



# Biological characteristics of tuna

Tuna and tuna-like species are very important economically and a significant source of food, with the so-called principal market tuna species - skipjack, yellowfin, bigeye, albacore, Atlantic bluefin, Pacific bluefin (those two species previously considered belonging to the same species referred as northern bluefin) and southern bluefin tuna - being the most significant in terms of catch weight and trade. These pages are a collection of Fact Sheets providing detailed information on tuna and tuna-like species.

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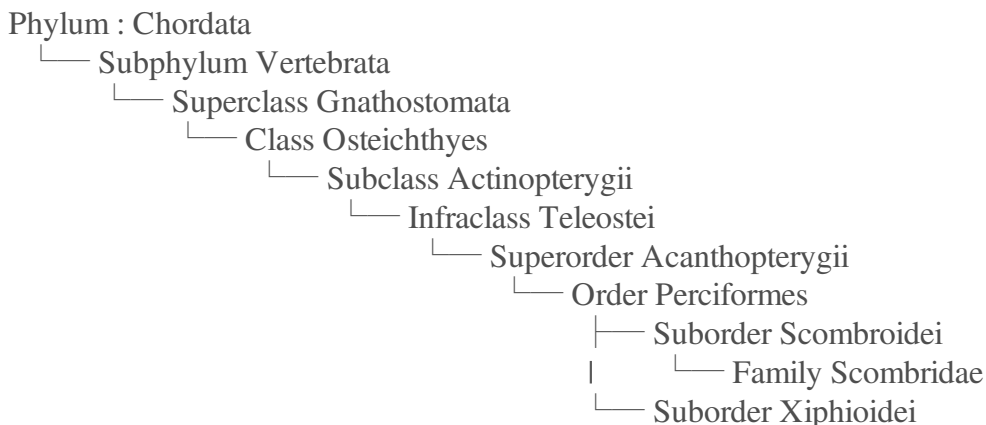
## Taxonomy and classification

[ **Family: Scombridae** ] : Scombrids

[ **Family: Istiophoridae** **Family: Xiphiidae** ] : Billfishes

### Upper systematics of tunas and tuna-like species

Scombrids and billfishes belong to the suborder of the Scombroidei which position is shown below:



## Scombrids

The scombrids belong to the family of the *Scombridae* which is comprised of 15 genera and 51 species. These species are assigned to 2 sub-families:

the *Gasterochismatinae* with only 1 species and the *Scombrinae* divided into 4 tribes:

- the mackerels (Scombrini)
- the seerfishes or Spanish mackerels (Scomberomorini)
- the bonitos (Sardini)
- the tunas (Thunnini)

The tribe *Thunnini* (tunas for ichthyologists) contains 14 species in 4 genera:

- Thunnus* (8 species)
- Katsuwonus* (1 species)
- Euthynnus* (3 species)
- Auxis* (2 species)

However, in several languages, the word "tuna" is used for members of both the tribe *Thunnini* and the tribe *Sardini* (8 species).

## Origin of the word "tuna"

The word "tuna" is applied to certain members of the family Scombridae, a group of marine fishes including tunas, bonitos, mackerels, seerfishes and the butterfly kingfish. However, for ichthyologists, tuna refers to any of the 14 species of the tribe Thunnini within the family Scombridae (Klawe, 1977). The word "tuna" seems to have come into use in the second half of the last century, but it is not clear why it replaced the older name "tunny". It may have been brought to southern California by fishermen originated from Europe, either from the Dalmatian coast of the former Yugoslavia or from the Iberian Peninsula (Klawe, 1976). The European names of tuna (thon in French, atún in Spanish, tonno in Italian, ...) find their origin in the latin name *thunnus* itself issued from the Greek name  $\theta \upsilon \nu \nu \omicron \varsigma$ , (*thýnnos*) derived from the verb "*thynno*" which means "to rush".

## Principal market tuna species

Among the 14 species of tuna, skipjack, yellowfin, bigeye and albacore constitute more than 80% of the world catch since 1950. Three other species, Atlantic bluefin, Pacific bluefin and southern bluefin tuna, are also commercially important due to the high prices paid for them. These above-mentioned species are often referred as "principal market tuna species".

## Billfishes

The billfishes include 2 families: the Xiphiidae with one species, the swordfish (*Xiphias gladius*) and the Istiophoridae with 12 species within 4 genera.

The systematics of billfishes was recently revised by Orell et al. (2006) and Collette et al. (2006) using molecular analyses of nuclear DNA and mitochondrial DNA cytochrome b, leading to the distinction of billfishes from scombroids in a separate suborder, Xiphioidei and to the revision of billfish genera. Moreover, the lack of justification for separating the Atlantic sailfish (*Istiophorus albicans*) from the Indo-Pacific sailfish (*I. platypterus*) leads the author to gather them both under the latter genus. Similarly, Atlantic blue marlin (*Makaira nigricans*) and Indo-Pacific blue marlin (*M. mazara*) are regrouped in the single genus *Makaira nigricans*.

## Scientific and common names in English, French and Spanish of tunas and tuna-like species

### Tunas and bonitos

Scientific name	Common names, acknowledged by FAO, in: English French Spanish; 3-alpha code			
<b><i>Thunnini</i></b>	<b>Tunas</b>	<b>Thons</b>	<b>Atún</b>	
<i>Thunnus alalunga</i>	Albacore	Germon	Atún blanco	ALB
<i>Thunnus albacares</i>	Yellowfin tuna	Albacore	Rabil	YFT
<i>Thunnus atlanticus</i>	Blackfin tuna	Thon à nageoires noires	Atún aleta negra	BLF
<i>Thunnus maccoyii</i>	Southern bluefin tuna	Thon rouge du Sud	Atún del Sur	SBF
<i>Thunnus obesus</i>	Bigeye tuna	Thon obèse	Patudo	BET
<i>Thunnus thynnus</i>	Atlantic bluefin tuna	Thon rouge de l'Atlantique	Atún rojo del Atlantico	BFT
<i>Thunnus orientalis</i>	Pacific bluefin tuna	Thon rouge du Pacifique	Atún aleta azul del Pacifico	PBF
<i>Thunnus tonggol</i>	Longtail tuna	Thon rouge du Pacifique	Atún tongol	LOT
<i>Katsuwonus pelamis</i>	Skipjack	Thon mignon	Listado	SKJ
<i>Euthynnus affinis</i>	Kawakawa	Listao	Bacoreta oriental	KAW
<i>Euthynnus alleteratus</i>	Little tunny	Thonine orientale	Bacoreta	LTA
<i>Euthynnus lineatus</i>	Black skipjack	Thonine commune	Bacoreta	BKJ
<i>Auxis rochei</i>	Bullet tuna	Thonine noire	Barrilete negro	BLT
<i>Auxis thazard</i>	Frigate tuna	Bonitou	Melvera	FRI
		Auxide	Melva	
<b><i>Sardini</i></b>	<b>Bonitos</b>	<b>Bonites</b>	<b>Bonitos</b>	
<i>Allothunnus fallai</i>	Slender tuna	Thon élégant	Atún lanzón	SLT
<i>Cybiosarda elegans</i>	Leaping bonito	Bonite à dos tacheté	Bonito saltador	LEB
<i>Gymnosarda unicolor</i>	Dogtooth tuna	Bonite à gros yeux	Tasarte ojón	DOT
<i>Orcynopsis unicolor</i>	Plain bonito	Palomette	Tasarte	BOP
<i>Sarda australis</i>	Australian bonito	Bonite bagnard	Bonito austral	BAU
<i>Sarda chiliensis</i>	Eastern Pacific bonito	Bonite du Pac.	Bonito del Pac.	BEP

<i>Sarda orientalis</i>	DOMMO Indo-Pacific bonito	oriental Bonite orientale	oriental Bonito mono	BIP
<i>Sarda sarda</i>	Atlantic bonito	Bonite à dos rayé	Bonito atlántico	BON
<b>Seerfishes and mackerels</b>				
<b>Scientific name</b>	<b>Common names, acknowledged by FAO, in: English French Spanish; 3-alpha code</b>			
<i>Grammatorcynus bicarinatus</i>	Shark mackerel	Thazard requin	Carite cazón	SHM
<i>Grammatorcynus bilineatus</i>	Double-lined mackerel	Thazard-kusara	Carite cazón pintado	DBM
<b><i>Scomberomorini</i></b>	<b>Seerfishes</b>			
<i>Acanthocybium solandri</i>	Wahoo			
<i>Scomberomorus brasiliensis</i>	Serra Spanish mackerel			
<i>Scomberomorus cavalla</i>	King mackerel		<b>Carites</b>	
<i>Scomberomorus commerson</i>	Narrow-barred king mack.	<b>Thazards</b>	Peto	
<i>Scomberomorus concolor</i>	Monterey Spanish mackerel	Thazard-bâtard	Serra	WAH
<i>Scomberomorus guttatus</i>	Indo-Pacific king mackerel	Thazard serra	Carite lucio	BRS
<i>Scomberomorus koreanus</i>	Korean seerfish	Thazard barre	Carite estriado	KGM
<i>Scomberomorus lineolatus</i>	Streaked seerfish	Thazard rayé	Carite de	COM
<i>Scomberomorus maculatus</i>	Atlantic Spanish mackerel	Thazard Monterey	Monterey	MOS
<i>Scomberomorus papuanus</i>	Papuan seerfish	Thazard ponctué	Carite del Indo-Pacífico	GUT
<i>Scomberomorus multiradius</i>	Australian spotted mackerel	Thazard coréen	Carite coreano	KOS
<i>Scomberomorus munroi</i>	Japanese Spanish mackerel	Thazard cirrus	Carite coreano	STS
<i>Scomberomorus nipponius</i>	Kanadi kingfish	Thazard atlantique	Carite rayado	SSM
<i>Scomberomorus plurilineatus</i>	Queensland school mackerel	Thazard papou	Carite atlántico	PAP
<i>Scomberomorus queenslandicus</i>	Cero	Thazard australien	Carite papuense	ASM
<i>Scomberomorus regalis</i>	Broad-barred king mackerel	Thazard oriental	Carite australiano	NPH
<i>Scomberomorus semifasciatus</i>	Pacific sierra	Thazard Kanadi	Carite oriental	KAK
<i>Scomberomorus sierra</i>	Chinese seerfish	Thazard de	Carite canadí	QUM
<i>Scomberomorus sinensis</i>	West African	Queensland	Carite de	CER
<i>Scomberomorus tritor</i>	Spanish mackerel	Thazard franc	Queensland	BBM
		Thazard tigre	Carite chinigua	SIE
		Thazard sierra	Carite tigre	CHY
		Thazard nébuleux	Carite sierra	MAW
		Thazard blanc	Carite indochino	
			Carite lusitánico	
<b><i>Scombrini</i></b>	<b>Mackerel</b>	<b>Maquereaux</b>	<b>Caballa</b>	
<i>Rastrelliger brachysoma</i>	Short mackerel	Maquereau trapu	Caballa rechoncha	
<i>Rastrelliger faughni</i>	Island mackerel	Maquereau des îles	Caballa isleña	RAB
<i>Rastrelliger kanagurta</i>	Indian mackerel	Maquereau des Indes	Caballa de la India	RAF
<i>Scomber australasicus</i>	Spotted chub mackerel	Maquereau tacheté	Caballa pintoja	RAG
	Atlantic chub	Maquereau espagnol de	Estorino del Atlántico	MAA

<i>Scomber colias</i>	Atlantic chub mackerel	l'Atlantico	Atlantico	MAS
<i>Scomber japonicus</i>	Indo-Pacific Chub mackerel	Maquereau espagnol indo-Pacífico	Estorino del Indo-Pacífico	MAS
<i>Scomber scombrus</i>	Atlantic mackerel	Maquereau commun	Caballa del Atlántico	MAC
<b><i>Gasterochismatinae</i></b>				
<i>Gasterochisma melampus</i>	Butterfly kingfish	Thon papillon	Atún chauchera	BUK
<b>Billfishes</b>				
<b>Scientific name</b>	<b>Common names, acknowledged by FAO, in: English French Spanish; 3-alpha code</b>			
<i>Xiphias gladius</i>	Swordfish	Espadon	Pez espada	
<i>Istiophorus albicans</i>	Sailfish	Voilier de l'Atlantique	Pez vela del Atlántico	
<i>Istiophorus platypterus</i>	Indo-Pacific sailfish	Voilier de l'Indo-Pacifique	Pez vela del Indo-Pacífico	SWO
<i>Istiompax indica</i>	Black marlin	Makaire noir	Aguja negra	SAI
<i>Makaira mazara</i>	Indo-Pacific blue marlin	Makaire bleu de l'Indo-Pacifique	Aguja azul del Indo-Pacífico	SFA
<i>Makaira nigricans</i>	Blue marlin	Makaire bleu de l'Atlantique	Aguja azul del Atlántico	BLM
<i>Kajikia albigata</i>	Atlantic white marlin	Makaire blanc de l'Atlantique	Aguja blanca del Atlántico	BLZ
<i>Kajikia audax</i>	Striped marlin	Makaire à rostre court	Marlín rayado	BUM
<i>Tetrapterus angustirostris</i>	Shortbill spearfish	Marlin rayé	Marlín trompa corta	WHM
<i>Tetrapterus audax</i>	Striped marlin	Marlin de la Méditerranée	Marlín rayado	SSP
<i>Tetrapterus belone</i>	Mediterranean spearfish	Makaire épée	Marlín rayado	MLS
<i>Tetrapterus georgei</i>	Roundscale spearfish	Makaire bécune	Marlín del Mediterráneo	MSP
<i>Tetrapterus pfluegeri</i>	Longbill spearfish		Marlín peto	RSP
			Aguja picuda	SPF

### Fossil records

The first fossil records of *Scombridae* are dated from the beginning of the lower Eocene epoch (60–40 million years ago) during the lower Tertiary period. As for the *Istiophoridae*, the oldest fossils are dated from the upper Cretaceous epoch (70–90 million years ago) during the Secondary period. For the *Xiphiidae*, the oldest fossil records are dated from the Paleocene epoch of the lower Tertiary period, i.e. 57–65 million years ago (Berg, 1958).

## Morphological characteristics

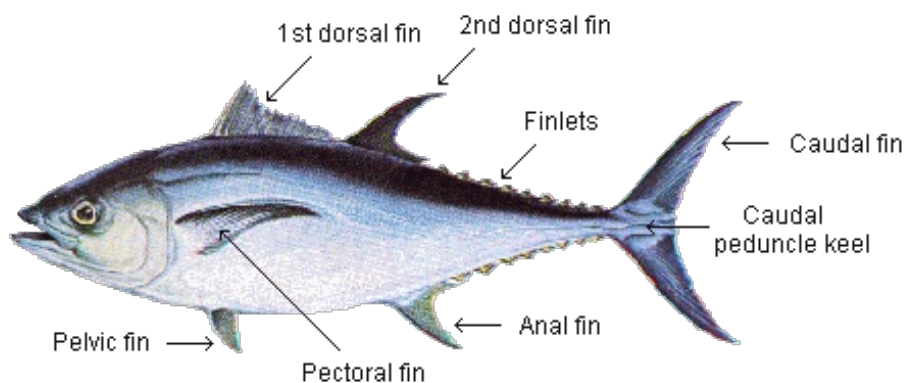
### Morphology of larvae

It is often difficult or impossible to identify larvae and, in some cases, early juveniles by anatomical characteristics or colour patterns. Biochemical or genetic methods can be used to distinguish the larvae of the various species (Elliott and Ward, 1995, Chow *et al.*, 2003). Diagnostic keys are available for larvae between 3 and 12 mm standard length. Larvae smaller than 3 mm are virtually indistinguishable (Nishikawa and Rimmer, 1987).

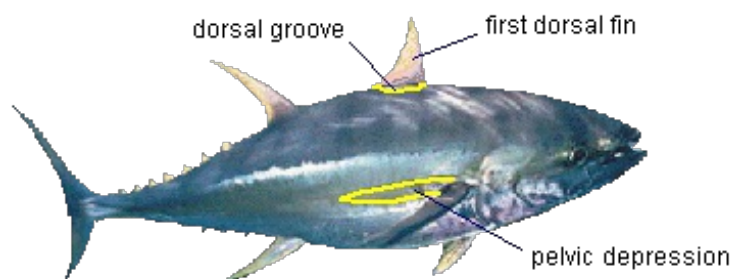
## Morphology of juveniles and adults

### Characteristics common to both scombrids and billfishes

Both scombrids and billfishes have two distinct dorsal fins, generally separated, the first one supported by spines and the second only by soft rays. The pelvic fins are inserted below the base of the pectoral fins. The caudal fin is deeply notched.



All scombrids and billfishes except swordfish have a pair of caudal keels on the middle of the caudal peduncle at the base of the caudal fin. The swordfish has only a large median caudal keel. The more advanced members of the Scombridae family also have a large median keel anterior to the pair of caudal keels. The bodies of all the Scombroidei are robust, elongate and streamlined. The first dorsal and first anal fins of all scombrids and billfishes, except swordfish, can fold down into grooves and the pectoral and pelvic fins into depressions when the fish is swimming rapidly.



The scombrids and billfishes, all have four gill arches on each side. The gill filaments are ossified as "Gill rays".

### Scombrids

The scombrids are characterised by the presence of at least four finlets behind the dorsal and the anal fins. The pelvic fins are smaller than the pectoral fins or of equal size. Except for the primitive butterfly kingfish, which body is covered by large cycloid scales, the body of all the

scombrids is naked or covered with small to moderate-sized scales. The bonitos (Sardini) are intermediate between the seerfishes and the tunas. As is the case for the tunas, they have a well-developed corselet of scales, but they lack the two longitudinal ridges on the upper surface of the tongue. The most primitive Scombrinae are the mackerels (Scombrini), the seerfishes (Scomberomorini) and the two-line mackerels (*Grammatorcynus* spp.). The mackerels have only two caudal keels, whereas the seerfishes and the two-line mackerels have a larger median keel in front of the pair of keels.

The tunas are the most highly evolved of the scombrids. They are unique among bony fish in having heat exchanger systems that allow them to regulate their body temperature, as can birds and mammals (see the thermoregulation).

## Billfishes

The billfishes are characterised by their rostrum, an extension of the upper jaw, which extends much beyond the lower jaw.



*Billfishes rostrum*

The rostrum has a flat, sword-like cross section for the swordfish and a rounded, spear-like cross section for the Istiophoridae.

The swordfish is also characterised by the absence of pelvic fins and scales. Its dorsal fins are well separated, and has only one large median caudal keel. The Istiophoridae have long, rigid, tapering pelvic fins. Their bodies are covered with small, elongate, bony scales. Their first dorsal fin has a long base and terminates close to the origin of the second dorsal fin.

## Geographical distribution

### Tropical and temperate tunas

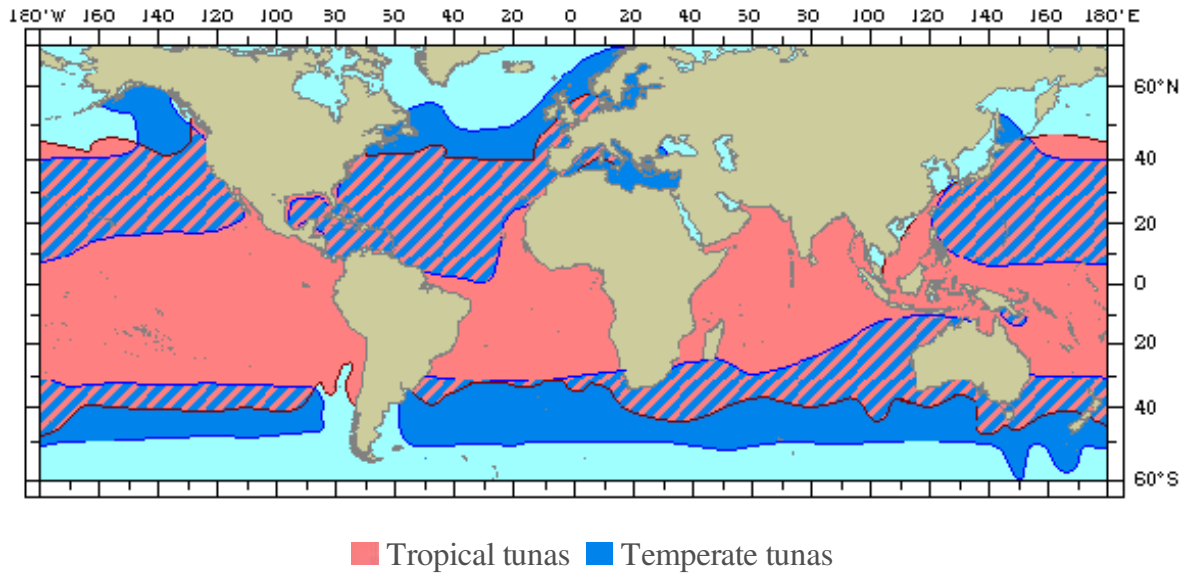
Because of different distributions due to their specific thermal tolerances and because of exploitation by different fisheries, a distinction is made between tropical and temperate tunas. Tropical tunas are found in waters with temperatures greater than 18° C (although they can dive in colder waters) whereas temperate tuna are found in waters as cold as 10°C, but can also be found in tropical waters (Brill, 1994).

Tropical tunas: skipjack and yellowfin

Intermediate tunas: bigeye

Temperate tunas: albacore, Pacific bluefin, Atlantic bluefin and southern bluefin

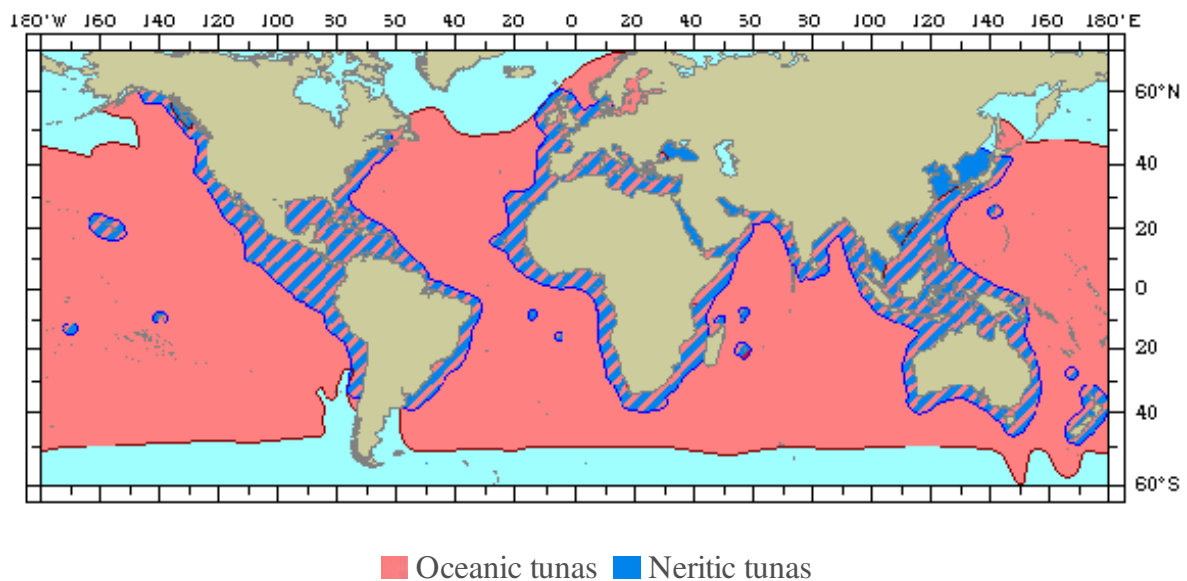
### *Tropical and temperate tunas*



## **Scomberids**

Tunas prefer oceanic waters, and 3 of the 8 species of *Thunnus* are found worldwide except in the Arctic Ocean. Most bonitos and little tunas (*Euthynnus* spp.) are primarily neritic, *ie* coastal fishes, but the distribution of individual species is often widespread. The frigate and bullet tunas (*Auxis* spp.) are probably both oceanic and coastal (Olson and Boggs, 1986). The slender tuna and the butterfly kingfish have circum-global distributions in the Southern Ocean. Most mackerels and seerfishes have restricted ranges of distribution. Exceptions are the Spanish mackerel and the wahoo which are found worldwide.

### *Oceanic and neritic tunas*





## Billfishes

Billfishes are widely distributed, at least, throughout the oceans in which they occur. The exception are the Mediterranean spearfish, which occurs only in the Mediterranean Sea, and perhaps the roundscale spearfish, which occur in the northeastern Atlantic Ocean around the Canary and Madeira Islands and in the western Mediterranean Sea. However, only the swordfish is cosmopolitan. All other *Istiophoridae* are being confined to the Atlantic Ocean or to the Indian and Pacific Oceans.

### Occurrence of the tuna, bonito and billfish species in the different oceans

Common names	Scientific name	Areas of occurrence
<b>Tunas and bonitos</b>		
Skipjack	<i>Katsuwonus pelamis</i>	worldwide
Yellowfin tuna	<i>Thunnus albacares</i>	worldwide
Bigeye tuna	<i>Thunnus obesus</i>	worldwide
Albacore tuna	<i>Thunnus alalunga</i>	worldwide
Atlantic bluefin tuna	<i>Thunnus thynnus</i>	Atlantic Ocean
Pacific bluefin tuna	<i>Thunnus orientalis</i>	Pacific Ocean
Southern bluefin tuna	<i>Thunnus maccoyii</i>	southern parts of Atlantic, Indian and Pacific Ocean
Longtail tuna	<i>Thunnus tonggol</i>	Indian Ocean, western Pacific Ocean
Blackfin tuna	<i>Thunnus atlanticus</i>	western Atlantic Ocean
Kawakawa	<i>Euthynnus affinis</i>	Indian, western and central Pacific Oceans
Black skipjack	<i>Euthynnus lineatus</i>	eastern Pacific Ocean
Little tunny	<i>Euthynnus alleteratus</i>	Atlantic Ocean
Bullet tuna	<i>Auxis rochei</i>	worldwide
Frigate tuna	<i>Auxis thazard</i>	Indian and Pacific Oceans
Slender tuna	<i>Allothunnus fallai</i>	Southern Ocean
<b>Billfishes</b>		
Swordfish	<i>Xiphias gladius</i>	worldwide
Atlantic sailfish	<i>Istiophorus albicans</i>	Atlantic Ocean
Indo-Pacific sailfish	<i>Istiophorus platypterus</i>	Indian and Pacific Oceans
Black marlin	<i>Makaira indica</i>	Indian and Pacific Oceans
Indo-Pacific blue marlin	<i>Makaira mazara</i>	Indian and Pacific Oceans
Atlantic blue marlin	<i>Makaira nigricans</i>	Atlantic Ocean
Atlantic white marlin	<i>Tetrapterus albidus</i>	Indian and Pacific Oceans
Striped marlin	<i>Tetrapterus audax</i>	Indian and Pacific Oceans

## Habitat and biology

## Ecological niche

Tunas are pelagic marine fish, spending their entire lives relatively near the surface of tropical, subtropical and temperate oceans and seas. Scombrids and billfishes live primarily in the water layers above the thermocline, but are able to dive to depth of several hundred meters (see the Vertical distribution section). Tuna species attaining only small sizes and juveniles of those attaining large sizes are encountered in epipelagic waters (from the surface to the thermocline) whereas large tunas tend to be mesopelagic and are found also in deeper and cooler waters.

Epipelagic tunas: skipjack and bonitos, juveniles of large tunas, billfishes

Mesopelagic tunas (adults): albacore, bigeye and bluefin

Tunas that are found at both depth ranges: yellowfin, swordfish

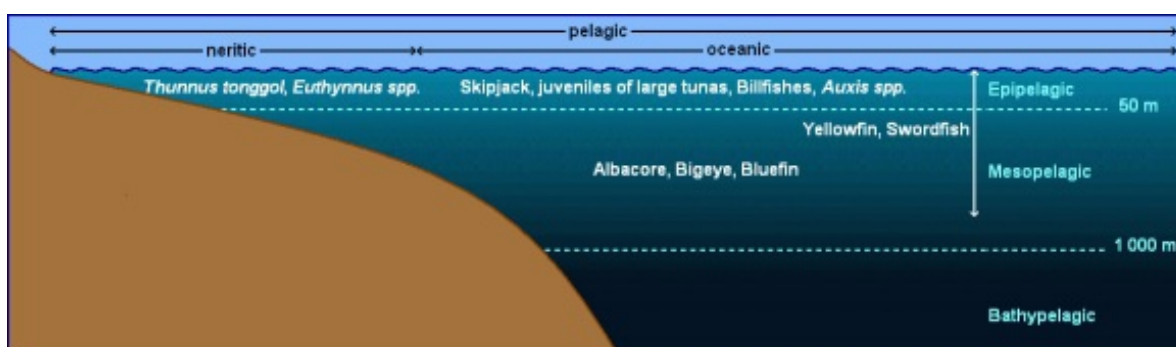
Some tunas are found in both offshore and coastal waters and others entirely, or almost entirely, in coastal waters.

Mid-ocean species: yellowfin and bigeye, swordfish

Coastal species: *Thunnus tonggol*, other bonitos than *Auxis spp.*, little tunas (*Euthynnus spp.*)

Species found in both waters: skipjack, albacore, Pacific bluefin, Atlantic bluefin and southern bluefin, *Auxis spp.*

Seerfishes are generally restricted to coastal waters and enter estuaries to feed. One species, the Chinese seerfish moves long distances in freshwater up the Mekong River in China.



*Distribution of*

*tunas and tuna-like in the water*

## Tuna and their environment

Important environmental parameters for tuna are the sea surface temperature, the quantity of dissolved oxygen in the water and the salinity. Lower thermal boundaries vary between 10°C for temperate tunas and 18°C for tropical tunas (see above; Brill, 1994). The minimum oxygen requirement is estimated between 2 to 2.7 ml/l for principal market tuna species except for bigeye tuna which can tolerate oxygen concentrations as low as 0.6 ml/l (Sharp, 1978 ; Lowe, 2000). Most tunas tend to concentrate along thermal discontinuities such as oceanic fronts (Sund, 1981).

## Vertical distribution constraints and diving behavior

The vertical distribution of most species of tunas is influenced by the thermal and oxygen structure of the water column. Tuna species attaining only small sizes and juveniles of those attaining large sizes tend to live near the surface, whereas adults of large species are found in deeper waters. The use of deep longlines showed that bigeye can be found at depths as great as 300 m (Suzuki *et al.*, 1977). Albacore are also caught under FADs at depths to about 200 m (Bard *et al.*, 1998). Acoustic telemetry has shown that billfishes are found near to the

surface during the day, descending more frequently to greater depths at night (Block *et al.*, 1992a).

Yellowfin, Bigeye, Bluefin and Swordfish show deeper dives than other species of tuna and billfish (Fromentin & Fonteneau, 2001; Sund, Blackburn & Williams, 1981):

- Yellowfin tuna has been observed to dive at more than 1100 m (record of 1200 m, L. Dagorn *et al.*, 2006).
  - Bigeye tuna are capable of diving to depths of more than 1200 m (record of 1800 m, Schaefer, *Comm. pers.*).
  - Atlantic bluefin tuna are able to dive to depths in excess of 1000 m, encountering an exceptionally wide range of temperatures (Block *et al.*, 2005).
  - Swordfish make large vertical excursions, coming close to the surface at night and diving as deep as 600 m during the day ((Carey and Robison, 1981); Sedberry and Loefer, 2001), even 900 m (Takahashi *et al.*, 2003).
- Other tunas show lesser diving capabilities, as, for example, *Euthynnus affinis* which have been observed at 400 m depth (Lee, 1982).

## Schooling behavior

Tunas use schooling to their advantage when they forage. Some tunas form parabolic-shaped schools to encircle their prey. Most tunas school according to size. Juveniles of tunas attaining large sizes are, therefore, often associated with tunas attaining only small sizes, such as skipjack or bonito. Schools of large adults consist of a few scattered individuals. Schooling offers protection for juvenile tunas by confusing predators and reducing the likelihood that any single fish will become a victim to a predator. Atlantic bluefin tuna can form giant schools spread over several nautical miles when migrating into the Mediterranean Sea to spawn during the summer. As is the case with the other fishes, the structure of tuna schools is maintained by the lateral line. Schools can gathered over 5000 individuals.

## Migration and other movements

All tunas and tuna-like fishes move constantly to search for food and to keep water passing over their gills. Migrations are seasonal movements, often over long distances, for the purpose of feeding or reproduction. Temperate tunas, i.e. albacore, Atlantic bluefin and Pacific bluefin, migrate long distances between temperate waters, where they feed, and tropical waters, where they spawn without moving among different oceans. Southern bluefin tuna migrates among the southern parts of Atlantic, Indian and Pacific Oceans. Although, the distribution of the three species of bluefin is quite extended, their spawning is restricted to relative small areas of tropical waters. Tropical tunas, i.e. skipjack and yellowfin, are less migratory in terms of long-distance directional movements, although several tagged yellowfin released in the western Atlantic have been recaptured in the eastern Atlantic. Bigeye have some of the characteristics of both temperate and tropical tunas. They apparently do not make trans-oceanic migrations, but like the temperate tunas, they migrate back and forth between feeding grounds in temperate waters and their spawning grounds in tropical waters. When they are not making directional migration, tunas move nearly all the time in search of areas where the food is most abundant. Fishermen are sometimes able to predict on the basis of oceanic conditions where the fish are likely to appear and then, they can transfer their operations to those areas. Less is known of the movements of billfishes, but apparently, they make seasonal migrations between temperate waters, where they feed, and tropical waters, where they spawn. For instance, blue marlin display extensive trans-equatorial and inter-oceanic movements from the Atlantic into the Indian Ocean (Ortiz *et al.*, 2003).

## Swimming

Tunas are excellent swimmers, and their bodies are designed for high performance at both sustainable and burst swimming speeds (Dickson, 1995). Tunas must swim constantly to satisfy their oxygen requirements and consequently stay alive. The direction of movements of some species, such as skipjack, seem to be dictated solely by the availability of food. The movement of other species, such as the three species of bluefin, seem to be influenced by both the distribution of food and the need to return to their ancestral spawning grounds at the

proper time. Tunas can move up to 15 km per night in order to forage on organisms that swim upward from deeper waters at that time.

Tunas have higher cruising speeds than do other active fishes, including other scombroids (Beamish, 1978; Block *et al.*, 1992b). The morphology of the body and caudal fin of tunas is optimal for prolonged, high-speed swimming. Similar body designs are found also in cetaceans, carangids, certain sharks and even the extinct reptilian ichthyosaurs. Webb (1984) lists the following morphological adaptations:

- a lunate tail of large span, but relatively small chord to maximize the thrust
- a narrow caudal peduncle to provide for locally-large amplitude displacements and to control the angle of attack
- a large anterior body depth and mass to reduce recoil energy losses
- a relatively-rigid streamlined body to both minimize the drag and maximize the thrust

## Long-range swimming

The net distances travelled by tunas and billfishes (shortest distances between the locations of release and recapture) exceed those of any other fish, as shown by the following records obtained from tagging studies (from Joseph *et al.*, 1988 for tunas and Orbesen *et al.*, 2008 for billfishes):

- 10,790 km for a Pacific bluefin tuna (from southeast of Japan to off Baja California)
- 10,680 km for a black marlin (from off Baja California to Norfolk Island in the South Pacific Ocean)
- 9,500 km for a skipjack tuna (from off Baja California to the Marshall Islands)
- 8,500 km for an albacore tuna (from off California to off Japan)
- 7,700 km for an Atlantic bluefin tuna (across the Atlantic Ocean)
- 15,744 km for a blue marlin (across the Atlantic Ocean)
- 14,556 km for a black marlin (across the Pacific Ocean, comm.. pers.)
- 6,523 km for a white marlin (across the Atlantic Ocean)
- 3,845 km for a sailfish (across the Atlantic Ocean)

In addition, net movements of more than 5,000 km have been recorded for yellowfin tuna, bigeye tuna, blue marlin, striped marlin and sailfish.

## Short-range, fast swimming

Scombrids and billfishes are adapted to fast swimming. The champions are, of course, the most highly evolved scombrids, the bonitos (Sardini) and the tuna (Thunnini) and the billfishes. They are able to exhibit startling bursts of speed, often exceeding one body length per second. The record (for all bony fishes) belongs to the black marlin (*Makaira indica*), which has been clocked at over 130 km/h.

### Estimated maximum swimming speed of some tunas and billfishes

Species	Sustained in m/s	Burst in m/s
<b>Scombrids</b>		
<i>Thunnus albacares</i> (1)	0.64	20.46
<i>Thunnus obesus</i> (1)	0.6	-

<i>Thunnus thynnus</i> (1)	3.49	-
<i>Katsuwonus pelamis</i> (1)	0.84	9.41
<i>Euthynnus affinis</i> (1)	0.76	5
<i>Auxis rochei</i> (1)	0.68	-
<i>Sarda chiliensis</i> (1)	0.88	3.70
<i>Sarda sarda</i> (1)	0.35	1.2
<i>Acanthocybium solandri</i> (1)	0.4	21.23
<i>Scomber japonicus</i> (1)	0.92	2.25
<i>Scomber scombrus</i> (2)	0.98	5.4
<b>Billfishes</b>		
<i>Tetrapterus audax</i> (3)	1.8	-
<i>Makaira indica</i> (3)	-	36.1
<i>Makaira nigricans</i> (3)	1	2.25
<i>Xiphias gladius</i> (2)	-	24.86

## References

- (1): Magnuson (1978)  
(2): Wardle and He (1988)  
(3): Block *et al.* (1992b)

## Physiological aspects of swimming

In order to swim at high speeds for long periods, tunas are capable of taking in and utilizing large amounts of oxygen.

In contrast to other fishes that contract their jaws and opercular muscles to pump water over their gills, tunas and billfishes (and some species of sharks) ram ventilate, that is they swim through the water with their mouths open, which forces water over their gills. This is an efficient way to get a large amount of water flowing through the gills at a low energetic cost, but it has an important drawback: tunas cannot stop swimming, or they will suffocate! They must swim at a speed of at least 0.65 m/s to provide sufficient flow of water over their gills.

The amount of gill surface of tunas is up to 30 times those of other fish, and for some tunas the absorptive surface approaches those of the lungs of mammals of comparable weight (Joseph *et al.*, 1988). This large surface enables the tunas to extract about half of the oxygen present in the water flowing over their gills.

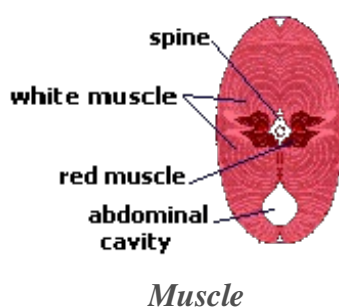
To transfer oxygen from the gills to the other tissues, tunas have hearts that are about 10 times the size, relative to the weight of the entire body, of those of other fish. The blood pressure of tunas is about three times those of other fishes, and their hearts pump blood at a rate about three times those of other fish. The blood of tuna has a hematocrit of 40%, a value usually associated with diving mammals.

Scombrids and billfishes, like most fish, have two types of muscle, white and red. The white muscles function during short bursts of activity, while the red muscles, which have a relatively large mass, allow the fish to swim at high speeds (up to 45 km/h) for long periods without fatigue, as demonstrated by tagging studies with conventional and sonic tags (Joseph *et al.*, 1988 ; Bushnell and Holland, 1997).

The proportion of red muscle is much greater for tunas than for other fish (Dickson, 1995) and their white muscles are capable of working in both aerobic and anaerobic conditions. Therefore, the increase in swimming speed can be portrayed as follows :

	sustained speed	high speed	burst speed
<b>red muscle in action</b>	yes		
<b>white muscle in action</b>			
aerobic condition	yes	yes	
anaerobic condition	yes	yes	yes

The red muscles are located deep within the body, and appear to be more important at the anterior part of the fish. They extend from the vertebral column to a lateral subcutaneous position. In contrast to other fishes, the proportion of red muscle does not seem to increase with the size of the tuna, probably because of greater muscle efficiency and labor sharing between red and white muscles, to which both endothermy and thermoregulation could contribute (Graham *et al.*, 1983).



Heart and white muscle aerobic capacities are significantly greater in tunas than in billfishes and other scombrids.

## Recovery from intense activities

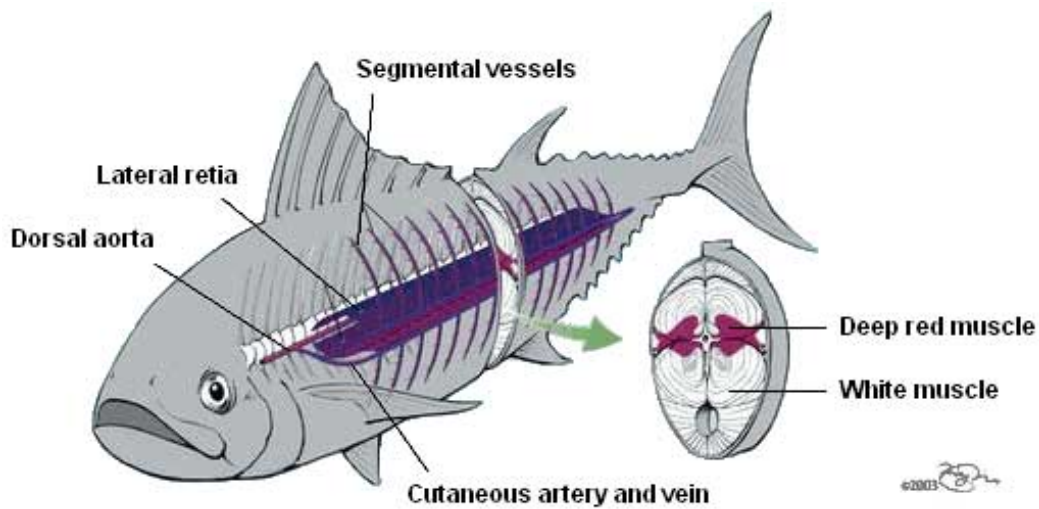
Furthermore, tunas and billfishes are capable of recovering more quickly than other fish after intense activities, such as that involved in capture of prey. For some tunas, the rates of removal of lactate from the blood and white muscle tissue approximate the rates measured in mammals, which allows the tuna to recover within two hours (Dickson, 1995).

## Thermoregulation in tuna

As a consequence of swimming constantly to maintain hydrostatic equilibrium (Magnuson, 1973) and oxygenate the blood (Roberts, 1978), muscular metabolism continuously generates heat as a byproduct. Tunas get rid of this excess, but, on the other hand, the heat can be used by the tuna to enable them to forage in cold waters.

## Metabolic mechanism for thermoregulation

Among all bony fish, the Thunnini are unique in their ability to regulate their body temperatures, due to a complex counter-current heat exchanger system, also called the *rete mirabile* (miraculous network) (Stevens and Neil, 1978). The only other fishes with this system are some sharks of the family *Lamnidae* (Collette, 1978).



*Rete mirabile* (from Weinheimer, 2003)

The tuna maintain their body temperatures above that of the ambient water by passing arterial blood through vascular countercurrent heat exchangers. All species of tuna have a lateral rete, consisting of small arteries branching from the lateral subcutaneous arteries and small veins emptying into the lateral veins (Graham *et al.*, 1983). In addition, many species of tuna also have a central rete within the vertebral haemal canal, consisting of arteries from the dorsal aorta and veins to the posterior cardinal veins (Stevens and Neil, 1978). The arterial blood is, then, warmed by the venous blood that flows through the red swimming muscles (Holland *et al.*, 1992).

The *rete mirabile* retains between 70 and 99 % of the heat produced by the red muscle fibers, and provide a barrier between the red muscle and the environment (Graham *et al.*, 1983). However, when excessive temperatures have been generated by heavy exercise, tunas appear to be able to control the efficiency of the heat exchangers by closing down some blood vessels of the *rete mirabile*, allowing heat to dissipate into the colder ambient water (Bushnell and Holland, 1997).

Measurements of body temperatures and ambient temperatures with histological analyses of the *rete mirabile* show that tunas as small as 207 mm in length can maintain their body temperatures more than 3°C above the ambient temperature, and thus can be considered to be endotherms (Graham *et al.*, 1983). Tuna body temperatures are often 10°C greater than those of ambient water. The maximum temperature difference was recorded for an Atlantic bluefin tuna, for which the body temperature was 21.5°C greater than the surrounding water (Graham *et al.*, 1983).

The thermoregulatory system cannot conserve heat indefinitely, and when a fish has been foraging in deep, cold water for an extended period, its body temperature decreases. When this happens, it can ascend to warmer water and disengage its thermoregulatory system to allow rapid warming of the tissues (Holland *et al.*, 1992).

## Behavioural mechanisms for thermoregulation

Combined with the physiological mechanisms, movements into cooler water will facilitate heat dissipation (Bushnell and Holland, 1997).

## Advantages of thermoregulation

Thermoregulation allows the tunas to sustain high swimming speeds for long periods and to recover quickly after prolonged exertion (Carey *et al.*, 1971), because most biochemical reactions proceed more rapidly at higher temperatures. Therefore, according to Bushnell and Holland, 1997, elevated body temperatures allow:

red muscle to contract more quickly, approaching the contraction rate of white muscle and consequently, contributing to high-speed swimming resulting from white muscle contractions  
more rapid transfer of oxygen from blood to muscle cells  
more rapid recovery, by enhancing the breakdown of lactic acid

In addition, being "warm bodied" allows the tunas to have a good vision at significant depths by maintaining their brains and eyes at greater than ambient temperatures (Bushnell and Holland, 1997). It also allows the tuna to be more sensitive to thermal gradients (Sharp and Dizon, 1978).

Also, because of that, the tuna can forage beneath the thermocline, in deep water, without suffering radical decreases in their core temperature. For example, a bigeye tuna was observed to dive 250 meters in one minute, going from 24°C to 9°C water (Holland *et al.*, 1992).

## Trophic relations and growth

### Growth stages

The following three stages can be distinguished:

larvae (recently hatched individuals which are considerably different in appearance from juveniles or adults)

juveniles (similar in appearance to adults, but sexually immature)

adults (sexually mature fish)

### Trophic position of larvae

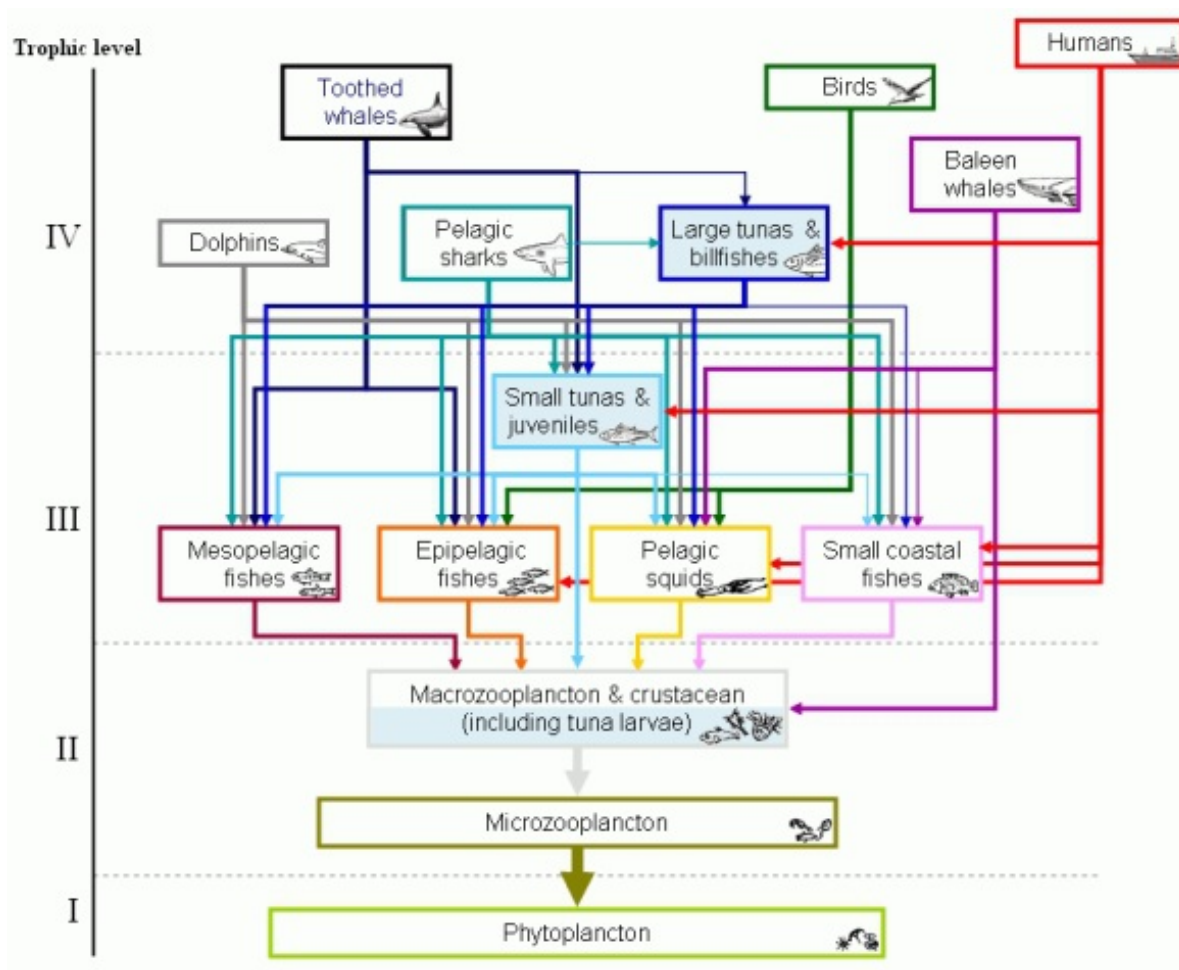
Larvae of tunas and tuna-like fishes live in warm surface waters and feed primarily on the zooplankton including small crustaceans and larvae of crustaceans, fishes, molluscs and jelly-fish. Larvae of tunas and tuna-like species are preyed upon by zooplankton foragers, such as larger larvae and early juveniles of pelagic fishes. Cannibalism is, therefore, an important cause of mortality for tuna larvae.

### Trophic position of juveniles and adults

#### Tunas and tuna-like fishes in the oceanic food web

Tunas and billfishes prey on fish, squid and crustaceans. The larger individuals (wahoo, bonitos, tunas and billfishes), which feed on pelagic fishes, are positioned at the top of the trophic web. The smaller individuals (juvenile tunas and billfishes, mackerels and seerfishes) prey on zooplankton (mainly crustaceans) and constitute part of the ration of large scombroids, sharks, cetaceans and seabirds. Analyses of stomach contents of yellowfin and skipjack tuna indicate that they feed on small epipelagic fishes between 1 and 10 cm in length (Roger, 1994). Since these preys of yellowfin and skipjack feed directly on zooplankton (mainly copepods), it seems that the tunas are at the top of a short food web, which is probably very efficient from the point of view of energetics.

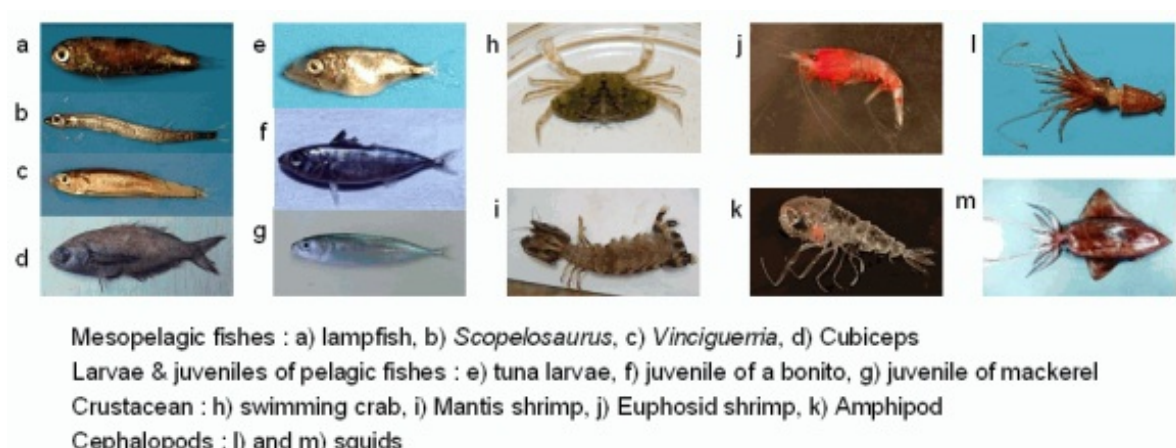




*Position of tunas and tuna-like fishes in the oceanic food web*

## Food items

Tunas and billfishes are opportunistic feeders. At the species level, they do not have strong preferences for certain types of prey. However, on a regional scale and at a given time, a few species may represent almost all of the food of fish of a specific age group (Cayré *et al.*, 1988). Tunas and billfishes prey on pelagic or epipelagic fishes (including juveniles of tunas), crustaceans and cephalopods (squids). Yellowfin and bigeye tunas as well as swordfish feed on mesopelagic fishes (Ménard *et al.*, 2000, Allain, 2005). Coastal tunas feed on neritic and epipelagic prey (Olson and Boggs, 1986). Larger tunas feed on small pelagic fishes such as mackerels, small tunas, carangids or flying fishes.



*Example of preys that are often found in the stomach of tunas*

## Foraging behavior of juveniles and adults

Tunas and billfishes are predators that locate their prey visually. To satisfy their food requirements tunas and billfishes have to swim long distances. Their type of locomotion is, therefore, particularly adapted to the search for prey in a large volume of water with the least expenditure of energy. However, they appear less effective than transient predators, such as esocids, in actually capturing the prey (Webb, 1984). To compensate for this, tunas tend to break up schools of prey, producing disorientation and straggling, and/or search for prey in schools (Webb, 1984; Partridge, 1982). Tunas can detect minute traces of scents of oils, proteins and amino-acids of the mucus layer produced by their prey. When prey is detected, some tunas show changes in their behavior consisting of a general increase of activity (also called frenzy): increase in swimming speed, change in swimming pattern, jaw snapping and display of dark stripes on the flanks. Tropical tunas and swordfish often dive down at significant depths below the thermocline (commonly at 500 m) to feed on mesopelagic fishes (Holland *et al.*, 1992). It is commonly believed that tunas feed during the day. However, sonic tracking experiments show that some tunas feed also at dusk, when mesopelagic micronecton migrate toward the surface (Bard *et al.*, 1998).

## Growth of juveniles and adults

### Growth rates

Most scombrids grow rapidly and reach their adult sizes in a few years. Average growth rates vary according to the species, the age and the location. In general, larger tunas grow to about 40 to 55 cm the first year, then the annual growth rate ranges between 20 to 30 cm per year decreasing with age. Tuna species attaining only small sizes grow to 20 to 35 cm in the first year and their annual length increments rapidly decrease to less than 10 cm. In the Atlantic and Indian Oceans, several studies have shown that yellowfin grow rapidly during the first year, slowing their growth during the next one or two years and then having again a fast growth before gradually slowing down as the maximum size is approached. Seerfishes and mackerels have also a fast growth during their first years of life. Sizes of 35 to 45 cm at age 1 year are common.

Billfishes can grow to more than 80 cm during their first year of life. After this very fast juvenile growth, adult growth rates are comparable to those of tunas.

### Weights and lengths ranges

The maximum weights attained by tunas range from about 1 to 2 kg for bullet and frigate tunas to more than 600 kg for Atlantic bluefin tuna. The maximum lengths attained by tunas range from about 50 cm for bullet and frigate tunas to more than 300 cm for Atlantic bluefin tuna.

Seerfishes, mackerels and bonitos are relatively small (less than 1 meter in length), except for some species of seerfishes such as the king mackerel or the narrow-barred king mackerel which grow to more than 240 cm, for 70 kg.

The smallest billfish is the Mediterranean spearfish, which reaches a maximum length of a little more than 180 cm. The largest billfishes are the black marlin and the Indo-Pacific blue marlin, which reach lengths of more than 4 m and weights of more than 600 kg.



Atlantic bluefin tuna



Bigeye tuna



Yellowfin tuna



Albacore tuna



Skipjack tuna

*Relative sizes of tuna main market species*

## Size

### Common and maximum sizes of tunas and billfishes

#### Tunas (1)

Scientific name	Common size (in cm)	Maximum size (in cm)	Maximum weight (in kg)
<i>Auxis rochei</i>	15-35	50	-
<i>Auxis thazard</i>	25-40	58	-
<i>Euthynnus lineatus</i>	30-65	70	9
<i>Euthynnus alleteratus</i>	30-80	100	12
<i>Euthynnus affinis</i>	30-60	100	13
<i>Katsuwonus pelamis</i>	40-80	108	33
<i>Thunnus atlanticus</i>	40-70	100	19
<i>Thunnus alalunga</i>	40-100	127	40
<i>Thunnus tonggol</i>	40-70	130	35
<i>Thunnus albacares</i>	60-150	200	175
<i>Thunnus maccoyii</i>	160-200	225	160
<i>Thunnus obesus</i>	70-180	230	200
<i>Thunnus thynnus</i>	80-200	300	650

### Bonitos (1)

Scientific name	Common size (in cm)	Maximum size (in cm)	Maximum weight (in kg)
<i>Cybiosarda elegans</i>	35-45	50	5
<i>Sarda sarda</i>	30-50	85	7.5
other <i>Sarda</i> *	30-50	100	-
<i>Allothunnus fallai</i>	65-95	96	10
<i>Orcynopsis unicolor</i>	40-90	130	13
<i>Gymnosarda unicolor</i>	65-150	200	131

\* *Sarda australis*, *S. chiliensis*, *S. orientalis*, *S. sarda*

### Seerfishes and mackerels (1)

Scientific name	Common size (in cm)	Maximum size (in cm)	Maximum weight (in kg)
<i>Rastrelliger faughni</i>	-	20	0.75
<i>Rastrelliger brachysoma</i>	15-25	35	-
<i>Rastrelliger kanagurta</i>	15-25	35	-
<i>Grammatorcynus</i> spp.	-	60	3.5
<i>Scomber</i> spp.	15-30	40-50	1
<i>Scomberomorus multiradius</i>	-	35	0.5
<i>Scomberomorus concolor</i>	-	75-80	-
<i>Scomberomorus guttatus</i>	-	75-80	-
<i>Scomberomorus niphonius</i>	-	100	4.5
<i>Scomberomorus cavalla</i>	20-70	170	-
<i>Scomberomorus commerson</i>	30-90	220	45
<i>Gasterochisma melampus</i>	74-164	164	-
<i>Acanthocybium solandri</i>	100-170	210	80

### Billfishes (2)

Scientific name	Common size (in cm)	Maximum size (in cm)	Maximum weight (in kg)
<i>Tetrapterus georgei</i>	-	160 BL	21
<i>Tetrapterus pfluegeri</i>	-	200 BL	45
<i>Tetrapterus angustirostris</i>	-	200 TL	52
<i>Tetrapterus belone</i>	-	240 BL	70
<i>Tetrapterus albidus</i>	130-210 BL	280 TL	82

<i>Tetrapterus audax</i>	140-280 BL	350 TL	200
<i>Istiophorus albicans</i>	125-210 BL	315 TL	58
<i>Istiophorus platypterus</i>	140-240 BL	340 TL	100
<i>Xiphias gladius</i>	115-190 BL	445 TL	540
<i>Makaira nigricans</i>	230-345 TL	375 TL	580
<i>Makaira indica</i>	170-210 BL	448 TL	700
<i>Makaira mazara</i>	200-300 BL	447 TL	900

Notes for billfishes: BL = Body Length, TL = Total Length

### References:

(1): Collette and Nauen (1983)

(2): Nakamura (1985)

## International Game Fish Association (IGFA) records

### Tunas and bonitos

Common name	Record size (in cm)	Record weight (in kg)	Location and year of capture
<i>Sarda chiliensis</i>	-	6.3	off Baja California, 1980
<i>Euthynnus lineatus</i>	-	11.8	off Baja California, 1991
<i>Euthynnus alleteratus</i>	92.7	12.2	off Florida, 1976
<i>Katsuwonus pelamis</i>	99	18.9	off Mauritius island, 1982
<i>Thunnus alalunga</i>	123	40	off Canary Islands, 1972
<i>Acanthocybium solandri</i>	-	72	off Baja California, 1996
<i>Thunnus albacares</i>	208	176.4	west coast of Mexico, 1977
<i>Thunnus obesus</i>	236	197.3	off Peru, 1957
<i>Thunnus thynnus</i>	304	679	off Nova Scotia, 1979

### Seerfishes and mackerels

Common name	Record size (in cm)	Record weight (in kg)	Location and year of capture
<i>Scomber japonicus</i>	-	1.9	off Baja California, 1986

### Swordfish and billfishes

Common name	Record size (in cm)	Record weight (in kg)	Location and year of capture
<i>Istiophorus albicans</i>	-	61.4	off Nigeria, 1991
<i>Tetrapterus albidus</i>	-	82.5	off Brazil, 1979

<i>Isuopnorus platypterus</i>	327.7	100.2	off Galapagos, 1947
<i>Tetrapterus audax</i>	-	224.0	off New Zealand, 1986
<i>Xiphias gladius</i>	445	536.1	off Chile, 1953
<i>Makaira mazara</i>	-	624.1	off Hawaii, 1982
<i>Makaira nigricans</i>	-	636.0	off Brazil, 1992
<i>Makaira indica</i>	442	707.6	off Peru, 1953

**Source:** IGFA (1995), note that IGFA records are not anymore on public domains and records might have changed since 1995.

## Sexual dimorphism

Sexual dimorphism is observed with billfishes. For example, Atlantic Blue marlin exhibits sexually dimorphic growth patterns: somatic growth of male slows at about 100 kg round weight and males do not exceed 150 kg, while females can reach up to 910 kg (Wilson *et al.*, 1991). Similarly, swordfish exhibit a sexual dimorphism of growth: males grow more slowly and reach a lower asymptotic length than females.

## Longevity

Longevities of tunas vary from a few years for the smaller tunas to 12 to 15 years for the larger tunas. The longevity record for tunas is about 20 years for the Atlantic bluefin tuna (Cort, 1990) or 25 years for the southern bluefin tuna (Gunn *et al.*, 2008). Longevities of 15 to 27 years (Pacific blue marlin) or 28 years (Indo-Pacific blue marlin) have been estimated for billfishes and for swordfish. Longevities of seerfishes and mackerels are moderate with some records at 16 years for the Spanish mackerel.

## Natural mortality

For larger tunas and billfishes, adult natural mortalities range from 0.2 to 0.6. Juvenile natural mortalities are higher. Little is known on natural mortalities of seerfishes and mackerels.

## Reproduction

### Spawning

#### Spawning behavior

Tuna spawn in open water close to the surface. Eggs are released by females in several batches. For example, yellowfin tuna in the Pacific spawn nearly every day. However, for some species like the bluefin species, spawning is more seasonal.

#### Spawning areas and seasons

Tunas spawn in areas where the survival of their larvae is greatest. Most species of tunas spawn only in waters where the surface temperatures are greater than 24°C. Tropical tunas appear to spawn in equatorial areas all year around and at higher latitudes during the warm seasons. Albacore and bigeye appear to migrate annually from temperate feeding areas to tropical spawning areas. Bigeye larvae are less abundant than those of other tropical tunas, and are found mainly in equatorial waters in which the temperatures are greater than 28°C (Collette and Nauen, 1983). Atlantic bluefin, Pacific bluefin and southern bluefin tuna exhibit a homing behavior when they mature, and return to restricted areas in the Atlantic, Pacific and Indian Oceans to spawn. It is commonly accepted that there is a homing behavior, but to a lesser extent, in yellowfin in the Atlantic Ocean. Billfishes appear to spawn seasonally in warm tropical and subtropical waters.

## Maturity and fecundity

### Maturity

With the exception of bluefin tunas (*Thunnus thynnus*, *T. orientalis* and *T. maccoyii*), most tunas, seerfishes, mackerels and billfishes reach their age of maturity between 2 and 5 years of age. Due to their sexual dimorphism of growth, male billfishes are mature at a smaller size than female billfishes.

### Maturity of tunas and billfishes

Tunas			
Scientific name	Age	Size	Weight
<i>Auxis rochei</i>		35 cm	
<i>Auxis thazard</i>		~30 cm	
<i>Euthynnus alleteratus</i>		40-50 cm	
<i>Euthynnus affinis</i>		45-50 cm	
<i>Katsuwonus pelamis</i>	2 years	Female: 42-50 cm Male: 45-52 cm	
<i>Thunnus alalunga</i>	5 years	90 cm	15 kg
<i>Thunnus albacares</i>	2.5-3 years	100-110 cm	20-30 kg

<i>Thunnus maccoyii</i>	11 years		
<i>Thunnus obesus</i>	3-3.5 years	100-110 cm	
<i>Thunnus thynnus</i> (East Atl.)	4 years	115 cm	30 kg
<i>Thunnus thynnus</i> (West Atl.)	8 years	190 cm	120 kg

### Bonitos

Scientific name	Age	Size	Weight
<i>Sarda sarda</i>		~40 cm	

### Seerfishes and mackerels

Scientific name	Age	Size	Weight
<i>Scomberomorus guttatus</i>	1-2 years	40-50 cm	
<i>Scomberomorus maculatus</i>	2 years	25-35 cm	
<i>Scomberomorus cavalla</i>	4 years	60-70 cm	
<i>Scomberomorus commerson</i>		70-90 cm	
<i>Acanthocybium solandri</i>	2 years	90 cm	



## Billfishes

Scientific name	Age	Size (LJFL)	Weight
<i>Tetrapterus pfluegeri</i>		150 cm	17-19 kg
<i>Tetrapterus albidus</i>		Female: ~150 cm Male: ~140 cm	
<i>Istiophorus albicans</i>		Female: 160-180 cm Male: ~140 cm	Female: 18-20 kg Male: 10 kg
<i>Xiphias gladius</i>	3.5 years	150-160 cm	
<i>Makaira nigricans</i>		Female: ~250 cm	120 kg
<i>Makaira indica</i>		Female: ~180 cm Male: 130-160 cm	Female: ~200 kg Male: 60-80 kg
<i>Makaira mazara</i>	2-4 years	130-140 cm	Female: 60-80 kg Male: 40-50 kg

## Fecundity

The batch fecundities of most species of tunas range from 2 to 70 million eggs, the lowest fecundity being for albacore and the highest for skipjack tuna and other small-sized tunas. Known batch fecundities of mackerels range from 300 000 to 1 500 000 eggs. Little is known on fecundities of seerfishes. Fecundity of wahoo has been estimated to 6 million eggs. Less is known on the reproductive biology of billfishes, but batch fecundity is estimated to range between 1 and 7 millions of ovocytes. Swordfish batch fecundity was estimated to 3.9 millions eggs in the Atlantic.

Maturity and fecundity parameters of the principal market species of tunas		
Scientific name	Size and age at maturity	Annual batch fecundity
<i>Katsuwonus pelamis</i>	about 3 years and 42 to 45 cm	7 to 76 million eggs
<i>Thunnus alalunga</i>	about 5 years and 90 cm	2 to 3 million eggs
<i>Thunnus albacares</i>	about 3 years and 100 cm	4 to 60 million eggs

<i>Thunnus obesus</i>	about 3 years and 100 cm	4 to 60 million eggs
<i>Thunnus thynnus</i>	about 4 years and 105 to 120 cm	5 to 30 million eggs

## Sex ratio

It has been shown that for yellowfin, bigeye and albacore, the sex-ratio changes with the age of the fish with a predominance of males for the larger sizes. A predominance of females has also been observed for medium-sized Atlantic bluefin tuna. For skipjack, differences in the numbers of males and females have been observed locally. Predominance of females at older ages is observed for several species of billfishes.

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## Bibliography

Allain, V., 2005. Diet of four tuna species of The Western and Central Pacific Ocean. *SPC Fisheries Newsletter* **114** (2005): 30-33.

Bard, F.X., E. Josse and P. Bach, 1998. Habitat, écophysiologie des thons. ICCAT symposium.

Beamish, F.W.H., 1978. Swimming capacity. pp 101-187. *In: Fish Physiology*, Vol. 7. W.S. Hoar and D.J. Randall (eds), Academic Press, New York.

Berg, L.S., 1958. System der rezenten und fossilen Fischartigen und Fische. VEB Verlag der Wissenschaften, Berlin.

Block, B.A., D.T. Booth and F.G. Carey, 1992a. Depth and temperature of the blue marlin, *Makaira nigricans*, observed by acoustic telemetry. *Mar. Biol.*, **114**: 175-183.

Block, B.A., D.T. Booth and F.G. Carey, 1992b. Direct measurement of swimming speeds and depth of blue marlin. *J. Exp. Biol.*, **166**: 267-284.

Block, B. A., S. L. H. Teo, A. Walli, A. Boustany, M. J. Stokesbury, C. J. Farwell, K. C. Weng *et al.*, 2005. Electronic tagging and population structure of Atlantic bluefin tuna. *Nature* **434**: 1121-1127.

Brill, R., 1994. A review of temperature and oxygen tolerance studies of tunas pertinent to fisheries oceanography, movements models and stock assessments. *Fish. Oceanogr.*, **3** (3): 204-216.

Brill, R. W. and M. E. Lutcavage, 2001. Understanding environmental influences on movements and depth

distributions of tunas and billfishes can significantly improve population assessments. *American Fisheries Society Symposium* **25**: 179-198.

Bushnell, P.G. and K.N. Holland, 1997. Tunas. *Virginia Mar. Res. Bull.*, **29** (1 & 2): 3-6.

Carey, F.G., J.M. Teal, J.W. Kanwisher, K.D. Lawson and J.S. Beckett, 1971. Warm bodied fish. *Am. Zool.*, **11** (1): 137-145.

Carey, F.G. and B.H. Robison, 1981. Daily patterns in the activities of swordfish, *Xiphias gladius*, observed by acoustic telemetry. *Fish. Bull. U.S.*, **79**: 277-291.

Cayré, P., J.B. Amon Kothias, J.M. Stretta and T. Diouf, 1988. La biologie des thons. pp 167-183 in A. Fonteneau and J. Marcille (eds), Ressources, pêche et biologie des thonidés tropicaux de l'Atlantique centre-est, *FAO Fish. Tech. Pap.*, **292**, Rome: 391 p.

Chow S, K. Nohara, T. , Tanabe, T. Itoh, S. Tsuji, Y. Nishikawa, S. Uyeyanagi, K.Uchikawa, 2003. Genetic and morphological identification of larval and small juvenile tunas (Pisces: Scombridae) caught by a mid-water trawl in the western Pacific. *Bull. of Fisheries Research Agency* **8**: 1-14.

Collette, B.B., 1978. Adaptations and systematics of the mackerels and tunas. pp 7-39 in G.D. Sharp and A.D. Dizon (eds), *The Physiological Ecology of Tunas*, Academic Press, New-York: 485 p.

Collette, B.B. and C.E. Nauen, 1983. FAO Species catalogue, vol. 2. Scombrids of the worlds. An annotated and illustrated catalogue of tunas, mackerels, bonitos and related species known to date. *FAO Fish. Synop.*, **125** (2), Rome: 137 p.

Collette, B.B., J.R McDowell and J.E. Graves, 2006. Phylogeny of recent billfishes (xiphoidei). *Bull. Mar. Sci.* **79**(3): 455-468.

Cort, J.L., 1990. Biología y peca del atún rojo, *Thunnus thynnus* (L.), del Mar Cantábrico. *Inst. Español Ocean., Publ. Esp.*, **4**: 272 p.

Dagorn L., K.N. Holland, J-P. Hallier, M. Taquet, G. Moreno, G. Sancho, D. G. Itano, R. Aumeeruddy, C. Girard, J. Million and A. Fonteneau. Deep diving behavior observed in yellowfin tuna (*Thunnus albacares*). *Aquat. Living Resour.* (2006)**19**, 85–88.

Dickson, K.A., 1995. Unique adaptations of the metabolic biochemistry of tunas and billfishes for life in the pelagic environment. *Env. Biol. Fish.*, **42**: 65-97.

Elliott, N.G. and R.D. Ward, 1995. Genetic relationships of eight species of Pacific tunas (Teleostei: Scombridae) inferred from allozyme analysis. *Mar. Fresh. Res.* **46** (7): 1021-1032.

Fromentin, J.-M. and A. Fonteneau, 2001. Fishing effects and life history traits: a case-study comparing tropical versus temperate tunas. *Fisheries Research* **53**:133-150.

Graham, J.B., F.J. Koehn and K.A. Dickson, 1983. Distribution and relative proportions of red muscle in scombrid fishes: consequences of body size and relationships to locomotion and endothermy. *Can. J. Zool.*, **61**: 2087-2096.

Gunn, J.S., Clear N.P., Carter T.I., Rees A.J., Stanley C.A., Farley J.H., Kalish J.M., 2008. Age and growth in southern bluefin tuna, *Thunnus maccoyii* (Castelnaud): Direct estimation from otoliths, scales and vertebrae. *Fisheries Research* **92**(2-3): 207-220.

Holland, K.N., R.W. Brill, R.K.C. Chang, J.R. Sibert and D.A. Fournier, 1992. Physiological and behavioural

thermoregulation in bigeye tuna. *Nature*, **358**: 410-412.

Joseph, J. W. Klawe and P. Murphy, 1988. Tuna and Billfish - fish without a country. 4<sup>th</sup> edition, Inter-American Tropical Tuna Commission (ed.), La Jolla, California: 69 p.

Klawe, W.L., 1976. Tuna as an English word for a scombrid fish. *Inter-Am. Trop. Tuna Comm. (Unpubl. Manuscr.)*.

Klawe, W.L., 1977. What is a tuna? *Marine Fisheries Review*, **39** (11), paper 1268: 5 p.

Lee, R.E.K.D., 1982. Thailand. Fishing for tuna. A report prepared for the pole-and-line fishing in Southern Thailand Project. FAO. FI: DP/THA/77/008:65 p.

Lowe, T.E., R.W. Brill, and K.L. Cousins, 2000. Blood oxygen-binding characteristics of bigeye tuna (*Thunnus obesus*), a high-energy-demand teleost that is tolerant of low ambient oxygen. *Marine Biology*, **136**: 1087-1098.

Magnuson, J.J., 1973. Comparative study of adaptations for continuous swimming and hydrostatic equilibrium of scombroid and xiphoid fishes. *Fish Bull.*, **71** (2):337-356.

Magnuson, J.J., 1978. Locomotion by scombrid fishes: hydrodynamics, morphology, and behavior. *Fish Physiol.*, **7**: 239-313.

Ménard, F., B. Stéquert, A. Rubin, M. Herrera, E. Marchal, 2000. Food consumption of tuna in the Equatorial Atlantic ocean: FAD-associated versus unassociated schools. *Aquat. Living Resour.*, **13** (2000): 233-240.

Nakamura, I, 1985. FAO Species catalogue, vol. 5. Billfishes of the world. An annotated and illustrated catalogue of marlins, sailfishes, spearfishes and swordfishes known to date. *FAO Fish. Synop.*, **125** (5), Rome: 65 p.

Nishikawa Y., DW Rimmer, 1987. CSIRO Mar. Lab. Report 186. Identification of Larval Tunas, Billfishes and other Scombroid Fishes (Suborder *Scombroidei*): an Illustrated Guide : 20pp.

Olson, R.J. and H. Boggs, 1986. Apex predation by yellowfin tuna (*Thunnus albacares*): independent estimates from gastric evacuation and stomach contents, bioenergetics and cesium concentrations. *Can. J. Fish. Aquat. Sci.* **43** (9):1760-1775.

Orbesen, E.S., J.P. Hoolihan, J.E. Serafy, D. Snodgrass, E. Peel, and E.D. Prince, 2008. Transboundary Movements of Atlantic Istiophorid Billfish Among International and US Domestic Management Areas Inferred by Mark-Recapture Studies. *Marine Fisheries Review*, **70** (1): 14-23.

Orrell, T.M., B.B. Collette, and G.D. Johnson, 2006. Molecular data support separate clades for Scombroidei (tunas and relatives) and Xiphioidei (billfishes). *Bull. Mar. Sci.* **79**(3): 505-519.

Ortiz, M., E.D. Prince, J.E. Serafy, D.B. Holts, K.B. Dary, J.G. Pepperell, M.B. Lowry and J.C. Holdsworth, 2003. Global overview of the major constituent-based billfish tagging programs and their results since 1954. *Mar. Freshwater Res.*, **54**:489-507.

Partridge, B.L., 1982. The structure and function of fish schools. *Scient. Amer.*, **247**: 114-123.

Roger, C., 1994. The plankton of the tropical western Indian ocean as a biomass indirectly supporting surface tunas (yellowfin, *Thunnus albacares* and skipjack, *Katsuwonus pelamis*. *Environ. Biol. Fish.* **39** (2):161-172.

Roberts, J.L., 1978. Ram gill ventilation in fish. pp. 83-88 in : G.D. Sharp and A.D. Dizon (eds), *The Physiological Ecology of Tunas*, Academic Press, New-York: 485 p.

Sedberry, S.R. and J.K. Loefer, 2001. Satellite telemetry tracking of swordfish, *Xiphias gladius*, off the eastern United States. *Mar. Biol.* **139**: 355-360.

Sharp, G.D., 1978. Behavioural and physiological properties of tunas and their effects on vulnerability to fishing gears. pp 397-450 In G.D. Sharp and A.E. Dizon (eds), *The physiological ecology of tunas*, Academic Press, New York: 485 p.

Sharp, G.D. and A.D. Dizon, 1978. *The Physiological Ecology of Tunas*. Academic Press, New-York: 485 p.

Stevens, E.D. and W.H. Neill, 1978. Body temperature relations of tunas, especially skipjack. pp 315-359 in : *Fish Physiology*, vol VII (ed. W.S. Hoar and D.J. Randall), New York : Academic Press.

Sund, P.N., M. Blackburn and F. Williams, 1981. Tunas and their environment in the Pacific Ocean: a review. *Oceanogr. Mar. Biol. Ann. Rev.*, **19**: 443-512.

Suzuki, Z., Y. Warashina, M. Kishida, 1977. The comparison of catches by regular and deep longline gears in the Western and Central Equatorial Pacific. *Bull. Far Seas Res. Lab.*, **15**: 51-90.

Takahashi, M., H. Okamura, K. Yokawa and M. Okazaki, 2003. Swimming behaviour and migration of a swordfish recorded by an archival tag. *Marine and Freshwater Research* 54 (4): 527-534.

Wardle, C.S. and P. He., 1988. Burst swimming speeds of mackerel, *Scomber scombrus* L. *J. Fish Biol.* **32** (3): 471-478.

Webb, P.W., 1984. Body form, locomotion and foraging in aquatic vertebrates. *Amer. Zool.*, **24**: 107-120.

Weinheimer, M., 2003. "Scombridae". Animal Diversity Web:  
<http://animaldiversity.ummz.umich.edu/site/accounts/information/Scombridae.html>.

Wilson, C.A., J.M. Dean, E.D. Prince, and D.W. Lee, 1991. An examination of sexual dimorphism in Atlantic and Pacific blue marlin using body weight, sagittae weight, and age estimates. *J. Exp. Mar. Biol. Ecol.* **151**: 209-225.

