

Fusarium wilt of bananas: Research advances, needs and regional initiatives



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What countries are doing Research on Fusarium wilt in LAC?

- ✓ Brazil (exports is limited, but national market rely on susceptible genotypes)
- ✓ Cuba (no export - it has been active because susceptible varieties and national interest)
- ✓ Venezuela (Almost no export)
- ✓ In countries with more export tradition Fusarium wilt has not been a priority in national research agenda in the last 60 years:
 - Ecuador
 - Costa Rica* (re-starting)
 - Colombia (at the beginning)

Little attention to a Foc in LAC in the last 10 years, even with the TR4 threat

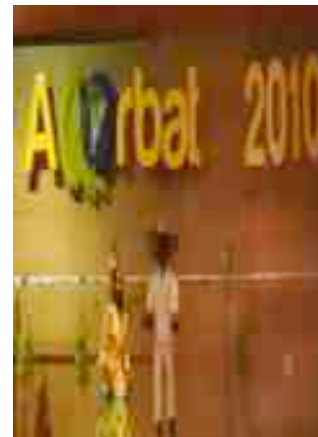
2004

2006

2008

2010

2013



| | |
|------------------|----------|
| Sigatoka = | 22 |
| Nematodes = | 4 |
| Others = | 6 |
| Fusarium= | 1 |

| | |
|-------------------|----------|
| Sigatoka = | 15 |
| Nematodes = | 4 |
| Others = | 13 |
| Fusarium = | 1 |

| | |
|-----------------|----------|
| Fusarium | 3 |
| Sigatoka | 20 |
| Nematodes | 5 |
| Other | 95 |
| Total | 123 |

| | |
|-----------------|----------|
| Fusarium | 6 |
| Sigatoka | 24 |
| Nematodes | 0 |
| Others | 59 |
| Total | 89 |

Fusarium: 11

Total: 145

Low number of indexed publications

Research Targets on Foc Tropical Race 4



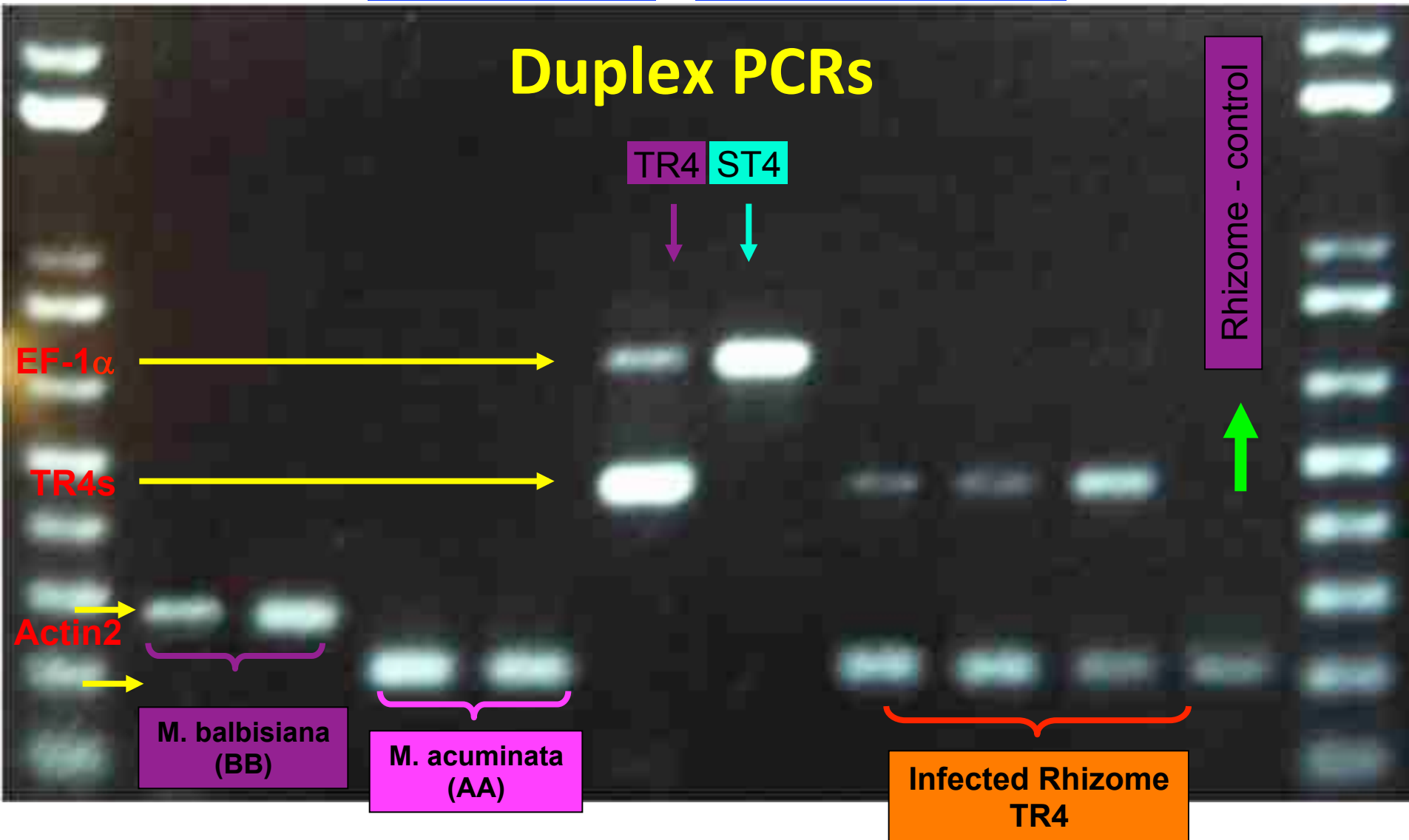
1. Prevention - early detection
2. Use of resistant varieties
3. Disease management

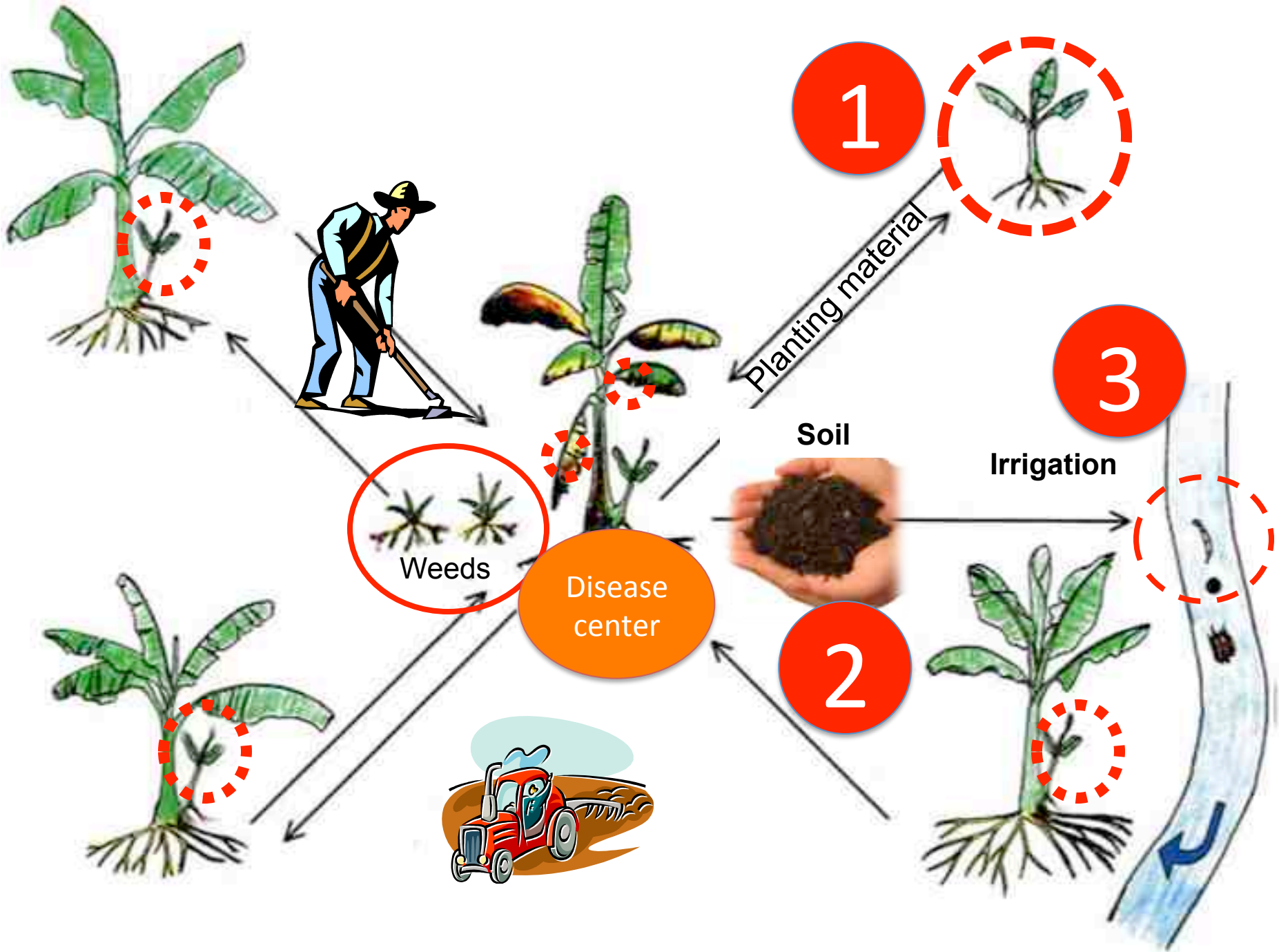
- a. Diagnostic tools for Foc TR4 and other important races/strains of Foc;
- b. Fast and reliable bioassays for high throughput phenotyping
- c. Characterize genotypes of Musa for TR4 resistance/ Identify Resistant sources
- d. Understand Foc- banana interaction at genetic, cytological and molecular level;
- e. Epidemiology for integrated management strategies - Eradication

Foc TR4 – Diagnostic

EF +FocTR4 // Actin2 + FocTR4

Duplex PCRs

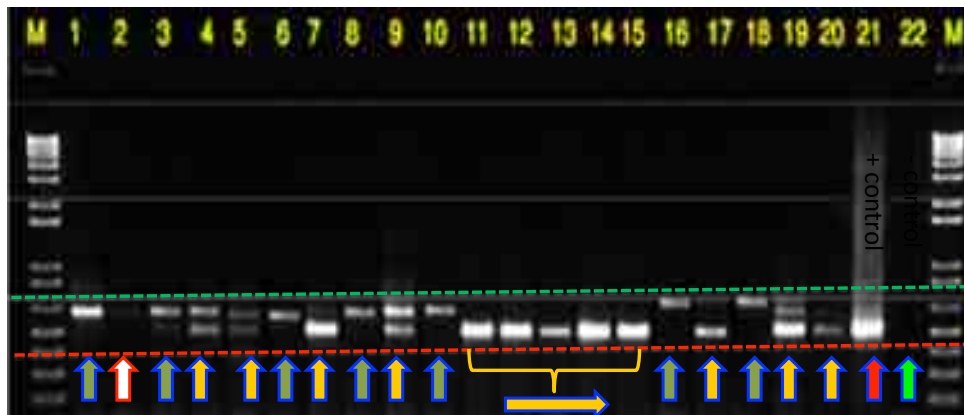




Detecting Foc TR4 in soil and symptomless tissues



DNA extraction



| Sample ID | Cultivar ^a | Plant stage | Location | Source | Sample processed | Single PCR | Nested PCR |
|-----------|-----------------------|-------------|----------|-------------|-------------------------|------------|------------|
| AuD1 | Cavendish | Symptomatic | Darwin | Australia | Plant | + | nd |
| AuH1 | Cavendish | Symptomless | Darwin | Australia | Plant | + | nd |
| TS1 | Cavendish | Symptomless | Chaojhou | Taiwan | Soil | - | + |
| | | | | | Plant | + | nd |
| TS3 | Cavendish | Symptomatic | Chaojhou | Taiwan | Soil | + | + |
| | | | | | Plant | + | nd |
| TS6 | Cavendish | Symptomless | Wandan | Taiwan | Soil | - | - |
| | | | | | Plant | - | nd |
| TS7 | Cavendish | Symptomatic | Jiuru | Taiwan | Soil | + | + |
| | | | | | Plant | + | nd |
| TS8 | Cavendish | Symptomless | Jiuru | Taiwan | Soil | - | + |
| | | | | | Plant | + | nd |
| TS9 | Cavendish | Symptomatic | Luye | Taiwan | Soil | + | + |
| | | | | | Plant | + | nd |
| Phi126C | Gran Naine | Symptomatic | Kapalong | Philippines | Plant | + | nd |
| Phi39B | Tall William | Symptomatic | Kapalong | Philippines | Plant | - | nd |
| Phi2SV | Latundan | Symptomatic | Kapalong | Philippines | Plant | - | nd |
| ChlamyD2 | n.a. | n.a. | n.a. | n.a. | Foc-colonized Substrate | + | + |



ITS

TR4

Water is an efficient way for FOC dissemination....

- Can we detect Foc TR4 easily in water samples?
- What would be the app/implications?



Risk Assessment and Surveillance of Foc TR4

Strengthening capacities and increasing preparedness for Foc TR4 in LAC

Bioversity,
MUSALAC
&
National Plant
Protection
Organizations



- ▲ Trained
- ▲ In the pipeline



RECOMENDACIONES PARA PREVENIR LA ENTRADA DE ENFERMEDADES CUARENTENARIAS DE ALTA IMPORTANCIA ECONÓMICA PARA BANANO Y PIÑA.

Dirigidas a: Empleados de las compañías fruteras, representantes de compañías de agroquímicos, inspectores de Compañías Certificadoras, representantes de ONG's, y cualquier otro visitante a plantaciones de banano en el Sur Este Asiático,

RECOMMENDATIONS TO PREVENT THE ENTRANCE OF QUARANTINE PLANT DISEASES OF MAJOR ECONOMIC IMPORTANCE FOR BANANA AND PINEAPPLE

Aimed at: Employees of the major fruit producing and processing companies, representatives of agrochemical companies and NGOs, certification inspectors, and any other visitor to banana plantations in South East Asia and pineapple plantations in South America.

RECOMMANDATIONS POUR PRÉVENIR L'ENTRÉE DE MALADIES DE QUARANTAINE DE GRANDE IMPORTANCE ÉCONOMIQUE POUR LES BANANIERS ET L'ANANAS.

Cible : Employés de compagnies fruitières, représentants de compagnies phytosanitaires, inspecteurs de Compagnies de Certification, représentants d'ONG, et pour tout visiteur de plantations de banane en Asie du Sud-est, et de plantations d'ananas en Amérique du Sud.

Contingency plan for Foc TR4

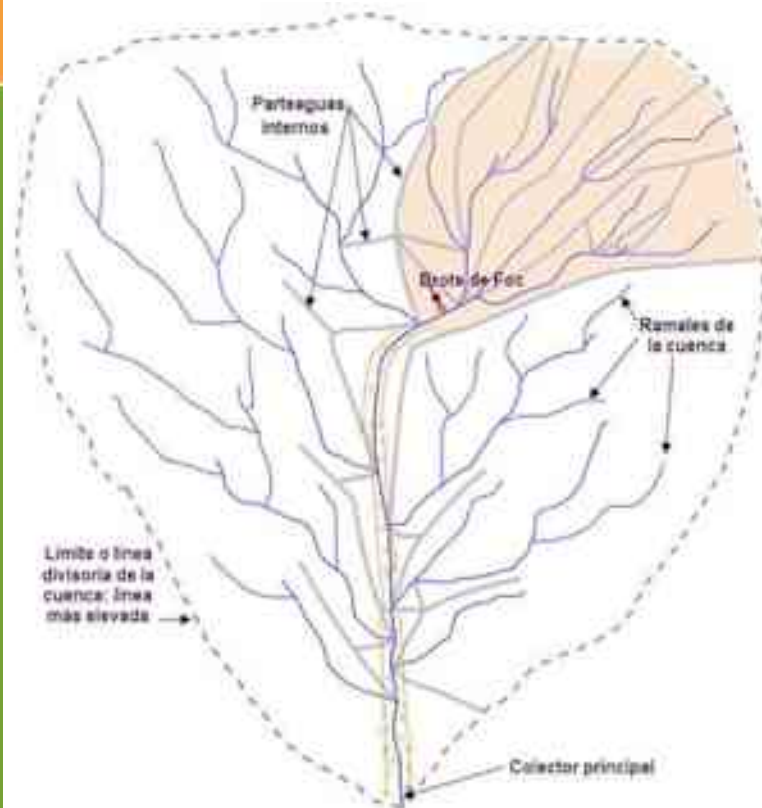
ORGANISMO INTERNACIONAL REGIONAL DE SANIDAD AGROPECUARIA
OIRSA

Plan de contingencia ante un brote de la raza 4 tropical de *Fusarium oxysporum* f. sp. *cubense*

En un país de la región del OIRSA



Elaborado por:
Miguel Ángel Dita Rodríguez
Plutarco Elías Echegoyén Ramos
y
Luis Fernando Pérez Vicente



Good example : Caravan for management Helicoverpa and other pests : Embrapa 2014

Caravana Embrapa



<http://www.cnpso.embrapa.br/caravana>

It is now too late? How to proceed?

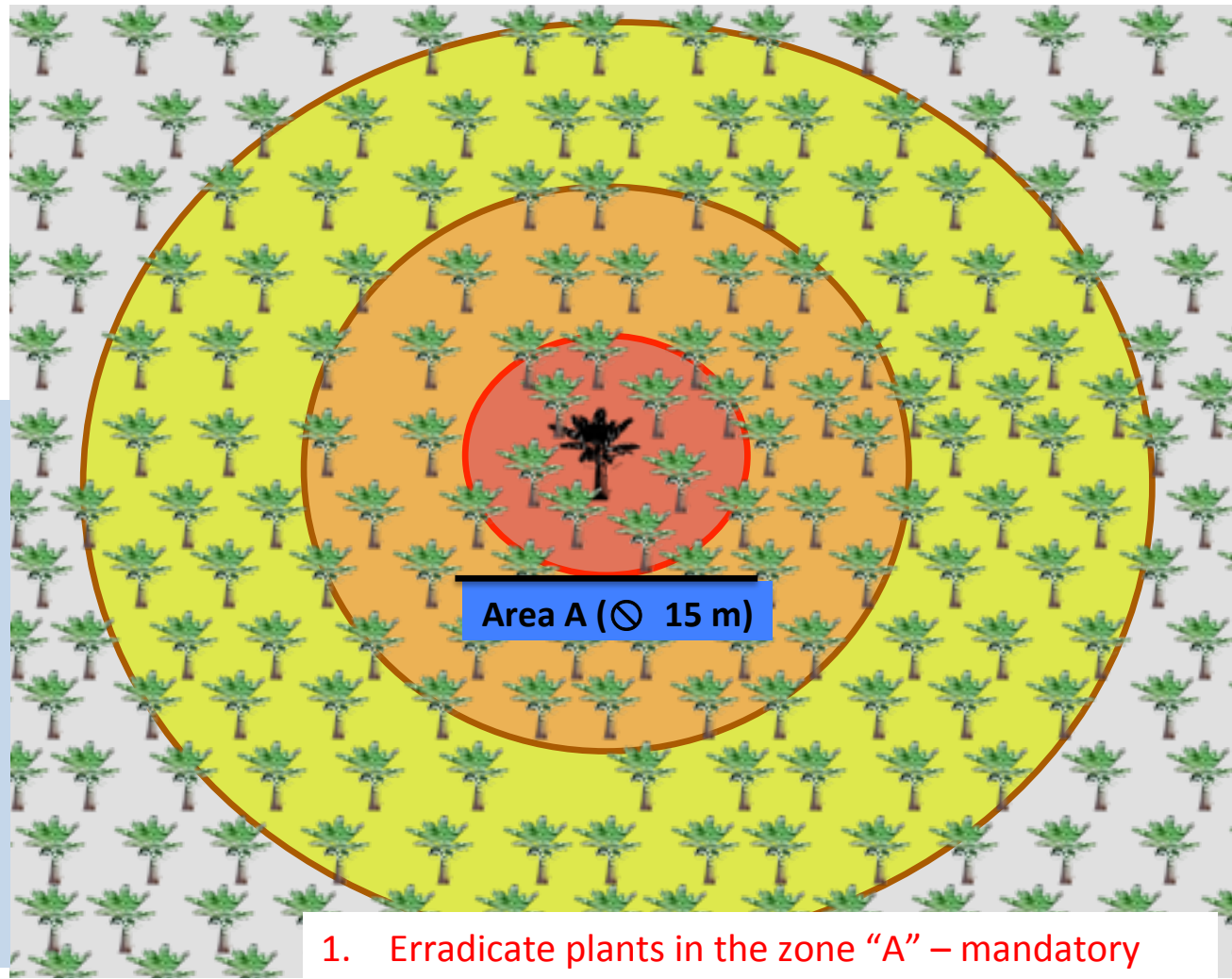
A. Molina



Early diagnostic of Fusarium wilt vs. management

When a plant is detected infected how many 'neighbors' we should destroy around?

Farmer and companies think more on bunches and boxes... they don't care too much on inoculum pressure, epidemiology ?
...but at the end of the day the scenario just becomes worst



1. Erradicate plants in the zone "A" – mandatory
2. Erradicate plant in zone B - Recommended

What else needs to be done on the the pathogen side?

Establishment and characterization of national collections

- Population biology studies
- Molecular characterization using informative markers (i.e genes related to pathogenicity/aggressiveness)
- VCG analyses
- Survey using specific primers for TR4
- Characterization for pathogenicity and aggressiveness on reference cultivars
- Identification and characterization of pathogenesis/aggressiveness -related genes



Population biology of *Foc* in Brazil (Costa et al., 2014)



Plant Pathology (2014)

Doi: 10.1111/ppa.12242

Genetic structure of *Fusarium oxysporum* f. sp. *ubense* in different regions from Brazil

S. N. Costa^{af}, C. A. D. Bragança^{bf}, L. R. Ribeiro^a, E. P. Amorim^c, S. A. S. Oliveira^c, M. A. Dita^c, F. F. Laranjeira^c and F. Haddad^{af}

^aFederal University of Rio de Janeiro de Bahia; ^bCAPES/Embrapa; and ^cEmbrapa Cassava & Fruit, Cruz das Almas 44380-000, Brazil

- 05 major regions
- 214 isolates
- 52 haplotypes detected

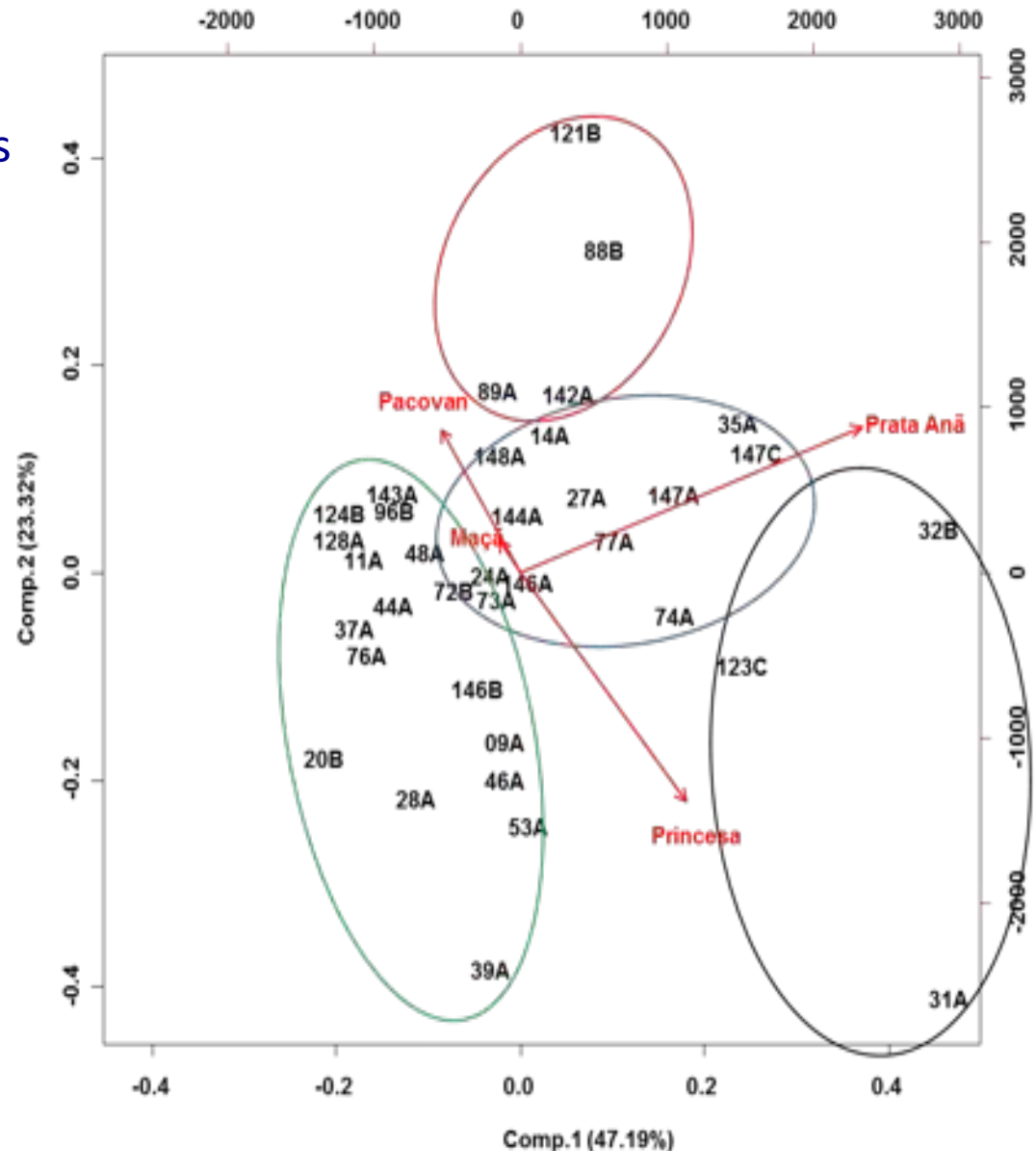


Migrations are represented by curves. Green curves: migrations between BA and other states; blue curves: migrations between MG and other states; brown curves: migrations between CE and other states; Yellow curve: migration between RN and RS. Nm: Number of migrants.

Population biology of FOC in Brazil (Costa et al., 2014)

Differential Interaction of isolates representing 52 haplotypes with varieties

Maça (Silk) = Susceptible
Prata anã = Susceptible
Pacovan : Moderately Resistant
Princesa : Resistant



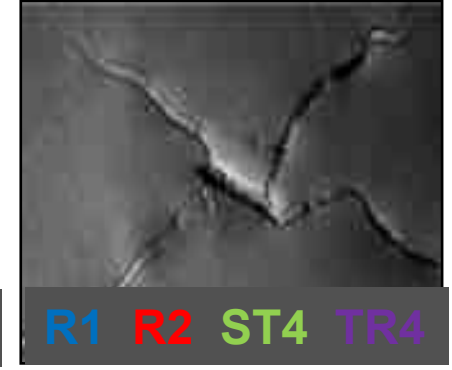
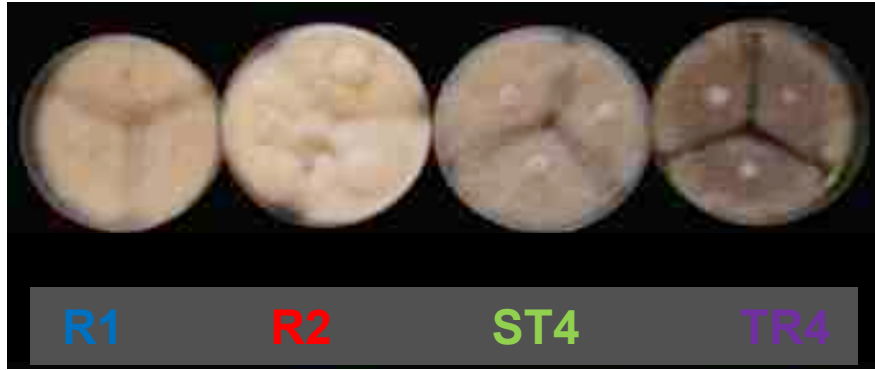
More info with: Dr. Haddad
fernando.haddad@embrapa.br

Comparative transcriptomic analysis of Foc [R1, R2, ST4 & TR4]

Mycelia

Conidia

Germinated conidia



RNA extraction

RNA pooling (mycelium + conidia + germinating conidia)

mRNA Isolation

cDNA from R1, R2, ST4, TR4 and 454 sequencing

Comparative EST analyses

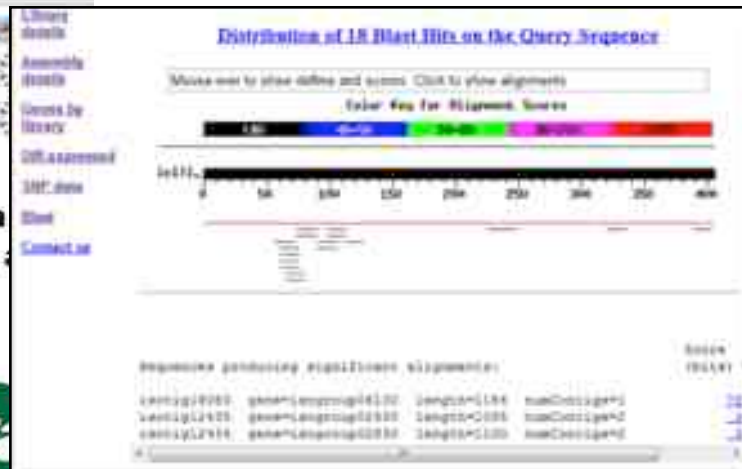
Foc web server: software, analysis, interaction

www.lba.cnptia.embrapa.br
www.lba.cnptia.embrapa.br/fusarium/



Home
Library details
Assembly details
Genes by library
Diff. expressed
SNP data
Blast
Contact us

Fusarium oxysporum transcriptome

Home
Library details
Assembly details
Genes by library
Diff. expressed
SNP data
Blast
Contact us

BLAST

Choose program to use and database to search:

Program: **blastn** Database: **Fusarium_Fox**

Enter sequence below in FASTA

Options: Use subject from file Check for repeats

Use subject from File Run reads in a single set Check related species: *F. lycopersi*, *F. verticillioides* and *F. graminearum*

The query sequence is **BLAST**

[Help](#) [Low complexity](#) [1](#) [5](#)

Expand [19](#) [Main](#) [BL00AMJ](#) [Perform suggested alignment](#)

Home
Library details
Assembly details
Genes by library
Diff. expressed
SNP data
Blast
Contact us

Description: Fisher's exact test between B1 and B2 Reads

This table displays significant gene terms B1 and B2 reads. Each gene is given a gene name in the associated transcriptome, which includes the normalized ERF12 value of each gene in the library B1, related to each of the normalized ERF12 reads of each gene in the library B2. log2(Fold change) represents the ratio B1 and B2 reads after adjustment. If positive, B1 is more expressed than B2, otherwise B2 is more expressed than B1. log2(Fold change) calculated is a combination of log2(Fold change) ratio, p-value adjustment (significant for each), as well as the ratio between the significant difference p-value (Saperas et al. 1998) is the false discovery rate (FDR) value. It used to increase the proportion of false positive rejected called the false discovery rate when the proportion that is rejected is equal to rate (Storey et al. 2002) number method to increase the proportion of false positives rejected called the false discovery rate. This gene is not a called significant. Significant p-value is not indicated by TEDE. It is also because the difference is smaller or equal than 0.05 (FDR) value defined for the corresponding gene. It will only be the library B1, B2 or both.

| GeneName | value1 | value2 | log2(Fold change) | log2(Fold change) nonadjusted | P |
|------------|---------|---------|-------------------|-------------------------------|---|
| contig1001 | 2007.31 | 18.2661 | 8.77987911658408 | 6.77643293005345 | 0 |

www.lba.cnptia.embrapa.br/Fusarium_Fox/blast/contig1001

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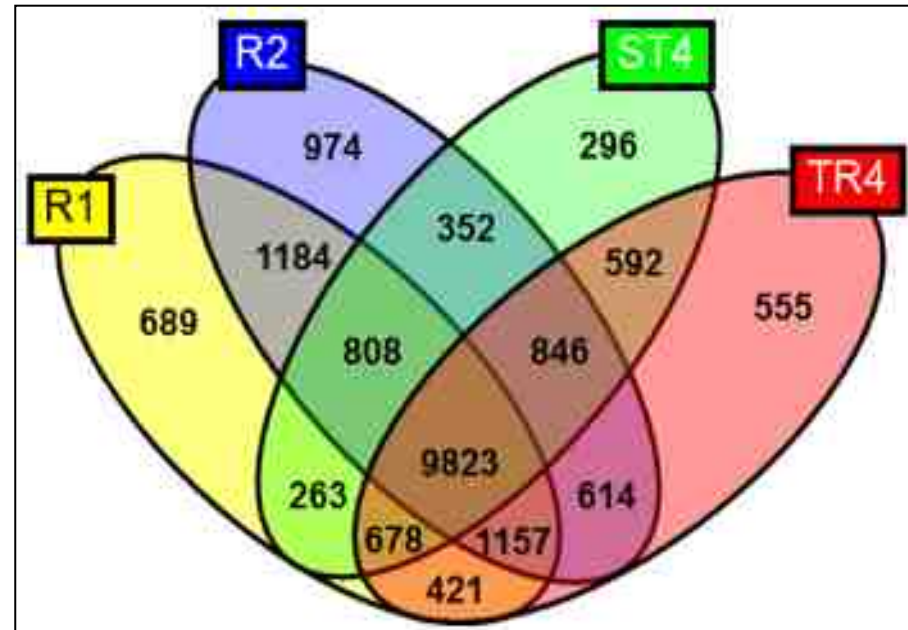
3490190196 gene#langroup00645 length=1472 nonCodingP1
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TCTTTGTTTATTATTTCTTATTTCTGATATGAGAGCACTGATATATGATATGATATGATAT
ACTTCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATAT
TTTTTCCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGAT
AAAAGCAATGCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGAT
GCTATTTTATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGAT
CGAAGCAGTCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGAT
GATGATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGAT
CGCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATG
CGCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATG
CACCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATG
GCCTGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATG
GATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATGATATG
  
```

Comparative transcriptomic analysis of Foc [R1, R2, ST4 & TR4]

Library statistics

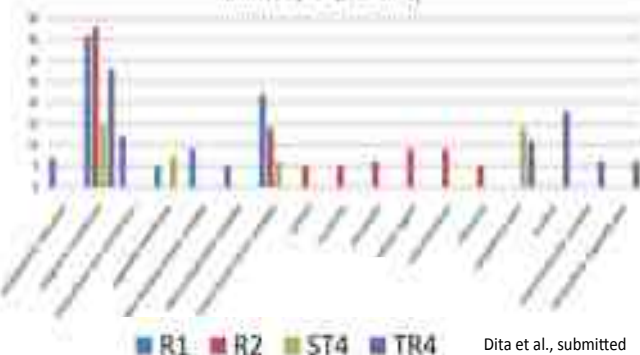
| | R1 | R2 | ST4 | TR4 | All Libraries |
|--------------------|---------|---------|---------|---------|---------------|
| Raw reads | 644.444 | 661.201 | 633.453 | 606.533 | 2.545.631 |
| rDNA | 2.988 | 3.255 | 1.811 | 2.746 | 10.800 |
| Passed reads | 637.670 | 653.748 | 629.498 | 601.136 | 2.522.052 |
| aver. length | 396 | 392 | 364 | 371 | 381 |
| % Singlets | 6.2 % | 6.3 % | 5.5 % | 7.2 % | 5.3 % |
| Assembled reads | 541.632 | 558.903 | 554.771 | 557.094 | 2.151.030 |
| # isogroups | 8.302 | 9.059 | 7.610 | 8.193 | 11.058 |
| # isotigs >50nt | 9.674 | 11.114 | 8.500 | 10.216 | 21.445 |
| mean isotig length | 1345.8 | 1440.5 | 1293.8 | 1406.3 | 2161.9 |
| Longest Isotig | 8.293 | 9.732 | 7.276 | 7.752 | 11.432 |

Gene distribution across Foc races

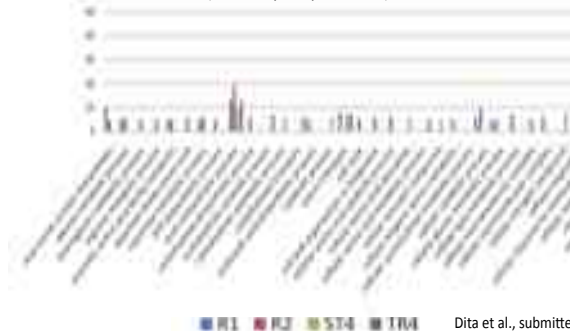


Annotation of UNIQUE genes

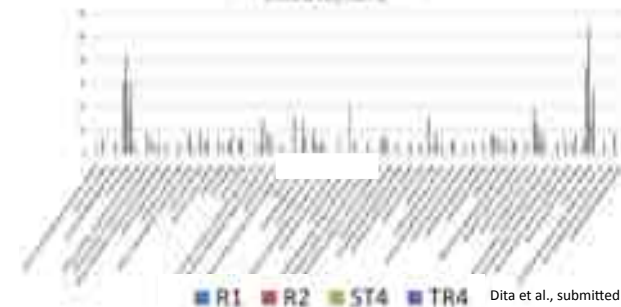
Sequence distribution (cellular component)
(filtered by #Seqs, cutoff=5)



Biological processes
(filtered by #Seqs, cutoff=5)



Sequence distribution (molecular function)
(filtered by #Seqs, cutoff=5)

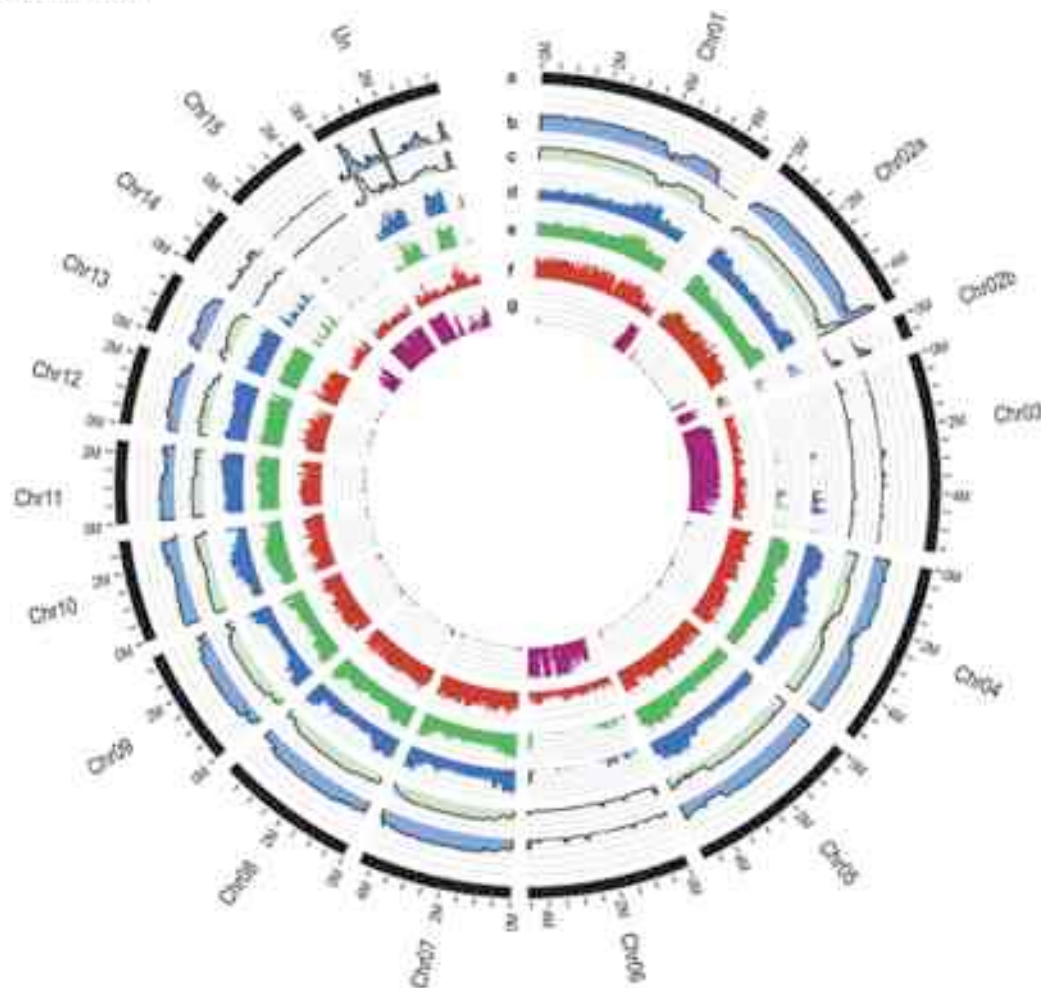




Genome and Transcriptome Analysis of the Fungal Pathogen *Fusarium oxysporum* f. sp. *ubense* Causing Banana Vascular Wilt Disease

Lijia Guo^{1*3}, Lijuan Han²³, Laying Yang¹³, Huicai Zeng², Dingding Fan³, Yabin Zhu³, Yue Feng³, Guofen Wang¹, Chunfang Peng³, Xuanting Jiang³, Dajie Zhou³, Peixiang Ni³, Changcong Liang¹, Lei Liu¹, Jun Wang¹, Chao Mao¹, Xiaodong Fang²¹, Ming Peng²¹, Junsheng Huang¹¹

1 Key Laboratory of Monitoring and Control of Tropical Agricultural and Forest Invasive Alien Pests, Ministry of Agriculture, Environment and Plant Protection Institute, Chinese Academy of Tropical Agricultural Sciences, Hainan, China, **2** Institute of Tropical Bioscience and Biotechnology, Chinese Academy of Tropical Agricultural Sciences, Hainan, China, **3** BGI-Sherzhen, Shenzhen, China

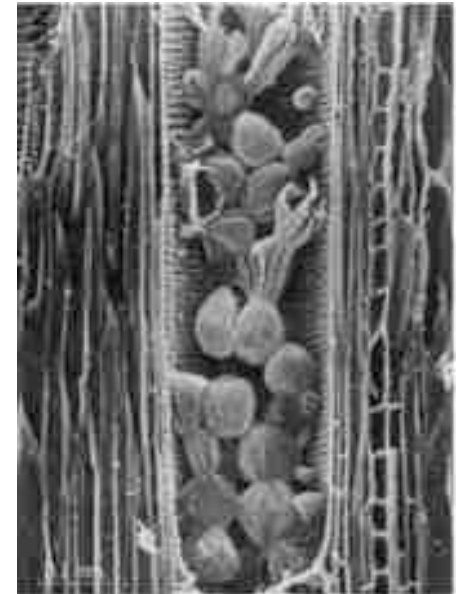
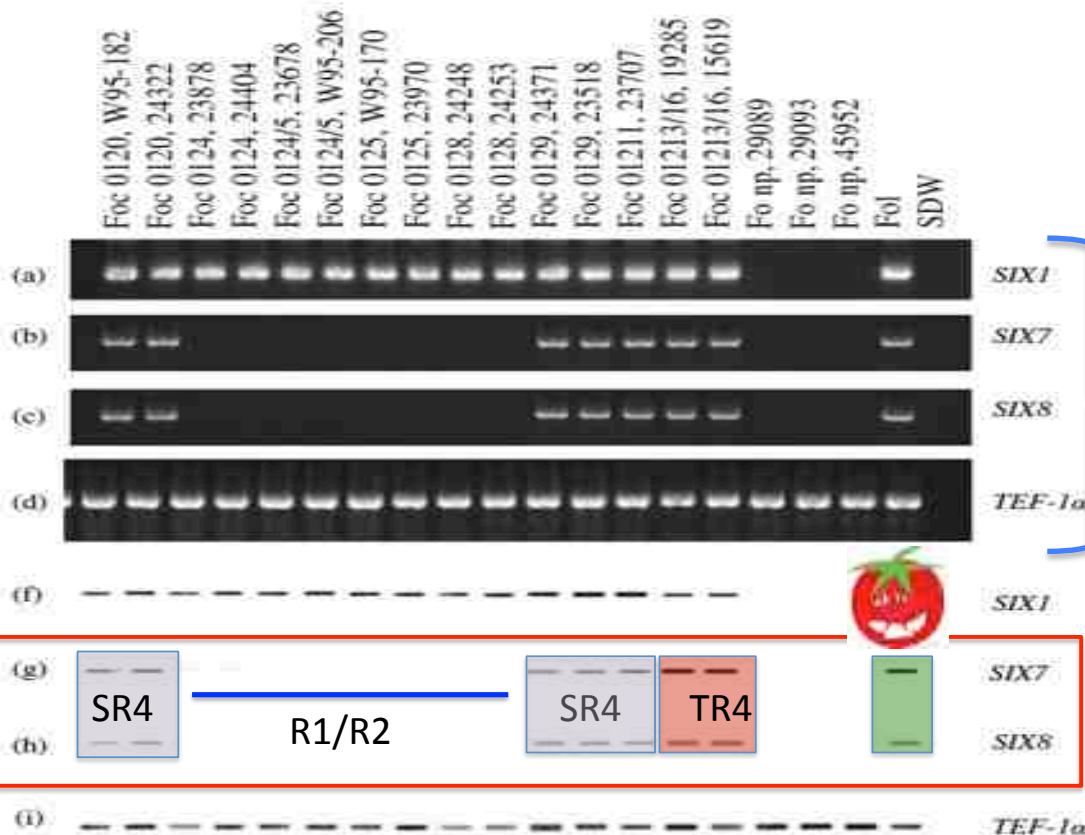


Presence of putative pathogenicity genes in isolates of *Fusarium oxysporum* f. sp. *ubense* from Australia

R. A. Meldrum • S. Fraser-Smith •
 L. T. T. Tran-Nguyen • A. M. Daly • E. A. B. Aitken

Secreted in Xylem Genes – SIX

Presence of putative pathogenicity genes



<http://www.accessscience.com/popup.aspx?figID=524000FG0060&id=524000&name=figure>

(G. E. VanderMolen
 University of Rhode Island)

PCR

Slot blot



Genome and Transcriptome Analysis of the Fungal Pathogen *Fusarium oxysporum* f. sp. *cubense* Causing Banana Vascular Wilt Disease

Lijia Guo^{1*†}, Lijuan Han^{3†}, Laying Yang^{1,3}, Huicai Zeng², Dingding Fan³, Yabin Zhu³, Yue Feng³, Guofen Wang¹, Chunfang Peng³, Xuanting Jiang³, Dajie Zhou³, Peixiang Ni³, Changcong Liang¹, Lei Liu¹, Jun Wang¹, Chao Mao¹, Xiaodong Fang^{3†}, Ming Peng^{2†}, Junsheng Huang^{1†}

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Table 2. The orthologs of *SIX*-genes in *Foc1* and *Foc4*.

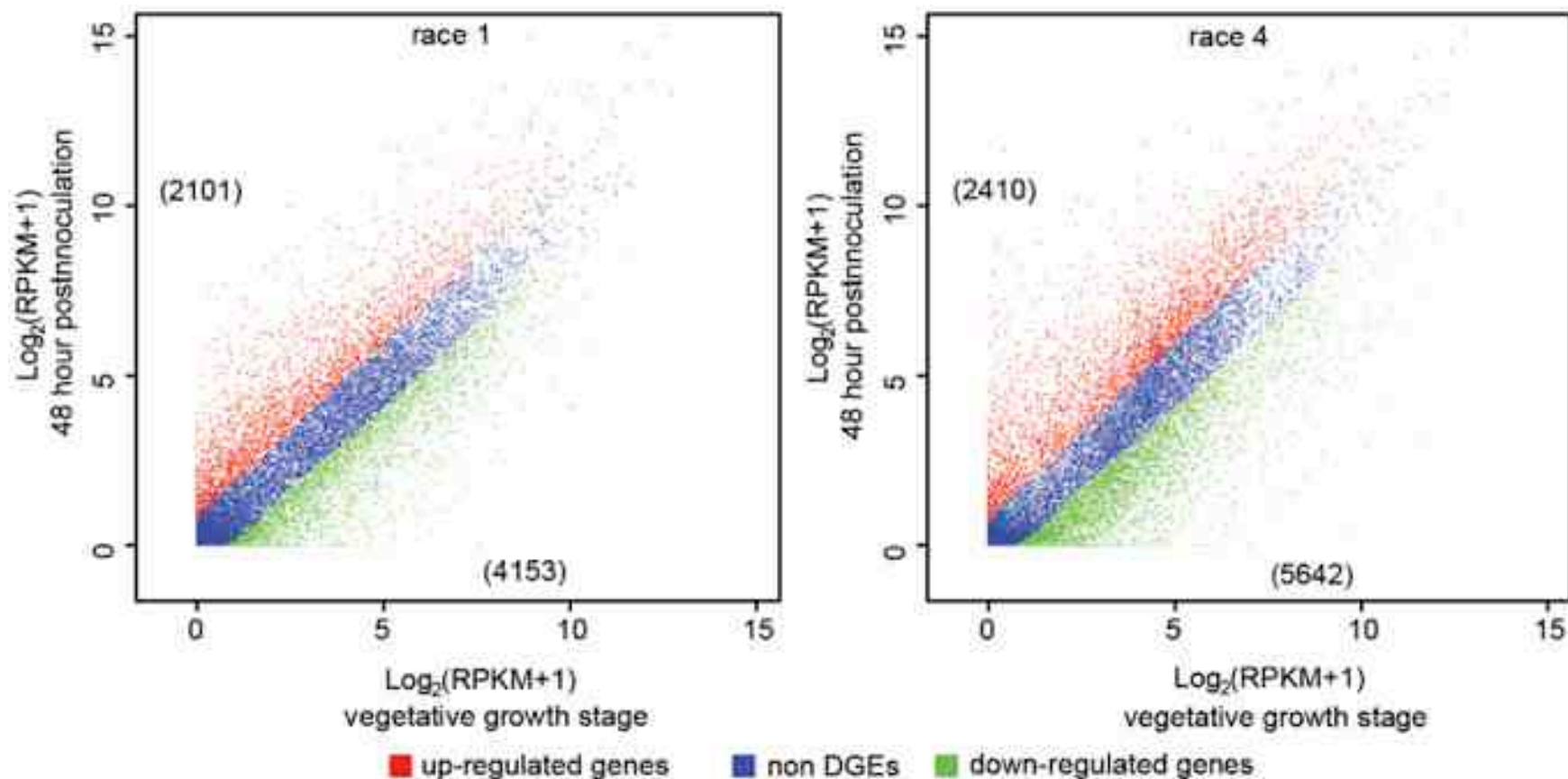
| <i>SIX</i> genes | <i>Foc1</i> gene ID | RPKM_0h | RPKM_48h | <i>Foc4</i> gene ID | RPKM_0h | RPKM_48h |
|------------------|---------------------|---------|----------|---------------------|---------|----------|
| <i>SIX1a</i> | Foc1g01632 | 5.75 | 39.16 | Foc4g00240 | 6.27 | 12.77 |
| <i>SIX1b</i> | NA | - | - | Foc4g00324 | 0.49 | 0 |
| <i>SIX1c</i> | NA | - | - | Foc4g00575 | 50.71 | 113.08 |
| <i>SIX2</i> | NA | - | - | Foc4g07631 | NA | NA |
| <i>SIX6</i> | Foc1g00211 | 14.32 | 13.31 | Foc4g00351 | 243.71 | 192.59 |
| <i>SIX8</i> | NA | - | - | Foc4g00520 | 0.16 | 0.63 |



Genome and Transcriptome Analysis of the Fungal Pathogen *Fusarium oxysporum* f. sp. *ubense* Causing Banana Vascular Wilt Disease

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Functional analysis in Foc



Agrobacterium-mediated insertional mutagenesis in Foc

(sporulation, pathogenicity, chlamyospore formation)

Flávia da Silva Fernandes



R1 sporulation



R2 sporulation



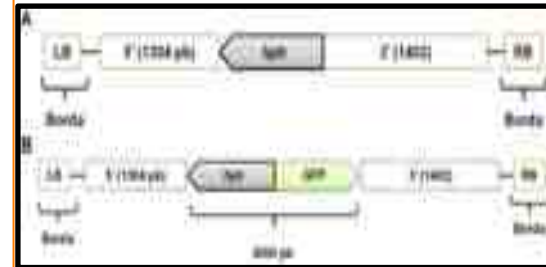
Work in progress, Race 1 and 2 strains in Brazil



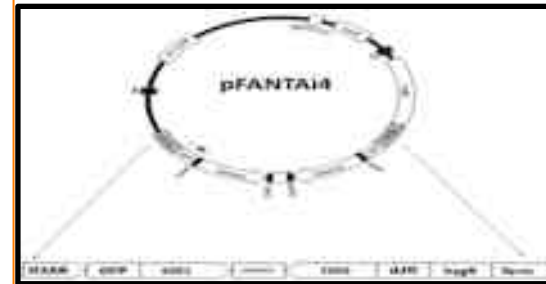
Targeted-gene approach

Joelma dos Santos Fernandes

Knockout



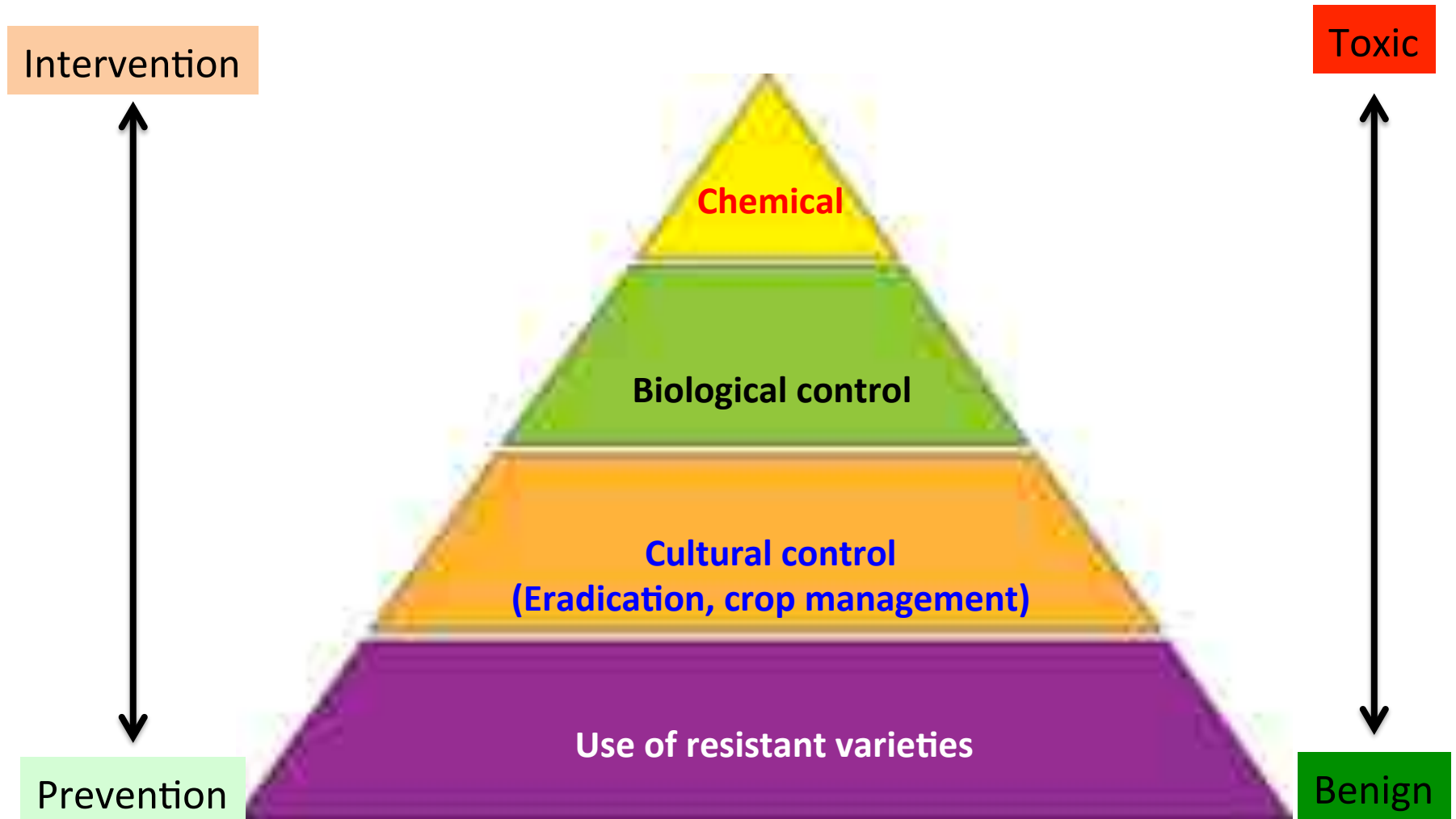
knockdown



Fow2,
SNF1,
Six1, --14
Sge1,
Frp1,
NEP,
MAPK,
PLS1.

RNAi.
I-Mutation

Management options



Resistant varieties vs. Foc TR4?



How many varieties are affected by Foc TR4?

Philippines

Indonesia

1. Raja (AAB)
2. Raja serai (AAB),
3. Buai (AAA),
4. P.Panjang (AAB),
5. Barangan (AAA)
6. Ambon kuning (AAA),
7. Ambon hijau (AAA)
8. Rejang (Aaw),
9. Jantan (AAB),
10. Kepok (ABB),
11. Mas kirana (AA)

Riska & Hermanto (2012)

| Variety | 1 st Crop | 2nd |
|---------------------|----------------------|-----|
| 1.Lakatan (AAA), | 100 | NA |
| 2.Latundan (AAA), | 41 | 100 |
| 3.Gran Nanine(AAA), | 97 | NA |
| 4.Williams (AAA) | 94 | NA |
| 5.GCTCV 119 (AAA) | 1 | 28 |
| 6.GCTCV 218 (AAA) | 1 | 24 |
| 7.FHIA 21 (AABB) | 81 | NA |

Molina (2011)

Australia

~ 26 susceptible Varieties (Walduck & Daly, 2007)

http://www.nt.gov.au/d/Primary_Industry/Content/File/horticulture/TAR2006_07/Banana_panama_pg11_15.pdf

~ 80% of bananas produced worldwide are from varieties susceptible to TR4

Banana cultivars reported with resistance to Foc TR4

| Variety | Resistance | Market | Sources |
|-------------------------------------|------------|--------|---|
| FHIA 01 | R | Seeded | Walduck & Daly 2007 Bingzhi et al. 2004 |
| FHIA 02 | R | Seeded | Bingzhi et al. 2004 |
| FHIA 18 | R | Good | Walduck & Daly 2007 Bingzhi et al. 2004 |
| FHIA 25 | R | Poor | Walduck & Daly 2007 Bingzhi et al. 2004 |
| Pisan Jai Buya | R | NA | Walduck & Daly 2007 Bingzhi et al. 2004 |
| *GCTV 119 | MR | Good | Walduck & Daly 2007 Bingzhi et al. 2004 Molina 2011 |
| GCTV 218 | MR | Good | Molina, 2011 |
| <i>M. accuminata malaccensis</i> ** | R | Seeded | Walduck & Daly 2007 |
| ABB varieties (Saba, Tanduk) | R | NA | Molina - unpublished |



* There are many GCTV, produced by TBRI, Taiwan, which are continuously replaced for new somaclons with some resistance levels

**Some *M.a. malaccensis* are susceptibles



At least 50 varieties are affected by TR4

~ 80 % of banana produced are from susceptible varieties [Ploetz et al. 2005]

In the Caribbean ~ 63% of 2,856,394 t produced in 2009 were from TR4-susceptible varieties

(Data: Lescot, 2010)

Cavendish Banana Cultivars Resistant to Fusarium Wilt Acquired through Somaclonal Variation in Taiwan



Shin-Chuan Hwang

Dr. Hwang is a plant pathologist and Director of the Taiwan Banana Research Institute. He received his B.S. degree from National Taiwan University and his M.S. and Ph.D. degrees from the University of Hawaii. His research interests cover all aspects of banana diseases, with special emphasis on Fusarium wilt. He was elected as one of the Ten Outstanding Young Men of the nation in 1982, received the Distinguished Agricultural Research and Extension Award from the Council of Agriculture of Taiwan in 1988, and was bestowed the Outstanding Applied Science and Technology Award by the National Science



Wen-Hsiung Ko

Dr. Ko is professor of plant pathology at Beaumont Agricultural Research Center of the University of Hawaii at Manoa. He received his B.S. degree from National Taiwan University and his Ph.D. degree from Michigan State University. His research interests include ecology and biological control of soilborne diseases of tropical fruit crops and biology of *Phytophthora*. He received the Ruth Allen Award in 1988 from the American Phytopathological Society for his contribution in hormonal regulation of sexual reproduction in *Phytophthora* and is a Fellow of the American Phytopathological Society.

La resistencia de los somaclones es dependiente de la presión de inóculo!

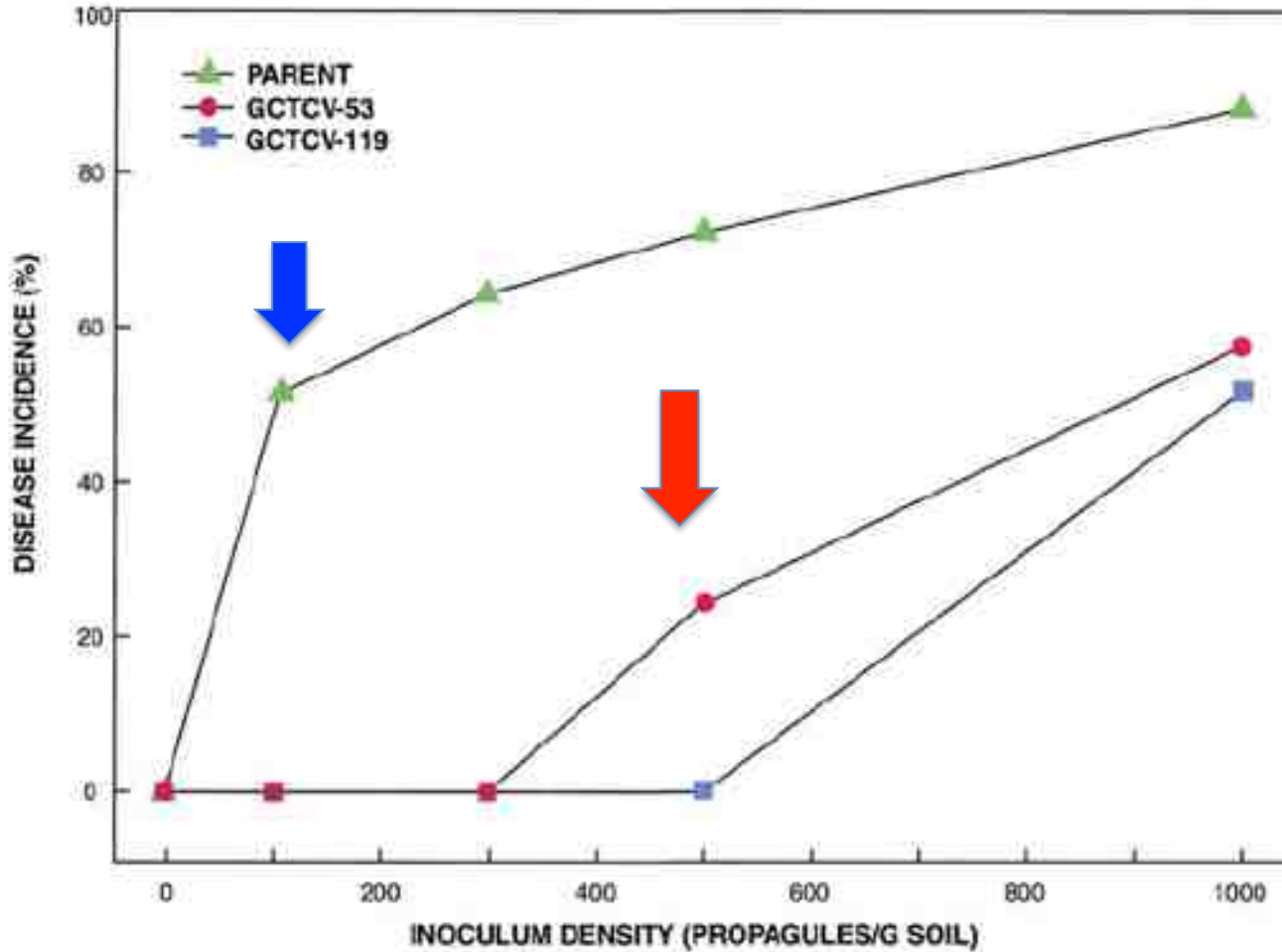


Fig. 5. Disease incidence of resistant somaclone in comparison with the susceptible Giant Cavendish parent in trays with various inoculum densities of race 4 of *Fusarium oxysporum* f. sp. *cubense*.

There are phenotypic variations environmentally-associated

Hwang & Ko 2004, Plant Disease / June 2004

Table 3. Horticultural characteristics of Fusarium wilt resistant clones derived from Giant Cavendish by somaclonal variation

| Clone | Resistance ^a | Horticultural characteristics |
|-----------|-------------------------|--|
| GCTCV-40 | High | Tall and slender pseudostem; weak petiole with narrow and drooping leaves; small bunch |
| GCTCV-44 | High | Short and slender pseudostem; weak petiole and drooping leaves; bunch normal but weak pedicel |
| GCTCV-46 | High | Black spots on pseudostem and leaf sheath; upright leaves; small bunch with short fingers |
| GCTCV-53 | High | Dark green pseudostem; drooping leaves; elongate male bud; small bunch with short fingers |
| GCTCV-62 | Moderate | Pale green pseudostem; fewer suckers; small bunch and fingers |
| GCTCV-104 | High | Pale green pseudostem; fewer fingers; long growth cycle |
| GCTCV-105 | High | Shorter and slender pseudostem; compact bunch with a greater number of short fingers |
| GCTCV-119 | High | Very tall; wavy leaves; short fruit stalk; long growth cycle; fewer hands but large fingers; sweeter fruit |
| GCTCV-201 | Moderate | Robust pseudostem; short fruit stalk; malformed hands |
| GCTCV-215 | High | Tall and slender pseudostem; leaf tip curl and splitting; fewer suckers; normal bunch but slender fingers; long growth cycle |
| GCTCV-216 | High | Very tall; very large and heavy bunch; long growth cycle |
| GCTCV-217 | High | Erect leaves; compact, but heavy bunch; less curved fingers |

^a High is <10% average disease incidence; moderate is 11 to 30% average disease incidence. Disease incidences were determined 1 year after planting.

Cavendish somaclons with resistance levels to Foc TR4, produced by TBRI, promoted by Bioversity-Philippines

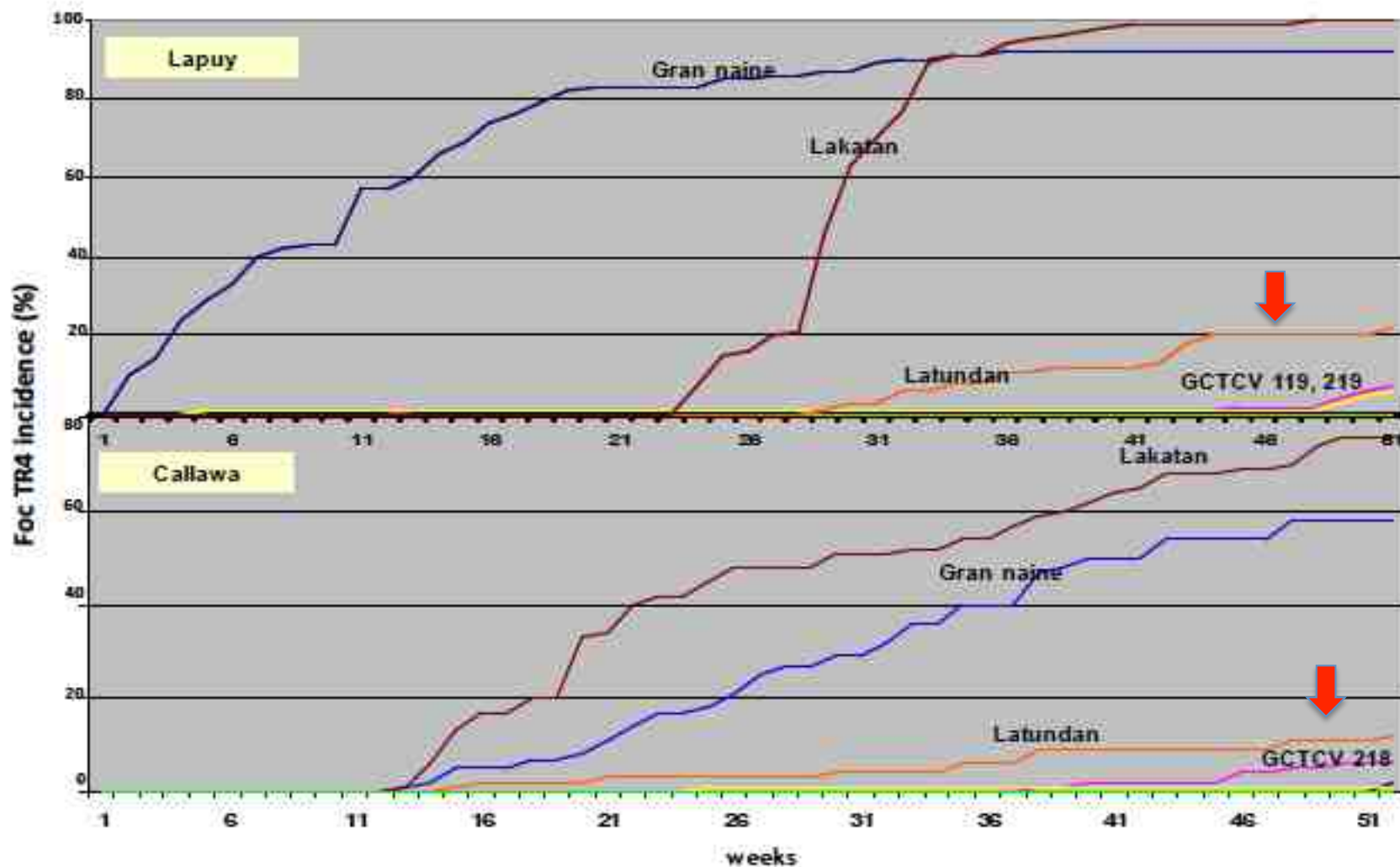


Figure 3. Fusarium incidence of different banana cultivars planted in Lapuy and Callawa, Davao City in 2009 - 2012)

Cavendish somaclons with resistance levels to Foc TR4, produced by TBRI, promoted by Bioversity-Philippines



© Miguel Dita

“No Cavendish types were identified as resistant, although GCTV119 from Taiwan and CJ19 from Indonesia show better survival than other Cavendish types; **however, still not enough to be commercially viable** (Walduck & Daly 2007)

http://www.nt.gov.au/d/Primary_Industry/Content/File/horticulture/TAR2006_07/Banana_panama_pg11_15.pdf

Fusarium wilt-resistant lines of Brazil banana (*Musa spp.*, AAA) obtained by EMS-induced mutation in a micro-cross-section cultural system

Y. F. Chen, W. Chen, X. Huang, X. Hu, J. T. Zhao, Q. Gong, X. J. Li and X. L. Huang*

The Key Laboratory of Gene Engineering of the Ministry of Education, School of Life Sciences, Zhongshan (Sun Yat-sen) University, Guangzhou 510275, China



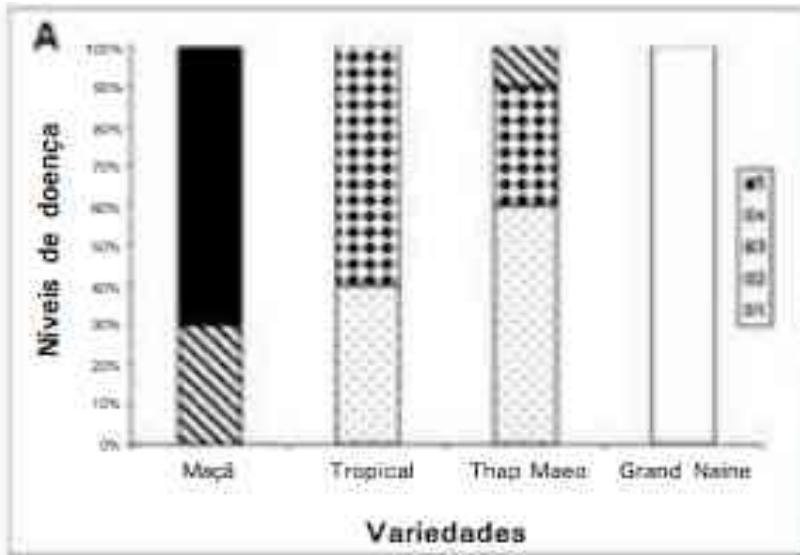
5 of 6 líneas resistentes en invernadero fueron resistentes en campo

Phenotyping for TR4 resistance- high-throughput system

Fast but reliable!



Screening for Fusarium wilt resistance



Comunicado 150
Técnico

ISSN 1809-502X
Cruz das Almas, BA
Dezembro, 2011

Metodologia para a caracterização de genótipos de bananeira quanto à resistência ao mal-do-Panamá em casa-de-vegetação



Miguel Angel Dita Rodriguez¹

Lindineia Ribeiro²

Edson Perito Amorim¹

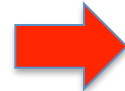
Zilton José Maciel Cordeiro¹

Sebastião de Oliveira e Silva²

Phenotyping for TR4 resistance- high-throughput system

Genotypes vs. Foc TR4

1. Grande Naine (AAA)
2. Gros Michel (AAA)
3. Maçã (Silk) – AAB
4. Prata (AAB)
5. *CIRAD 930 (AA) -*M. a. malaccensis*
6. *Pahang (AA) - *M. a. malaccensis*
7. Matavia (Bluggoe) – ABB
8. Banksii (AA)
9. Tuu Gia (AA)



* Parcial resistance

Evaluation of East African Highland Banana (AAA) and Plantain (AAB) genotypes in Asia for Foc TR4 resistance

| ITC code | Cultivar | Genome | Subgroup |
|----------|-------------------------------------|---------|------------------------|
| ITC0081 | Igitsiri (Intuntu) | AAA | EAHB |
| ITC0179 | Inkira | AAA | EAHB |
| ITC0084 | Mbwazirume | AAA | EAHB |
| ITC0166 | Ingagara | AAA | EAHB |
| ITC1354 | Enzirabahima | AAA | EAHB |
| ITC1355 | Kazirakwe | AAA | EAHB |
| ITC1465 | Ibwi | AAA | EAHB |
| ITC0215 | Mbi Egome 1 | AAB | Plantain - French |
| ITC0217 | Akpakpak | AAB | Plantain - French |
| ITC0519 | Obubit Ntanga green mutant | AAB | Plantain - French |
| ITC0121 | Ihitisim | AAB | Plantain - Horne |
| ITC0208 | Atali Kiogo | AAB | Plantain - False Horne |
| ITC1165 | Curare | AAB | Plantain - False Horne |
| ITC1325 | Orishele | AAB | Plantain - False Horne |
| ITC0570 | Williams (Bell, South Johnstone) | AAA | Cavendish |
| | GCTCV-119 | AAA | Cavendish SC variant |
| | Grand Nain | AAA | Cavendish |
| | Lakatan | AAA | (dessert type) |
| | Latundan | AAB | Silk |
| | Cardaba | ABB/BBB | Saba |
| | Baxi | AAA | Cavendish |
| | Guangfen No.1 | ABB | Pisang Awak |

Banana Varieties resistant to Race 1 and 2 Embrapa's breeding program



Caprichosa
Type: Prata

YS : RESISTANT
BLS: RESISTANT
FW: RESISTANT



Garantida
Type : Prata

YS : RESISTANT
BLS: RESISTANT
FW: RESISTANT



Tropical
Type- Silk

YS : RESISTANT
BLS: SUSCEPTIBLE
FW: RESISTANT



Princesa
Type : Silk

YS : RESISTANT
BLS: SUSCEPTIBLE
FW: RESISTANT

We don't know yet their reaction for TR4

Banana Varieties resistant to Race 1 and 2 Embrapa's breeding program

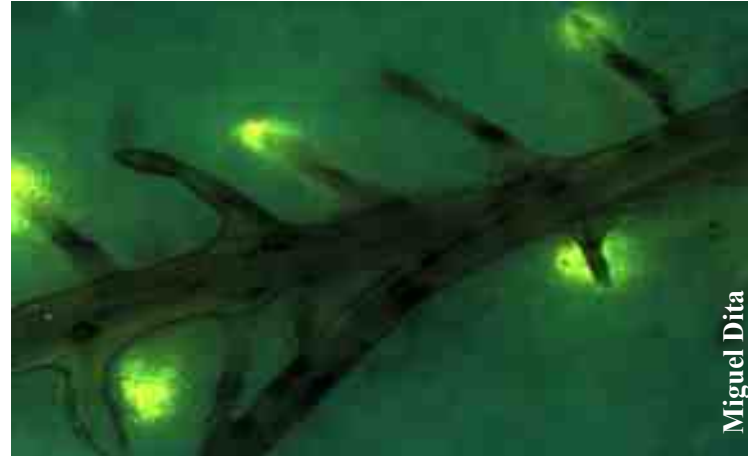
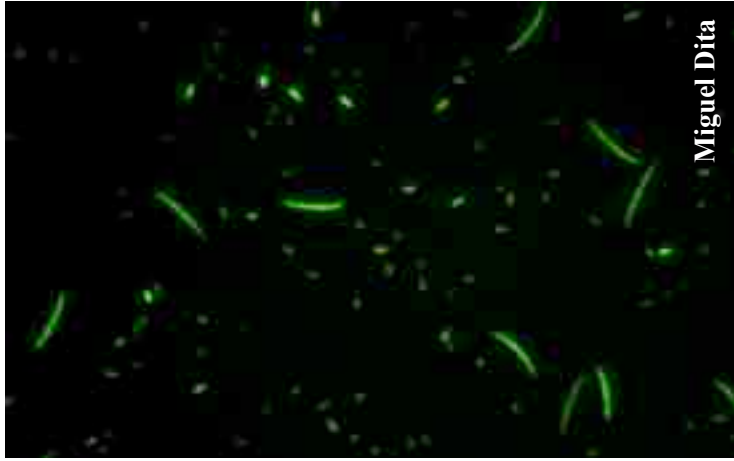


We don't know yet the reaction for TR4

More information with Dr. E. Amorim
edson.amorim@embrapa.br

Understanding genetic and molecular basis of resistance

Comparative transcriptome analyses of compatible and incompatible Musa- FOC interactions



X

Dita et al. 2010

Immune



Partial resistant



Susceptible



454- seq



EST data

Some old questions:

- When and where is Foc stopped in Cavendish during Race 1 infection?
- Which gene(s) are involved? At what level?
- Which mechanisms are responsible for resistance (effective >60 years)

Banana genome vs. Foc genome: 1st Insights

Banana genome

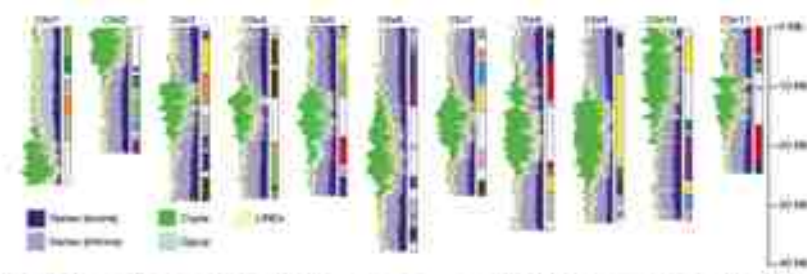
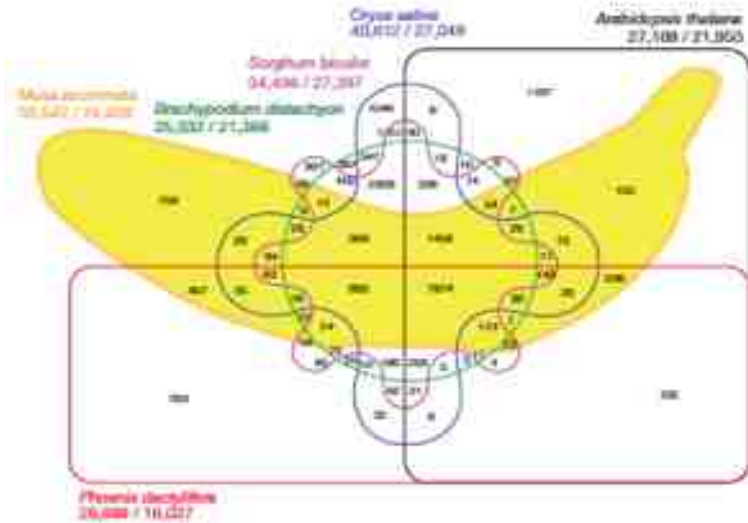


Figure 1. Chromosomal distribution of the banana genome. The figure shows the distribution of the banana genome across 11 chromosomes, with different colors representing different cellular localizations. The legend at the bottom identifies the localizations: cytoskeleton (blue), cytosol/mitochondria (green), endoplasmic reticulum (yellow), extracellular (red), golgi apparatus (purple), mitochondria (orange), mitochondria/nucleus (brown), nucleus (pink), peroxisome (grey), and plasma membrane (light blue).

D'Hont et al. Nature 488, 2012)

Foc TR4 In vitro

Berg et al. 2012
Li-Jun Ma Team

| BLAST Match Characteristics | Number of Proteins | Number of Predicted Secreted Proteins | Number of Predicted Secreted Proteins < 200 AA |
|--|--------------------|---------------------------------------|--|
| Conserved Lineage Specific (to Fo4287) | 15238 | 1526 | 294 |
| Unique to TR4 | 140 | 7 | 2 |
| | 451 | 40 | 12 |

Foc TR4 and R1 in planta

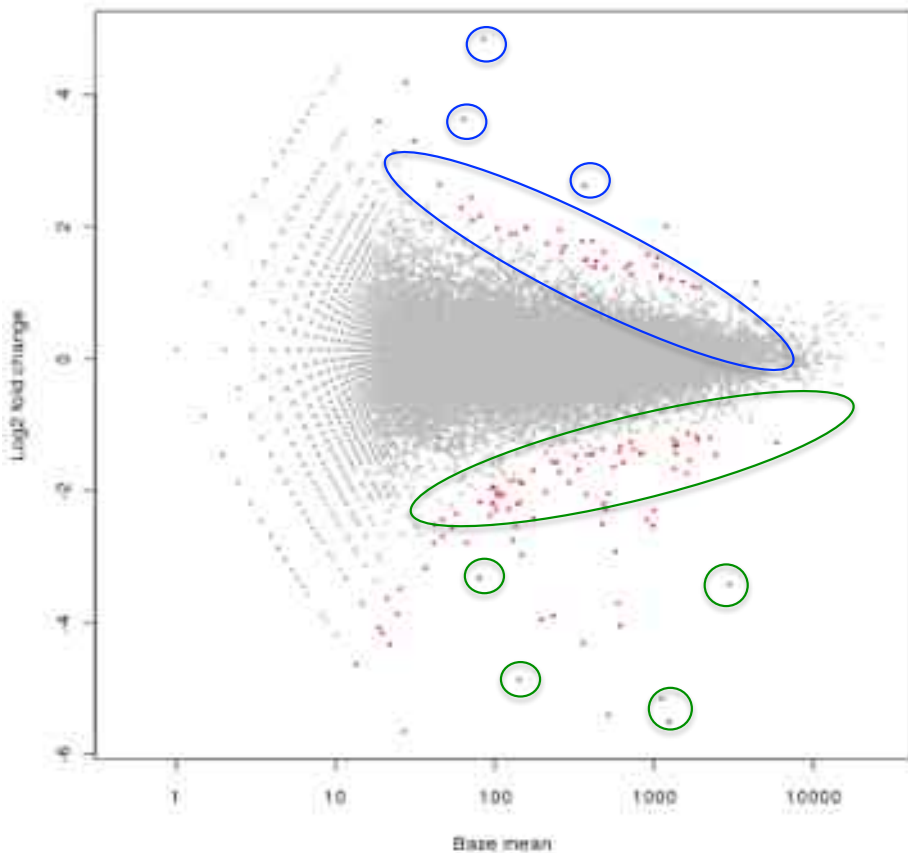
| Cellular localization | Gran Naine- TR4 | Pahang-TR4 |
|-----------------------|-----------------|------------|
| cytoskeleton | 198 | 54 |
| cytosol/mitochondria | 26 | 10 |
| endoplasmic reticulum | 6 | 0 |
| extracellular | 858 | 177 |
| golgi apparatus | 3 | 1 |
| mitochondria | 1332 | 487 |
| mitochondria/nucleus | 9 | 2 |
| nucleus | 2747 | 672 |
| peroxisome | 24 | 2 |
| plasma membrane | 1223 | 264 |
| Total | 8397 | 2290 |

Aligned against *Foc (draft)* -software TopHat [Dita un published]

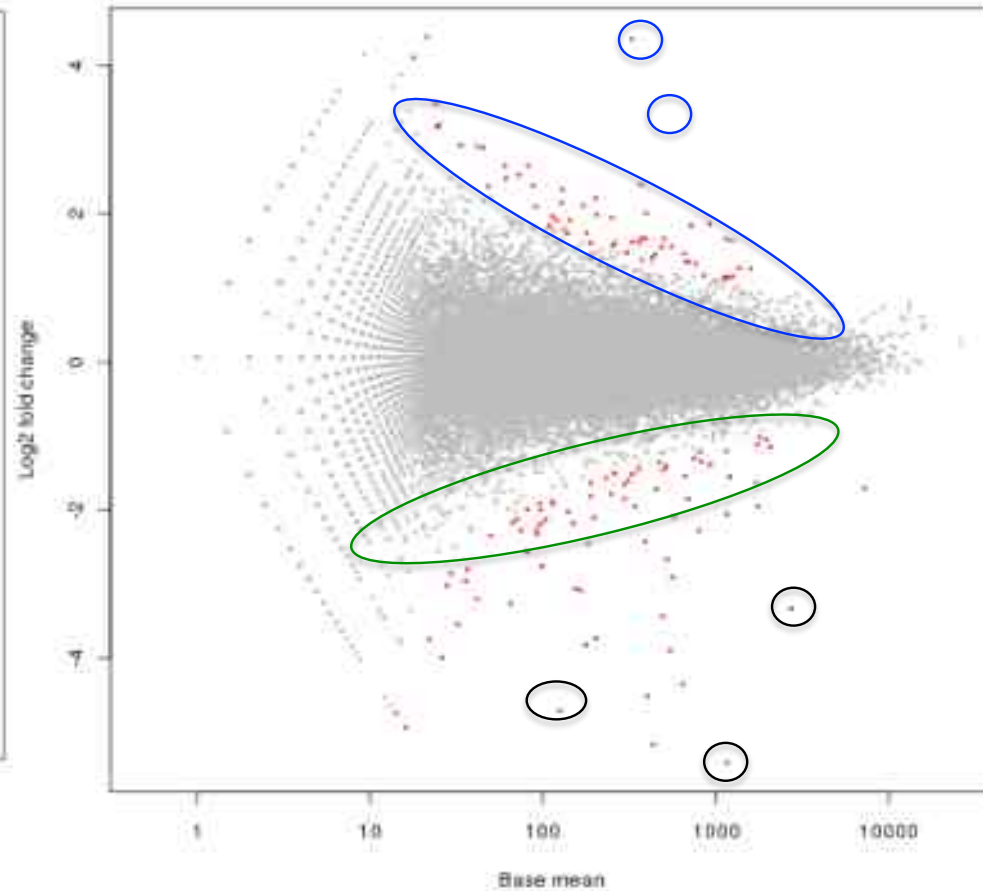
Comparative transcriptome analyses of compatible and incompatible Musa- FOC interactions

Musa Genes

G. Naine **Control** vs. **G. Naine Foc R1**



Gran Naine **Control** vs. **Gran Naine Foc TR4**

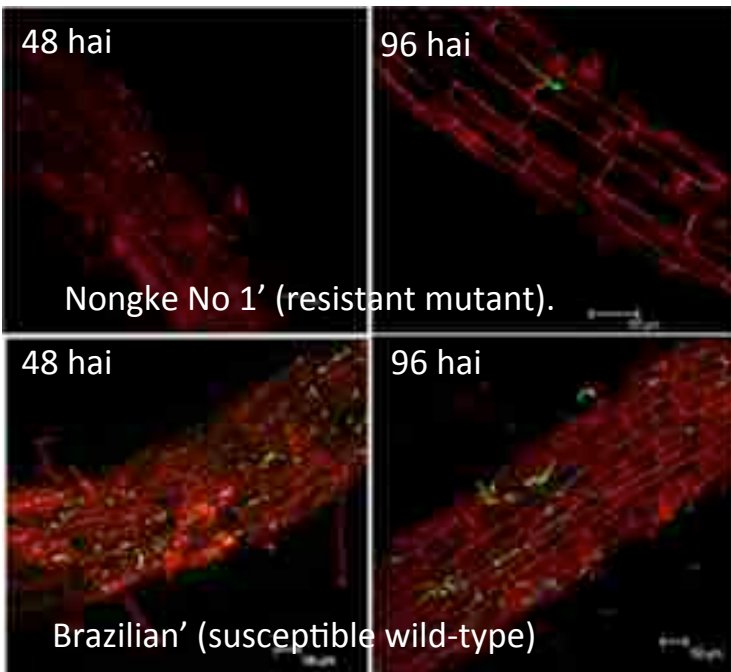


RESEARCH ARTICLE

Open Access

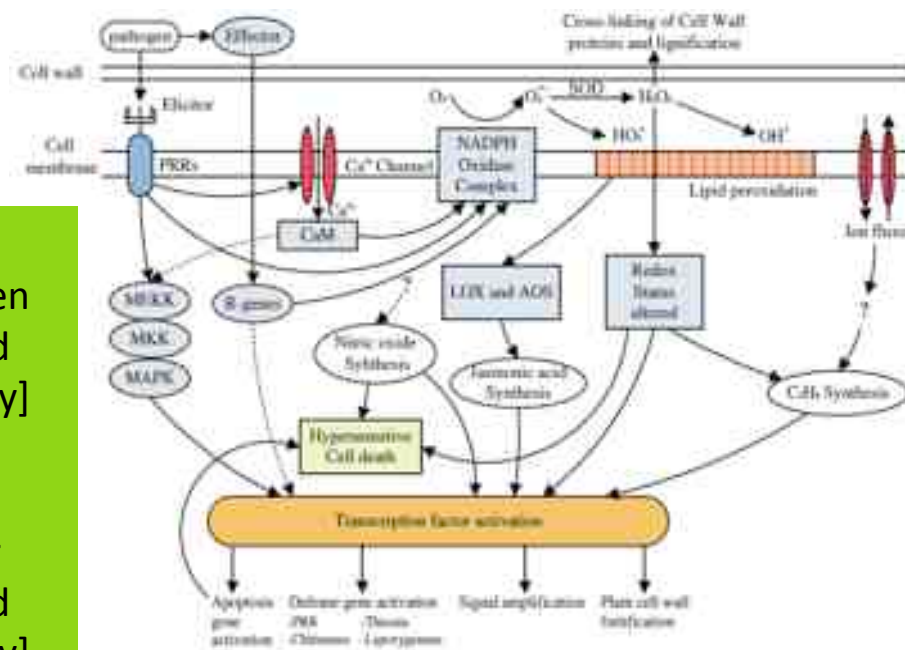
Transcriptome profiling of resistant and susceptible Cavendish banana roots following inoculation with *Fusarium oxysporum* f. sp. *cabense* tropical race 4

Chun-yu Li^{1,2†}, Gui-ming Deng^{3†}, Jing Yang^{3†}, Altus Viljoen⁴, Yan Jin^{1,2}, Rui-bin Kuang^{1,2}, Cun-wu Zuo³, Zhi-cheng Lv³, Qiao-song Yang^{1,2}, Ou Sheng^{1,2}, Yue-rong Wei^{1,2}, Chun-hua Hu^{1,2}, Tao Dong^{1,2} and Gan-jun Yi^{1,2*}



PTI
[pathogen triggered immunity]

ETI
[Effector triggered immunity]



Comparative proteomic analyses: Musa- FOC interactions

Li et al. *Proteome Science* 2013, **11**:41
<http://www.proteomesci.com/content/11/1/41>



RESEARCH

Open Access

Proteomic analysis of *Fusarium oxysporum* f. sp. *cubense* tropical race 4-inoculated response to *Fusarium* wilts in the banana root cells

Xingshen Li^{1,2}, Tingting Bai^{1,2}, Yunfeng Li², Xiaofei Ruan^{1,2} and Huaping Li^{1,2*}



Figure 1 Disease symptoms from *Musa* roots with treatment of *F. oxysporum*. **A** Banan roots (H₂O control), and **B** Banan roots (Fusarium). (susceptible fruit, moderately resistant Nongke No.1 and highly resistant Yuyoukang) were treated at 30 d after *F. oxysporum* inoculation, compared with water-treated control.

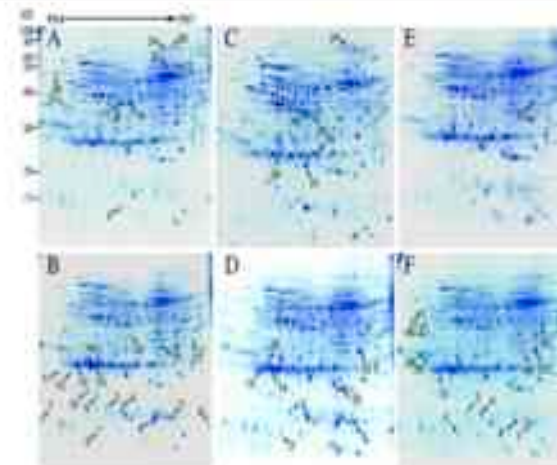


Figure 2 Representative 2-DE of protein from *Musa* roots with treatment of *F. oxysporum*. **A**, Brazil; **D** control; **B**, Brazil (Fusarium); **C**, Nongke No.1; **H₂O** control; **E**, Yuyoukang; **F** (H₂O control); **F**, Yuyoukang (Fusarium). Proteins (200 µg) were loaded on an 18 cm 8% and pH 4-7 to 18. Following electrophoresis of 12% SDS-PAGE and DBI 2-DE staining. The different expression spots present at the position of arrows are compared with their respective controls.

Comparative proteomic analyses: Musa- FOC interactions

Li et al. *Proteome Science* 2013, **11**:41
<http://www.proteomesci.com/content/11/1/41>



RESEARCH

Open Access

Proteomic analysis of *Fusarium oxysporum* f. sp. *cubense* tropical race 4-inoculated response to *Fusarium* wilts in the banana root cells

Xingshen Li^{1,2}, Tingting Bai^{1,2}, Yunfeng Li², Xiaolei Ruan^{1,2} and Huaping Li^{1,2*}

Conclusions:

This report is the first to use proteomic profiling to study the molecular mechanism of banana roots infected with Foc4.

The differentially regulated proteins involved in different defense pathways are likely associated with different resistant levels of the three banana cultivars.

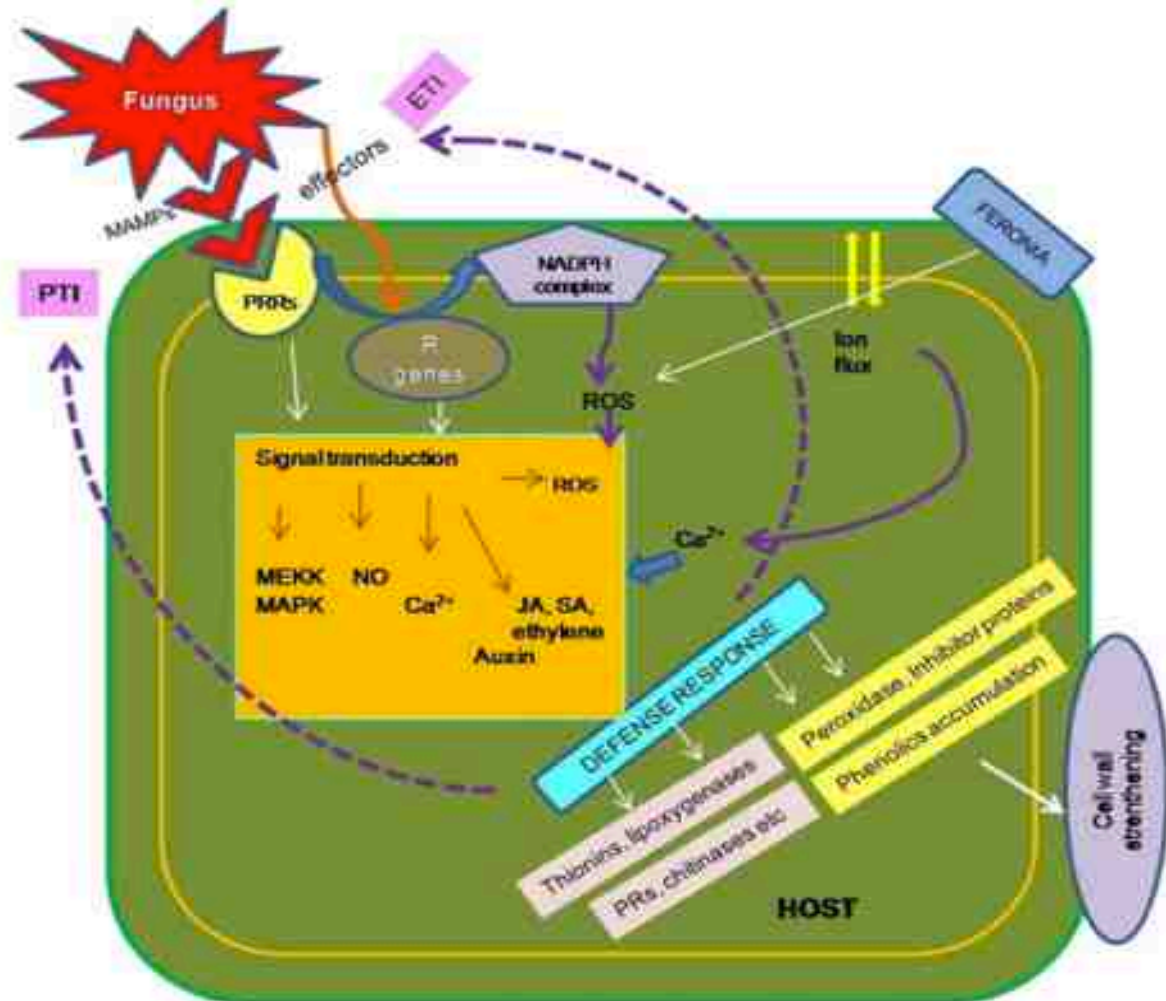
Plant defense response against *Fusarium oxysporum* and strategies to develop tolerant genotypes in banana

V. Swarupa · K. V. Ravishankar · A. Rekha

738

Planta (2014) 239:735–751

Fig. 1 Recognition of the pathogen and elicitation of defense response by the host. When the pathogen attacks the host through their typical molecular signatures called microbe-associated molecular patterns (MAMPs) and/or effectors, they are recognized by the host through specific pathogen recognition receptors (PRRs) and R-genes leading to the activation of multiple signal transduction pathways such as salicylic acid (SA), jasmonic acid (JA) and ethylene. Pathogen recognition results in change in membrane potential leading to activation of NADPH complex that results in production of reactive oxygen species (ROS), which acts as major signaling molecules and in turn, may induce other signaling molecules. Change in ion flux also induces Ca^{2+} signaling. Defense responses against MAMPs are called pathogen-triggered immunity (PTI) and against effectors are called effector-triggered immunity (ETI)



Is Transgenic an alternative ? What do we know so far?

Journal of Integrative Plant Biology

Formerly *Acta Botanica Sinica*

2005, 47 (8): 971–977

<http://www.blackwell-synergy.com>

<http://www.chineseplantscience.com>

Creation of Transgenic Bananas Expressing Human Lysozyme Gene for Panama Wilt Resistance

Xin-Wu PEI¹, Shi-Kai CHEN², Rui-Ming WEN², Shang YE², Jia-Qin HUANG², Yong-Qiang ZHANG¹,
Bing-Shan WANG¹, Zhi-Xing WANG¹ and Shi-Rong JIA^{1*}

(1. *Biotechnology Institute, Chinese Academy of Agricultural Sciences, Beijing 100081, China;*

2. *Xinyi Institute of Agricultural Science and Technology, Xinyi, Guangdong 523530, China)*

Plant Biotechnology
Journal



Plant Biotechnology Journal (2011) 9, pp. 1141–1148

doi: 10.1111/j.1467-7652.2011.00639.x

Apoptosis-related genes confer resistance to Fusarium wilt in transgenic 'Lady Finger' bananas

Jean-Yves Paul¹, Douglas K. Becker¹, Martin B. Dickman², Robert M. Harding¹, Harjeet K. Khanna¹ and James L. Dale^{1,*}

¹*Centre for Tropical Crops and Biocommodities, Faculty of Science and Technology, Queensland University of Technology, Brisbane, Qld, Australia*

²*Institute for Plant Genomics and Biotechnology, Department of Plant Pathology and Microbiology, Texas A&M University, College Station, TX, USA*

Is Transgenic an alternative ? What do we know so far?

Scientia Horticulturae (2013) 393–399

Contents lists available at ScienceDirect

Scientia Horticulturae

journal homepage: www.elsevier.com/locate/scihorti

Transgenic banana cv. Rasthali (AAB, Silk gp) harboring *Ace-AMP1* gene imparts enhanced resistance to *Fusarium oxysporum* f.sp. *cubense* race 1

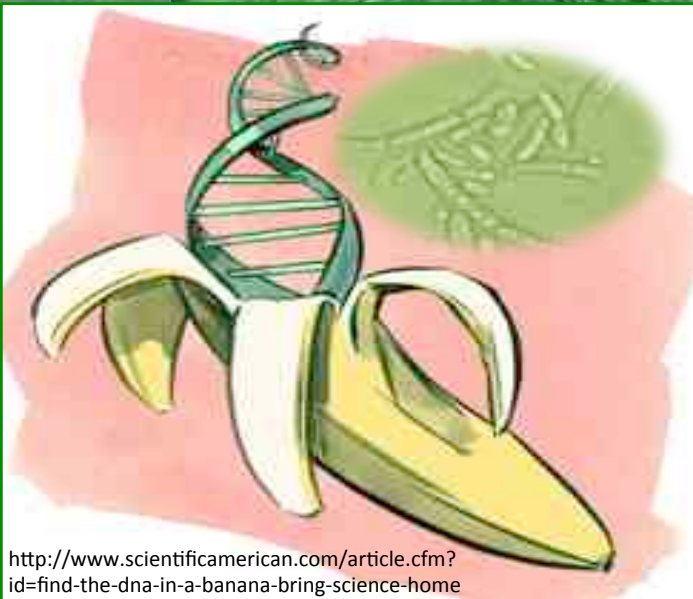
Sukhada Mohandas^{a,*}, H.D. Sowmya^a, A.K. Saxena^a, S. Meenakshi^a, R. Thilaka Rani^a, Riaz Mahmood^b

^a Indian Institute of Horticultural Research, Hebbal, Bangalore 560 089, India
^b Department of Studies and Research in Biotechnology & Biopharmatics, Ravenshaw University, Bhubaneswar 751 031, India



Conclusion: Significantly higher AMP-protein content in transgenic lines was negatively correlated and higher root colonization was positively correlated with disease symptoms. Six transgenic lines were selected for field evaluation.

Fig. 6. Progress of disease symptoms caused by *Fusarium oxysporum* f.sp. *cubense* race 1 after root challenging of transformed banana plants with untransformed control. (a) External symptoms of transformed banana plants (plants – 6, T3, 14, 18 and 19) and control plants fully devastated on the left. (b) Vascular discoloration of transgenic plants ((c–e) Plants 19, 14 and 6) along with control treated with Foc.



<http://www.scientificamerican.com/article.cfm?id=find-the-dna-in-a-banana-bring-science-home>

There are no resistant varieties, so what ?



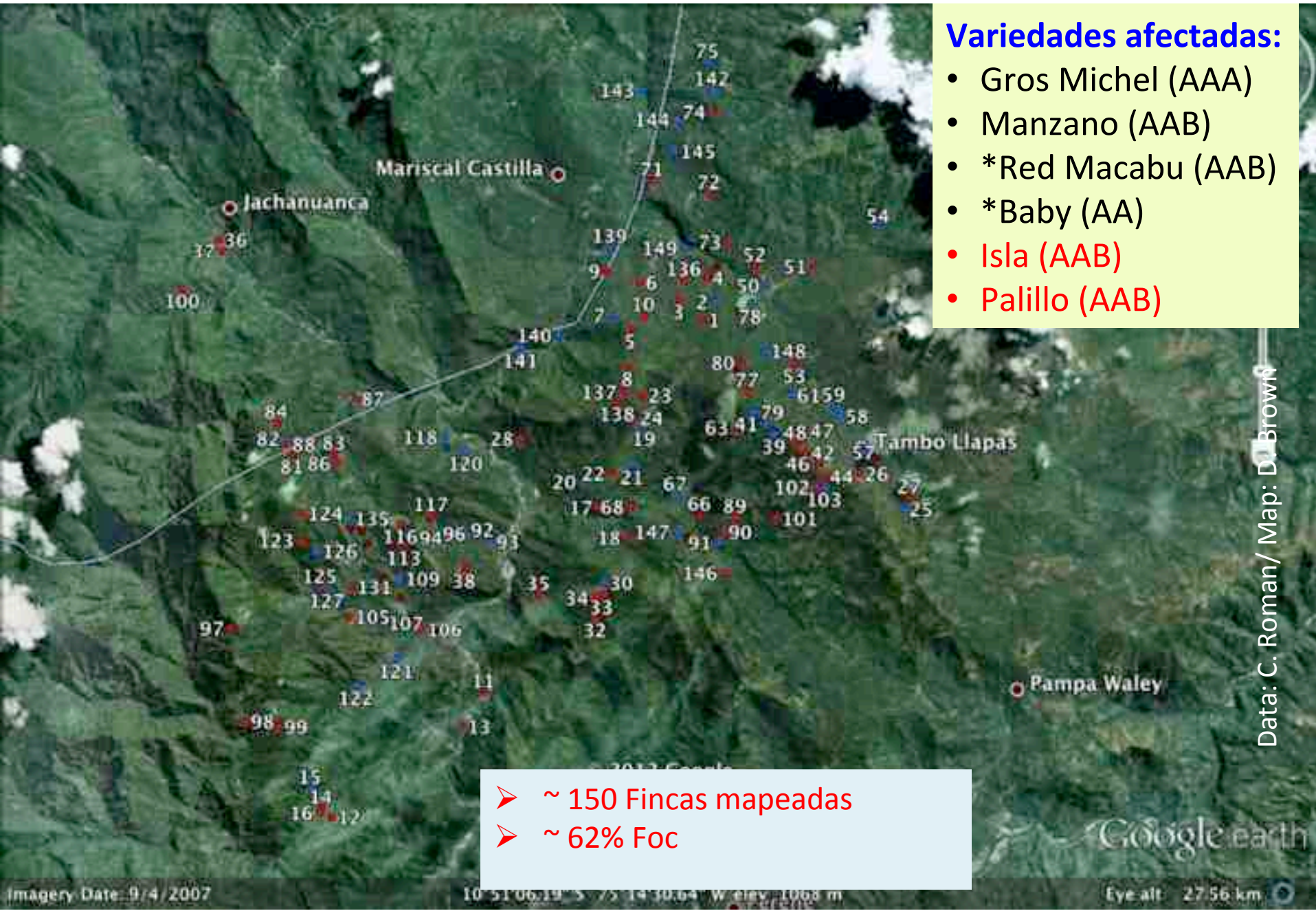
Mapping and epidemiological studies of Foc in Peru



Mapping and epidemiological studies of Foc in Peru

Variedades afectadas:

- Gros Michel (AAA)
- Manzano (AAB)
- *Red Macabu (AAB)
- *Baby (AA)
- Isla (AAB)
- Palillo (AAB)

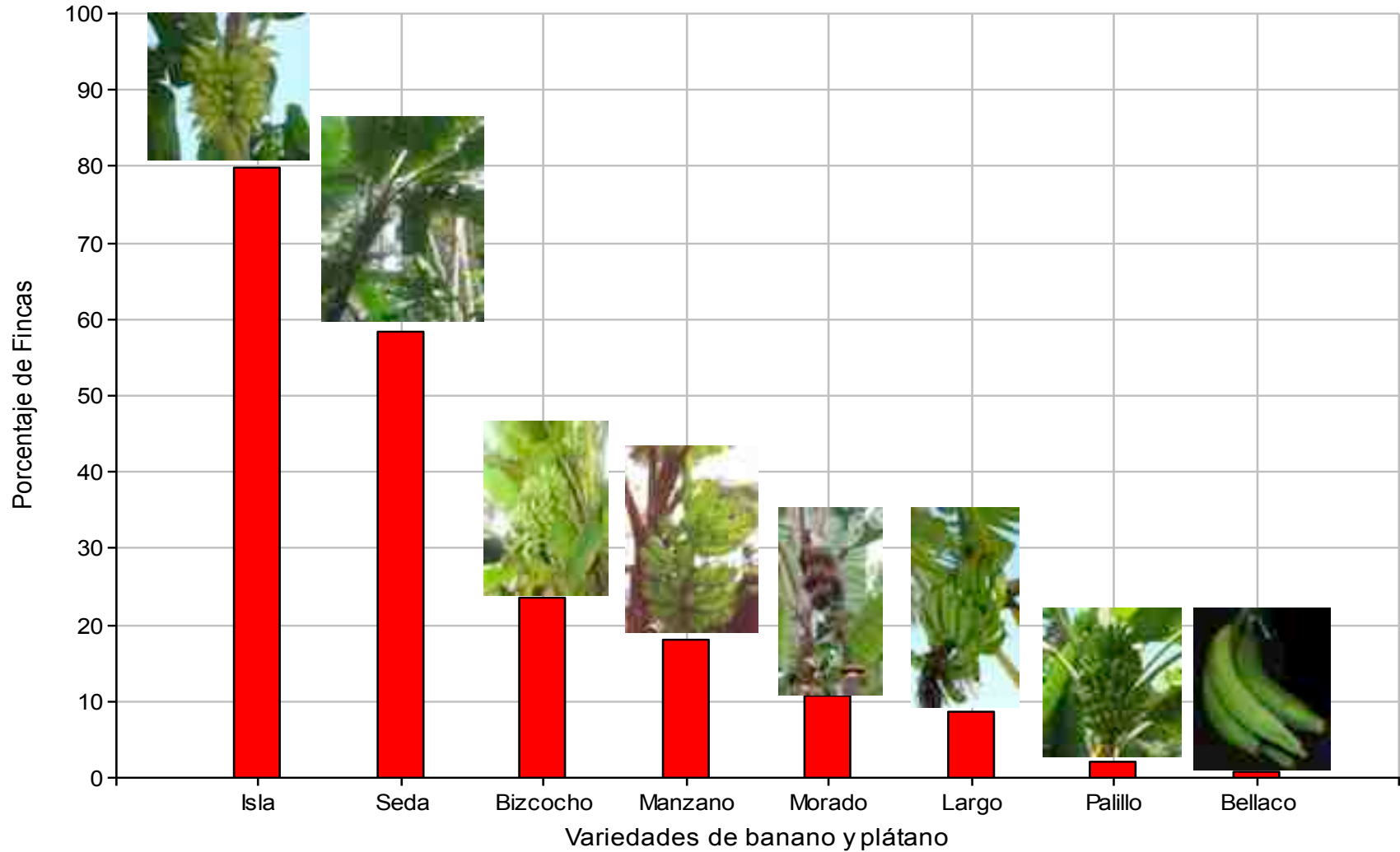


- ~ 150 Fincas mapeadas
- ~ 62% Foc

Data: C. Roman/ Map: D. Brown

Google Earth

Banana varieties in San Luis de Shuaro District, Peru



Use of bioinputs



Fences



Destronque



Prunning



Long spacement between plants

Elements of managemnt present on farm with absence or lower levels of Fusarium wilt incidence



CaOH



Coffe debris



Capacitation



Eliminate disese plants
Apply wood ash



Cut & apply CaOH

Association of some practices with Fusarium wilt intensity in Peru

(Roman, 2012)

| Activity | Association with FOC |
|---|----------------------|
| Introduction of planting material and soil from other farms | High |
| Leave infected plants in the farm without isolation | High |
| Presence of transit areas across the farm | High |
| Remove roots and soil attached to suckers in the farm | High |
| Discard infected plants in rivers or irrigation channels | High |
| Presence of water flow towards the farm and poor drainage | High |
| Presence of FOC in neighboring farms | Medium |
| Tools are frequently sterilized | Medium |
| Apply measures to avoid contaminated soils in the farm | Medium |
| Selection of clean planting material | Low |
| Diversification of varieties | Low |
| Ca identify the disease correctly | Low |
| Use of natural barriers | Low |
| Use of warning systems on the entry points | Low |
| presence of fences | Low |
| Apply crop management strategies | Low |
| Organization and capacitation of workers | Low |
| Monitoring and surveillance for early detection | Low |
| Eliminate/isolate infected plants immediately | Low |



High
 Medium
 Low

Cultural control

Silicon Suppresses Fusarium Wilt Development in Banana Plants

ALESSANDRO ANTONIO FORTUNATO¹, FABRÍCIO ÁVILA RODRIGUES¹, JÚLIO CEZAR PARPAIOLA BARONI¹, GUSTAVO CÉSAR BARBOSA SOARES¹, MIGUEL ANGEL DITA RODRIGUEZ² and OLINTO LIPARINI PEREIRA¹

Authors' addresses: ¹Department of Plant Pathology, Viçosa Federal University, Zip Code 36570-000, Viçosa, Minas Gerais, Brazil; ²Embrapa Mandioca e Fruticultura, Zip Code 44380-000, Cruz das Almas, Bahia, Brazil (correspondence to F. A. Rodrigues. E-mail: fabricio@ufv.br)

Received April 25, 2012; accepted July 27, 2012

Table 5

Area under darkening of rhizome-pseudostem progress curve (AUDRPPC) and the area under relative lesion length progress curve (AURLLPC) for banana plants from the Grand Nain and Maçã cultivars grown in soil amended with silicon (+Si) or without silicon (-Si) and inoculated with *Fusarium oxysporum* f. sp. *cubense*.

| Treatments | AUDRPPC | | AURLLPC | |
|------------|------------|-----------|------------|-----------|
| | Grand Nain | Maçã | Grand Nain | Maçã |
| -Si | 114.29 Aa | 110.86 Aa | 367.96 Ba | 606.92 Aa |
| +Si | 117.29 Aa | 93.14 Bb | 322.05 Aa | 354.79 Ab |
| C.V. (%) | 8.87 | | 15.13 | |

C.V. = coefficient of variation.

Means within each column followed by the same lowercase letter or means within each row followed by the same uppercase letter are not significantly different ($P = 0.05$) as determined by Tukey's test.



Cultural control

Chinese Leek (*Allium tuberosum*) suppress Foc

Dr. Yi Ganjun in the 7th BAPNET SC Meeting, Hanoi, Vietnam

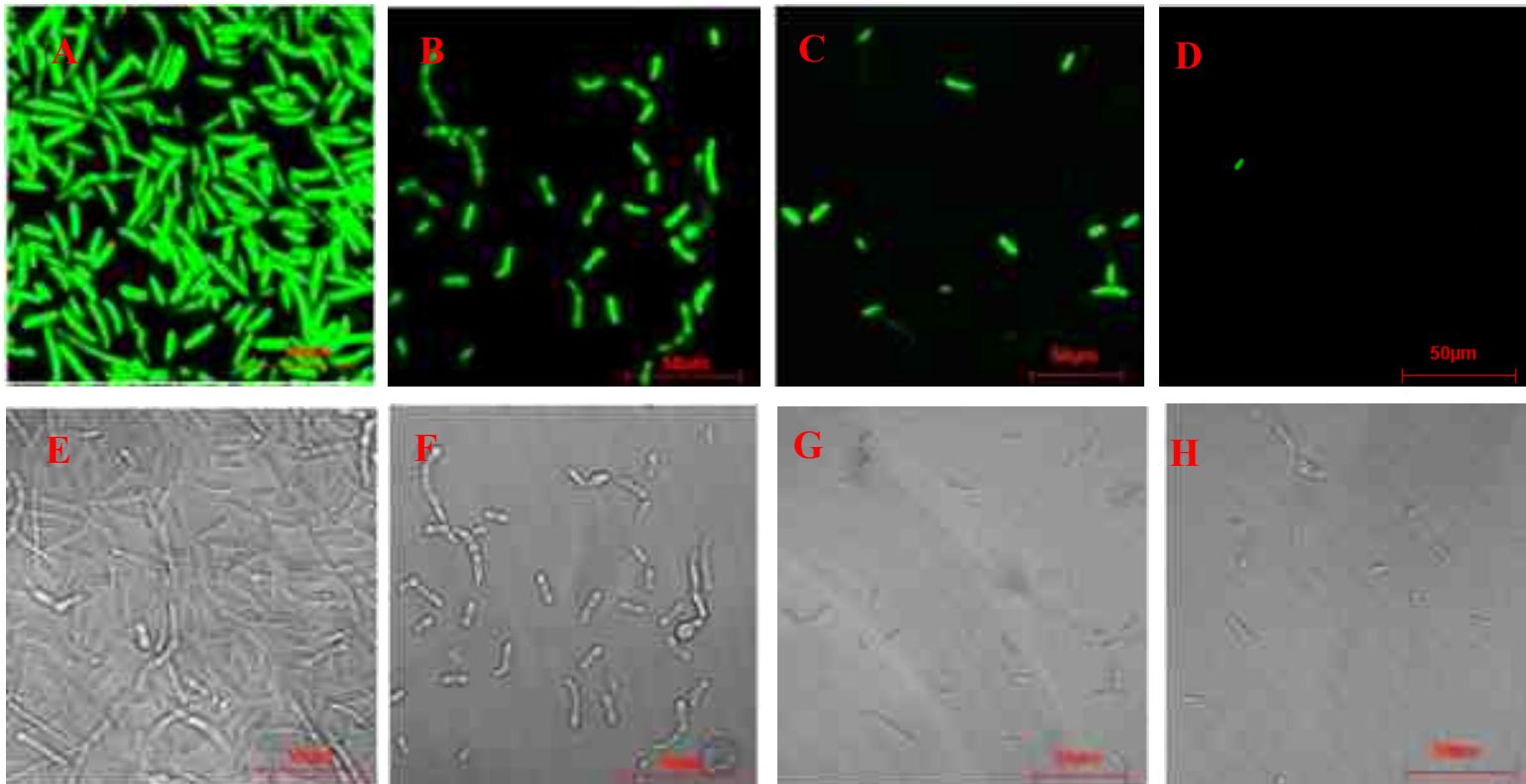


Chinese leeks have properties that destroy the cell structure of Fusarium wilt rendering the virus inert and incapable of infecting banana plants.

*Plots planted to Chinese leeks
(Photo by Yi Ganjun).*

Control of Fusarium wilt in banana with Chinese leek

Y. H. Huang • R. C. Wang • C. H. Li • C. W. Zuo •
Y. R. Wei • L. Zhang • G. J. Yi



Cultural control

Allium tuberosum vs. Foc

Some considerations for LAC



Photo by Yi Ganjun

Vs



Photo by R. Ploetz

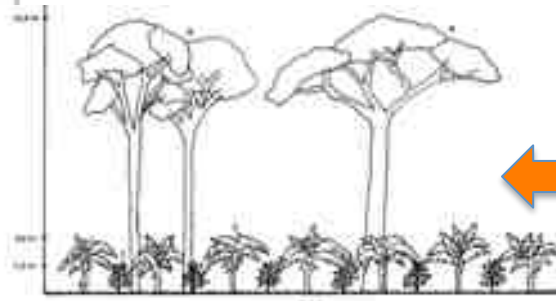
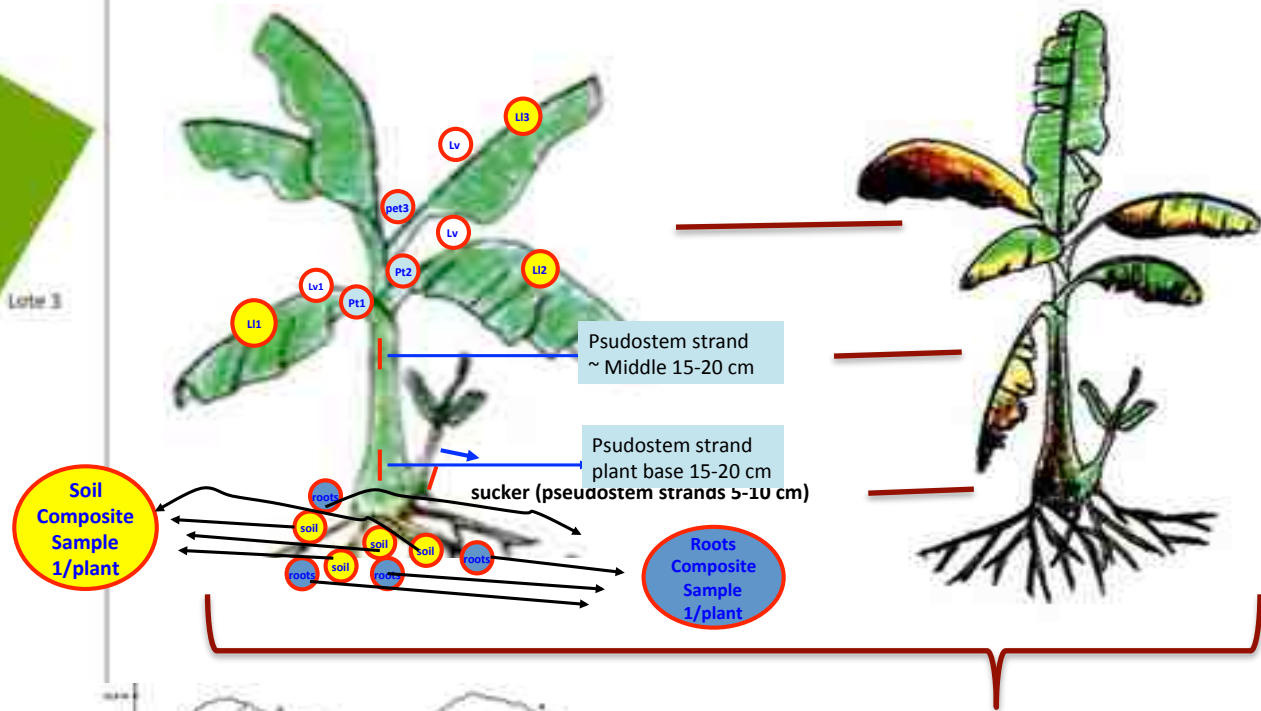


- Is this specie adapted to LAC?
- What is the practicity for bid banana plantations of monocrop – and for agroforestry systems?

Biocontrol in Foc needs to be associated to soil health and suppressive soils need a better understanding



Farm 1: Alto Varas, Turrialba, Costa Rica
Watershade, Turrialba river



Landscapes characterization

454 sequencing
Metagenomics analysis
Functional diversity

Integrated approach for disease management and enhancing productivity in Foc-susceptible bananas

**TC plants
+
target endophytes**



**Use of Beneficial
Microbial Diversity
Multifunctional isolates**

**“Biotized”/bio-fortified
healthy plants acclimatized –
Plant fitness increased!**



**Use of tailored fertilization based
on organic amendments
(Biochar, manures, compost) +
beneficial soil microbes
(Trichoderma & CIA)**

**Healthy plants are
established on a healthy soil**



**Soil -health oriented practices
and crop Integrated management**

Sustainable production of banana from High value, but Foc- susceptible varieties in Market

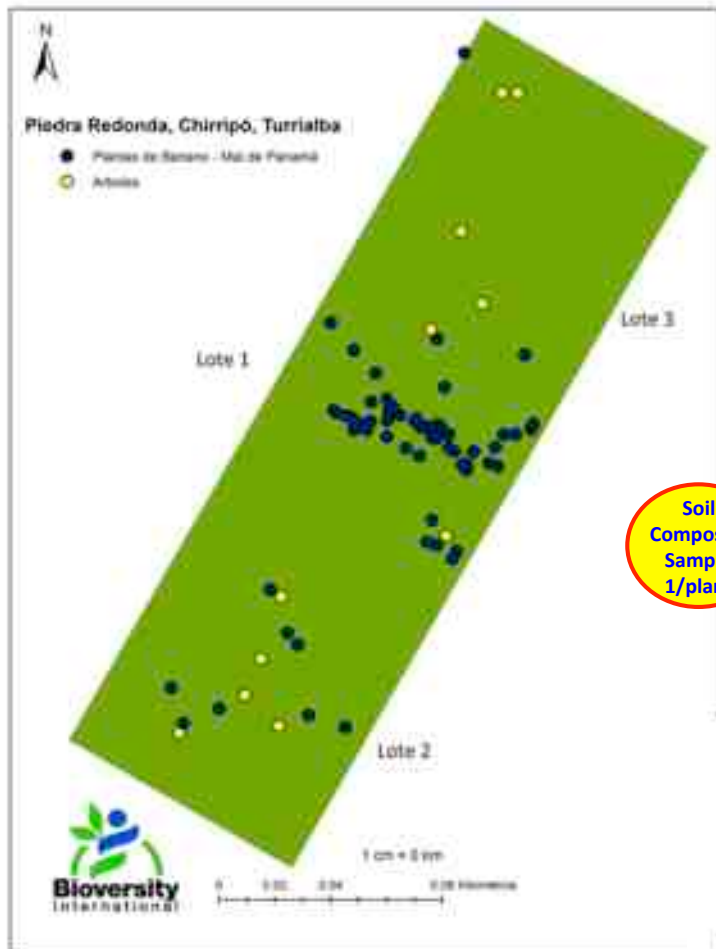
Plant health through soil health management: boosting soils for increasing plant defenses and suppressiveness to reduce losses caused by Fusarium wilt on bananas

Generating a toolbox of practices based on agrobiodiversity for sustainable management of Foc in banana

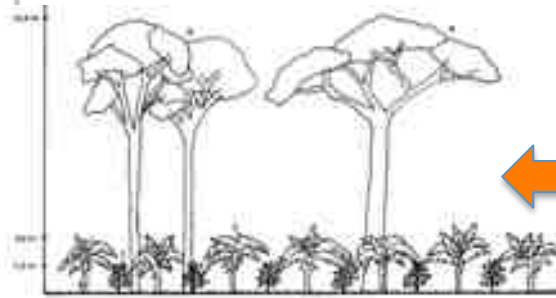
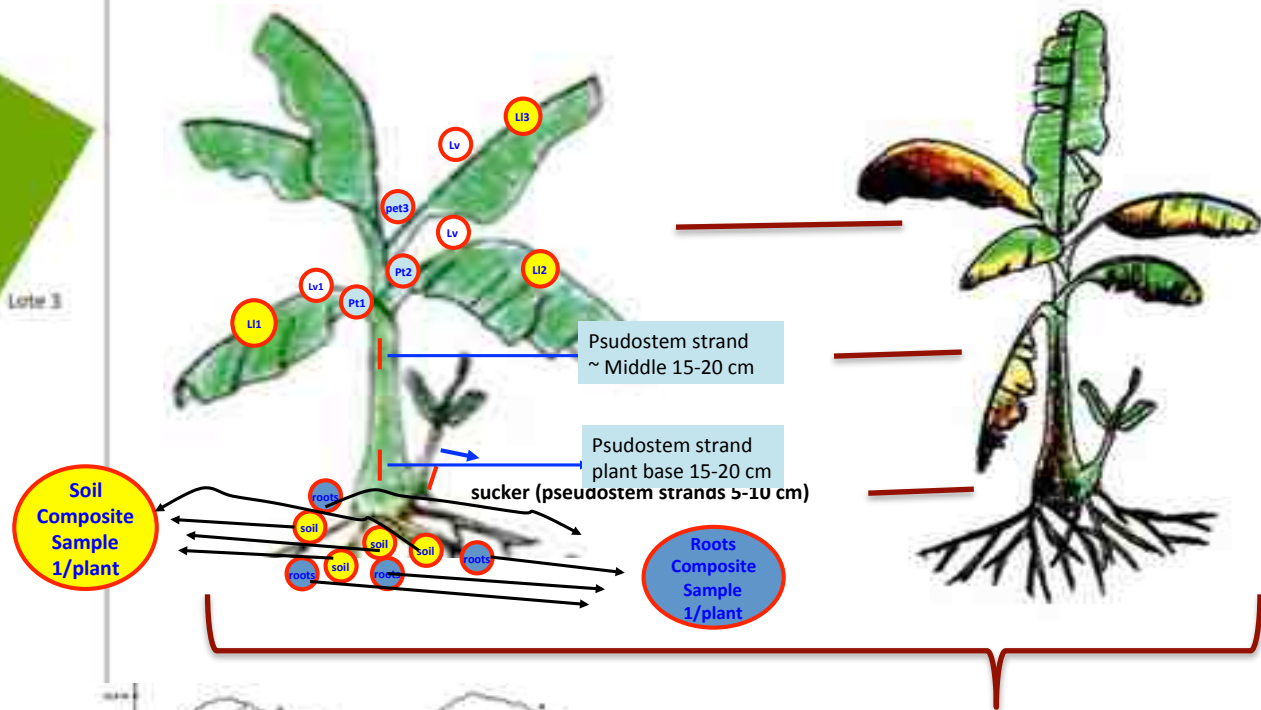


Understanding soil suppressiveness in Foc-banana

- 2 countries [Nicaragua & Costa Rica]
- 3 farms/ country, 3 areas/farm/ 5 plants/ area/



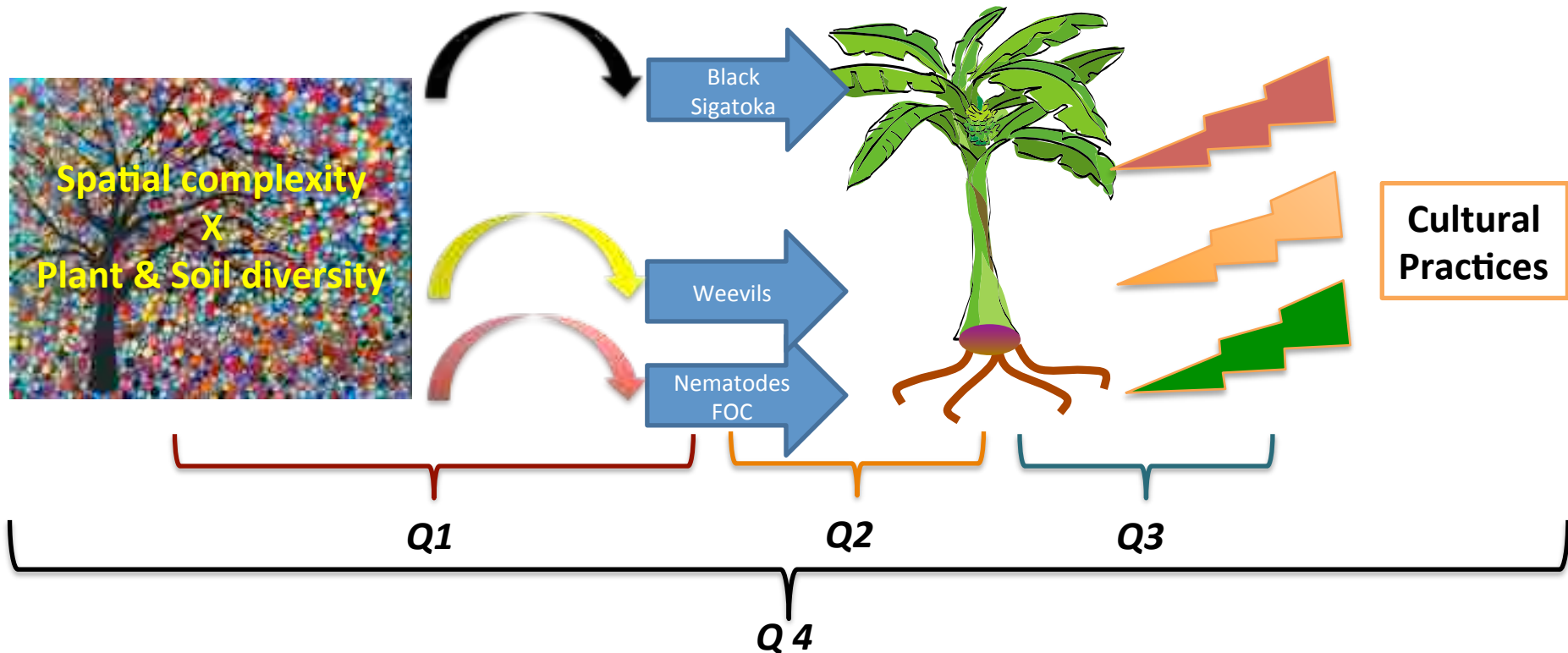
Farm 1: Alto Varas, Turrialba, Costa Rica
Watershade, Turrialba river



Landscapes characterization

454 sequencing
Metagenomics analysis
Functional diversity

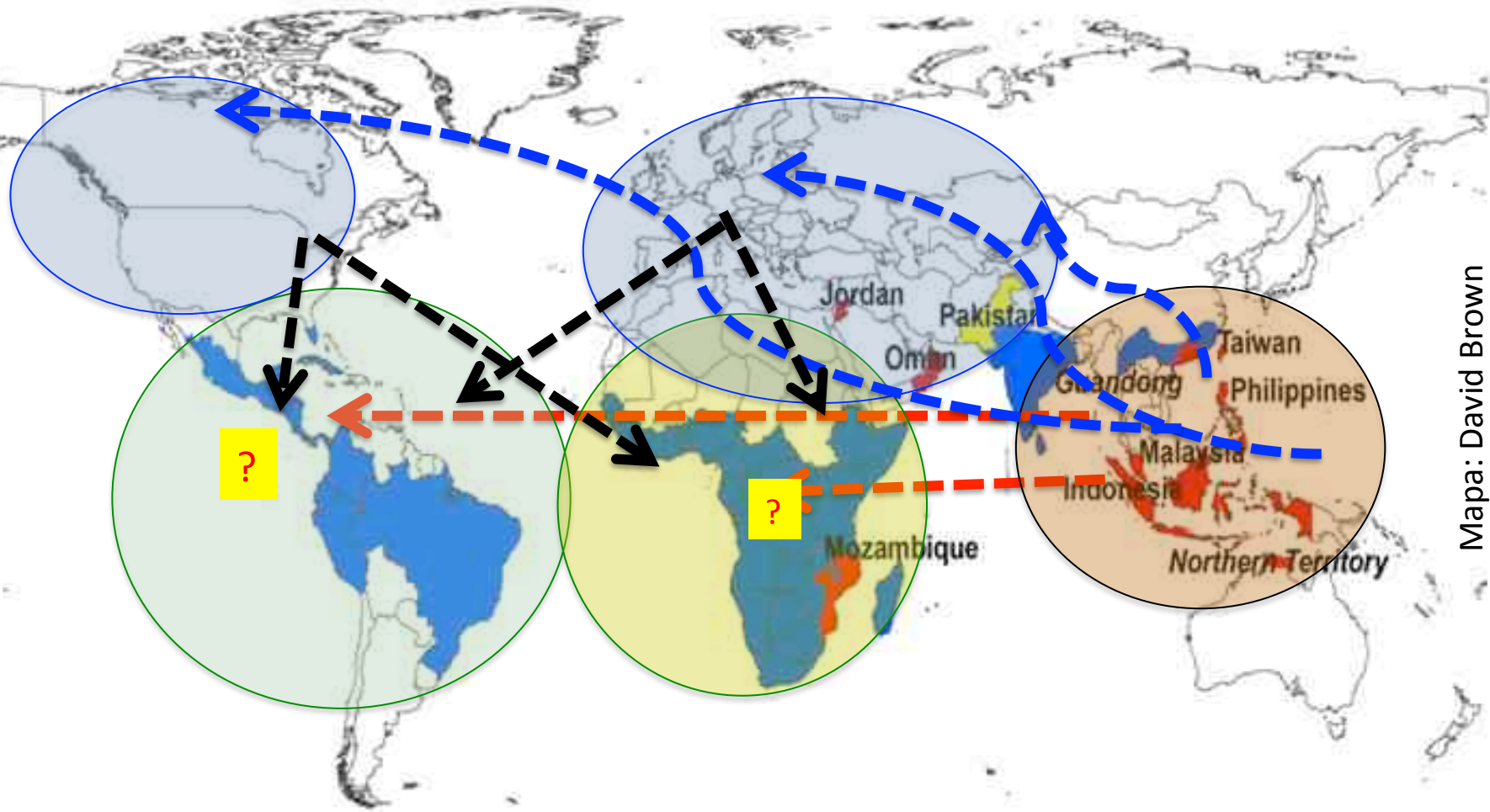
Agrobiodiversity and management factors vs. Foc & overall pests






- Q 1. How plant and soil diversity + spatial organization alter pests abundances?
- Q 2. How a profile of pests alter plant growth?
- Q 3. Which plant management practices should minimize pests effects on yield?
- Q 4. How to manage spatio-temporal factors and functional biodiversity to guarantee ecosystem services and resilience without relevant yield penalties?

Fusarium wilt of banana: Global distribution

Possible scenarios for global spreading



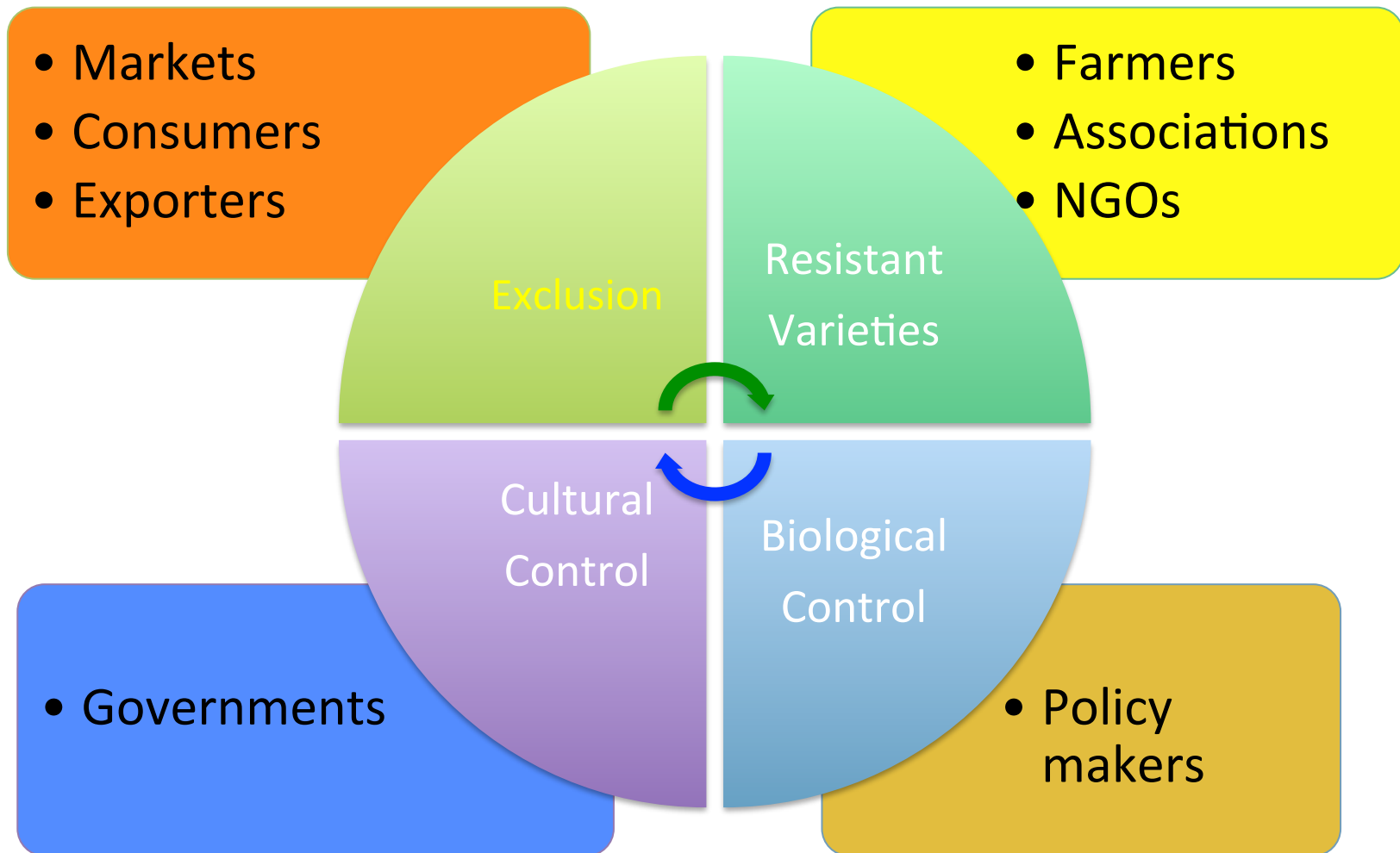
Mapa: David Brown

-  Races 1, 2
-  Tropical race 4, 1, 2
-  Under evaluation for Tropical race 4

This MAP is only a tentative approximation of the distribution of Foc to be used for academic purposes. It is not based on any scientific studies and should not be used by authorities for decision making purposes.

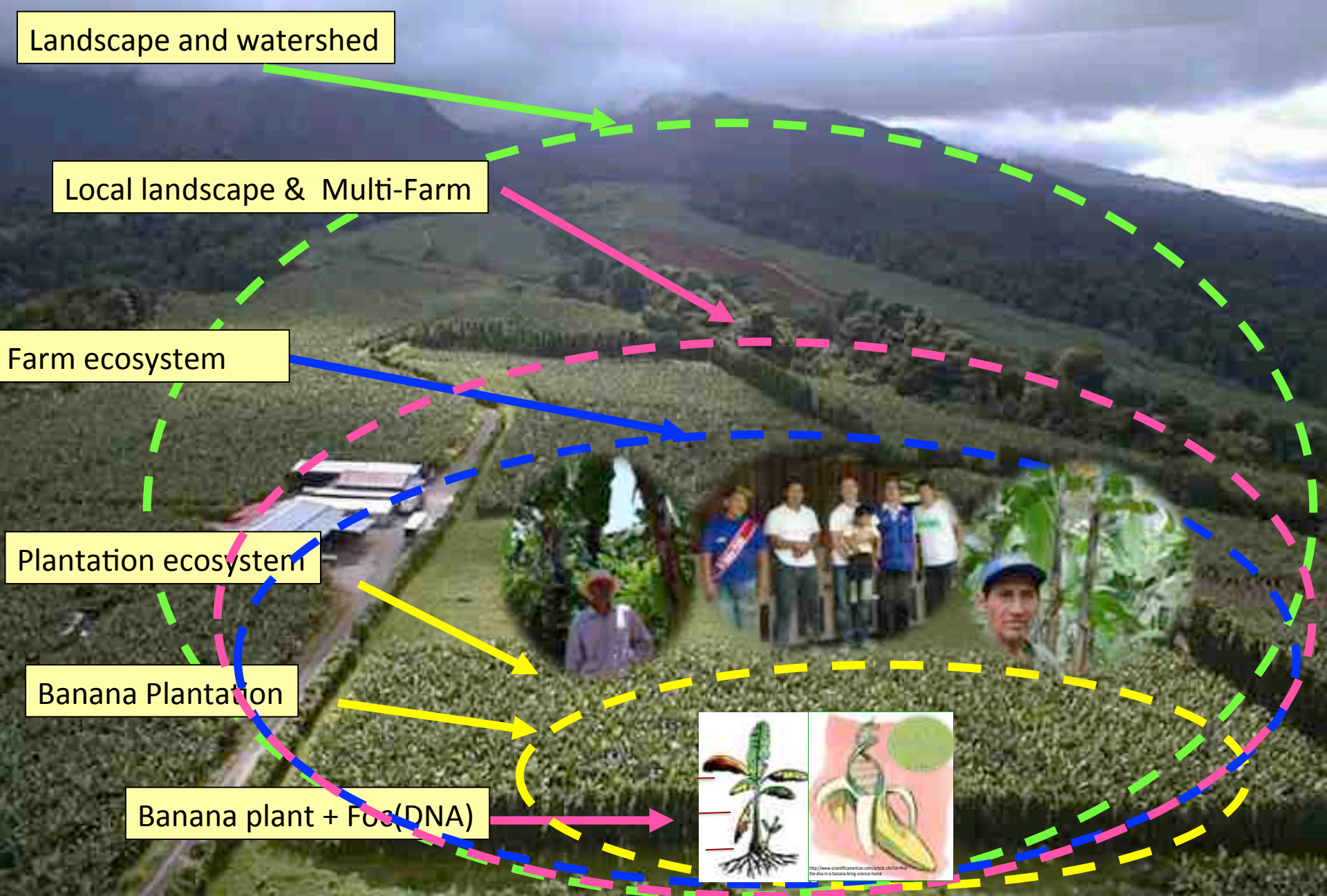
What would be the best scenario to fight against TR4 or any other high impact pest?

Integration



Take home message

Adapted from Staver 2011
Photo –ENDURE Guadalupe



Landscape and watershed

Local landscape & Multi-Farm

Farm ecosystem

Plantation ecosystem

Banana Plantation

Banana plant + Foo(DNA)

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