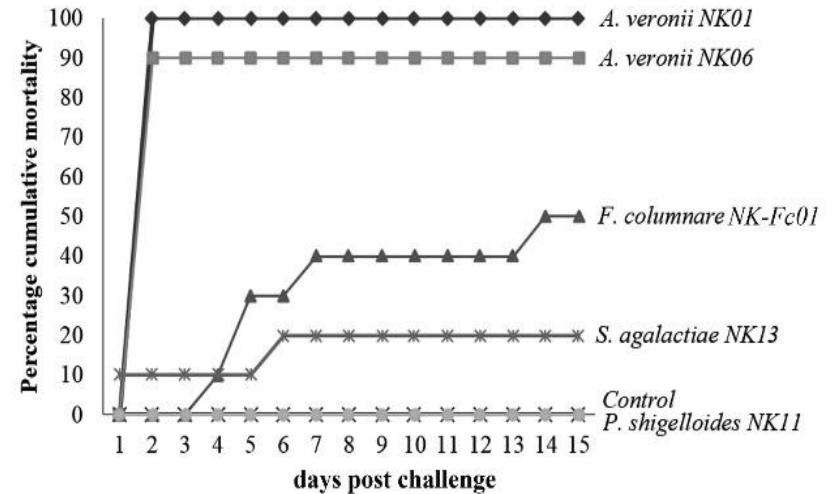
Red tilapia, Thailand (HT Dong)

THINK
DIFFERENTLY



Single infection vs. Multiple infections

Miscellaneous Disease in Tilapia



- ✓ Concurrent infections of 5 bacteria and a virus in cultured tilapia farms
- ✓ Each fish was infected with 2-4 pathogens
- ✓ ***A. veronii*** and ***F. columnare*** were most dominant and exhibited high virulence

Miscellaneous Disease in Tilapia

JOURNAL OF FISH DISEASES

Original Article

Natural coinfection by *Streptococcus agalactiae* and *Francisella noatunensis* subsp. *orientalis* in farmed Nile tilapia (*Oreochromis niloticus* L.)

G B N Assis, G C Tavares, F L Pereira, H C P Figueiredo, C A G Leal ✉

First published: 04 May 2016 | <https://doi.org/10.1111/jfd.12493> | Cited by: 9



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Aquaculture

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Short communication

A case of natural co-infection of Tilapia Lake Virus and *Aeromonas veronii* in a Malaysian red hybrid tilapia (*Oreochromis niloticus* × *O. mossambicus*) farm experiencing high mortality

M.N.A. Amal ^{a, f} ✉, C.B. Koh ^b, M. Nurliyana ^a, M. Suhaiba ^a, Z. Nor-Amalina ^c, S. Santha ^c, K.P. Diyana-Nadhirah ^c, M.T. Yusof ^d, M.Y. Ina-Salwany ^{c, f}, M. Zamri-Saad ^{a, f}

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Miscellaneous Disease in Tilapia



Coinfections of *Flavobacterium columnare* and *Francisella noatunensis* subsp. *orientalis*



F. columnare and *Saprolegnia* sp.

Thank you for your kind attention